

# EFFECT OF COW URINE DISTILLATE ON THE GROWTH AND BIOCHEMICAL COMPOSITION OF CLADOCERA, DIAPHANOSOMA EXCISUM

# Karpagam. R<sup>1</sup>, Venkatalakshmi. S<sup>1\*</sup>

<sup>1</sup>\*Department of Zoology, Government College for Women (Autonomous), Kumbakonam, Thanjavur (Dt), Tamilnadu, India – 612001. (Affiliated to Bharathidasan University, Tiruchirapalli, Tamilnadu, India)

#### \*Corresponding author: - Karpagam. R \*Email: teajuroshan24@gmail.com

#### Abstract

Diaphanosoma excisum, the fresh water cladocera was cultured with different concentrations of cow urine distillate (CUD) to evaluate its growth performance, nutritional composition and the antibacterial potential of hemolymph. The *D.excisum* showed a dose dependent increase in growth and concentration of biomolecules with the treatment of CUD. The LC50 concentration of CUD against *D.excisum* was 0.683  $\mu$ g/ml. The hemolymph isolated from the *D.excisum* culture with CUD showed effective antibacterial activity. The study thus suggests that CUD could be utilized for the enhanced of growth and nutritional composition of *D.excisum* under optimized growth conditions and thus can be mass cultured to provide the live food in aquaculture.

Keywords: D.excisum; Zooplanktons; Cow urine distillate; Antibacterial activity; Nutritional composition.

**DOI:** 10.48047/ecb/2023.12.si10.0051

## Introduction

Zooplankton organisms have been identified as critical components of aquatic ecosystems. Through grazing, they aid in the regulation of algal and microbial productivity as well as the transfer of primary productivity to fish and other consumers (1). They help to improve water quality by grazing on phytoplankton and bacteria. As a result, zooplanktons are regarded as water quality indicators (2). Furthermore, zooplanktons provide vital live feed for fish and crustaceans, which are the primary sources of animal protein in developing countries. Thus, the development of aquaculture and inland fisheries has been phenomenal in the last two decades (3). There is a growing demand for live zooplankton of appropriate size and quality to serve as prey for crustaceans and fish larvae. Artemia (4) and rotifer species (5) are common prey, but cladocerans (6-7) and copepods are gaining popularity (8).

Imported cysts of marine Artemia were the only live feed for commercially cultured fish larvae for several decades. The high cost of Artemia has been a main constraint in the commercial production of fish larvae and fingerlings in fish hatcheries since the 1980s. This issue is particularly prevalent in developing countries intending to import the commodity (6,9-10). Scientific research was initiated to identify and culture suitable cost-effective local zooplankton to replace imported Artemia. Cladocera are a type of zooplankton found in freshwater. Daphnia is the dominant representative in the temperate region. Diaphanosoma, on the other hand, is the most diverse pelagic genus in the tropical region (11). They are an important link between primary production and higher trophic levels because they are filter feeders and a food source for planktivorous fish and other predators(11). D. excisum is a pan-tropical species that can be found in Australia, Africa, and Asia (12).

*Diaphanosoma* (13) species exhibit potential aquaculture characteristics (14). *Diaphanosoma excisum*, a common freshwater zooplankton, has significant potential as a starter diet for fish larvae (6, 9). *Diaphanosoma*, also known as 'tropical Daphnia,' is widespread in the tropics and subtropics (11, 15-16). *Diaphanosoma excisum* and *Diaphanosoma dubium* are the two most common and dominant *Diaphanosoma* species. They are parapatrically distributed in warmer waters and rarely coexist in the tropics-subtropics transition zone (17-18). *D. excisum* is frequently the only Cladoceran present in the tropics (19).

Previous research concentrated on the effect of cow urine distillate on the growth of the copepod *Mesocyclops leukarti* in the laboratory (20).

Cow urine distillate has the potential to stimulate the growth of zooplanktons. The composition of essential elements in urine influences the different components of urine (21). Among the components obtained from urine, 75-90% of nitrogen excreted is primarily in the form of urea, with the remainder released as uric acid, creatinine, and amino acid (22). Urine also contains ions such as Na<sup>+</sup>, Cu<sup>2+</sup>, Mg<sup>2+</sup>, and Cl, which are essential for plant growth (23). Because of the abundance of these substances in urine, it is critical to thoroughly review how these substances can be captured and reused. Cow urine is useful for a variety of therapeutic purposes due to the presence of various constituents. For example, Allantoin in cow urine makes it an excellent substrate for wound healing (24). Because cow urine contains copper and gold hydroxide, it acts as an antipoison and is used to eliminate the toxic effects of medicine residues (25). Furthermore, the presence of hippuric acid (a common component of urine that increases with increased consumption of phenolic compounds such as tea, fruit juices, and wine), phosphates, and uric acid makes it useful as a diuretic agent. Because of its high nitrogen content, it is beneficial in stimulating the kidney. In fact, cow urine taken orally has been shown to boost the immune system, improve memory, relieve tension, and lower cholesterol levels (24-25). In this context, our goals were to examine Diaphanosoma excisum growth capacities in monospecific cultures under optimised laboratory conditions, as well as the effect of cow urine distillate on Diaphanosoma excisum population density and nutrient composition.

# MaterialsandMethodsCollection of Plankton Samples

Plankton samples were collected from the subsurface layer of Banapureeshwar temple at a depth of 50 to 100 cm (26) and were stained with Eosin for the morphological identification with a compound microscope (27). The isolated cladocera were maintained in 500 ml of tap water contained within a beaker.

# Culture survivability

The cultivated cladocera were subjected to survival tests to determine their suitability for further research in accordance with Nandakumar, (28). 10 healthy adults of dominant cladocera species were hand-selected and placed in a 500 ml glass beaker containing tap water filtered with a 1 filter bag. They were fasted for twenty-four hours before to the trial. The survival of the cladocera was inspected and tallied daily. The studies were conducted in duplicate for a total of fourteen days. The number of surviving cladocera on the final day was counted to determine their survival (28).

#### **Pure culture preparation**

The morphological characteristics of *Diaphanosoma excisum* was recognised under a microscope (10x) using practical guide (29). Then, the cladocera species were isolated, inoculated, and fed with yeast media in a 2-liter container (28)

#### Collection cow urine and preparation of CUD

Six disease-free cows (Tag Nos. 0206, 0177, 0184, 0468, 0133, and 0201) from Goshala, Sri Vittal Rukmini Samsthan, Govindapuram, near Kumbakonam, were selected for urine collection. Six cows' first urine was collected between 4:00 and 5:00 a.m., pooled and transported to the laboratory in airtight, sterile containers (30). The obtained cow urine samples were distilled for 5 to 6 hours at 50 to 60°C using a distillation device (31). It was utilised immediately for treatment without storage.

#### D.excisum susceptibility to CUD

For the toxicity tests, only actively swimming adult animals from the stock culture were chosen. The concentrations of CUD used were 200, 400, 600, 800, and 1000  $\mu$ l/ml. Twenty animals were utilised for each of the five concentrations and the control group. Animals were divided into two groups of 10 for each concentration (in duplicate). In all tests, well-aerated tap water was used. Each test lasted 24 hours, and deaths were recorded every 6, 12, 18 and 24 hours. The LC<sub>50</sub> concentration of CUD was determined by probit analysis using IBM SPSS software.

#### Effect of CUD on D.excisum

D.excisum (500 nos) were introduced into tanks containing 20 ml water. Different of concentrations (0.01, 0.025, 0.05 and 0.1 µl/ml) of CUD based on the LC<sub>50</sub> value were added to the experimental tanks. The control tank was maintained by adding an equal volume of tap water without CUD. After 60 days, biochemical composition of CUD-treated plankton and untreated plankton were evaluated.

#### **Isolation of hemolymph**

Using a hand lens 20 mature *Diaphanosoma* excisum adults were separated and homogenized

with sterile physiological saline solution in a tissue homogenizer. The homogenate was centrifuged for 10 min at 6000 rpm. The resulting supernatant was collected that contained the hemolymph (20).

### **Biochemical analysis**

The quantitative estimation of carbohydrates was determined using the method of Roe, (32). The Lowry's method was adapted for the estimation of protein content. The total lipid was extracted following Folch et al (33) and estimated using Banes and Blackstock's (34) method. All the experiments were carried out in triplicates.

#### Antimicrobial activity of hemolymph

Using the well diffusion method, the antimicrobial activity of hemolymph of D.excisum was determined against Escherichia coli. Staphylococcus.aureus, Pseudomonas.aeruginosa and Rhodococcus rhodochrous. Sterilization of Nutrient agar media was used. After the Nutrient agar had solidified, wells were cut with a well borer. The bacterial pathogens under test were swabbed on the surface of Nutrient agar plates. 25µl of the test sample was injected into each well. For 30 min, the plates were incubated to allow the samples to diffuse into the medium. The plates were incubated at 37°C for 24 hours before measuring the diameters of the zones of inhibition in millimetres. Each antibacterial assay was carried out in triplicate, and the mean results were reported (35).

#### **Result and Discussion**

The most important factor in considering zooplanktons as live feed in aquaculture is the nutritional value of the selected zooplankton species and its sustainability for large scale synthesis (16). Cladocerans offers the mass production in a short span of time under optimized growth conditions (36-37) Previous literatures reported the use of horse manure, chicken droppings, cow dung and fish faces in the culture of cladocerans (38-39). The cladocerans holds high values in aquaculture and thus are the most used live feeds. Hence, the present study involved in the identification of cladoceran species from the fresh water Banapureeshwar temple pond and used cow urine distillate for the laboratory rearing of D.excisum for the evaluation of their nutritional dominant species of cladocera values. The identified in the fresh water pond includes D.excisum, D.brachvurum, D.sarsi and D.magna was assessed for their survivability under laboratory conditions. The survival test was

carried out for a period for 14 days and the population density of the species at the end of experimental period was determined (Fig.1). The survival of cladocera species based on population density was observed to be in the order *D.excisum* > *D.brachyurum* > *D.magna* > *D.sarsi*. Based on the survival test, the highly populated species *D.excisum* was considered for further evaluation.

To determine the toxicity of cow urine distillate as well as the tolerance of *D.excisum*, different concentrations of CUD (200, 400, 600, 800, and 1000 µl/ml) were used. The mortality of *D.excisum* was recorded after 24 h of treatment. The percentage mortality and the derived LC<sub>50</sub> value were presented in Table 1. 96.67% mortality was recorded after 24 h of treatment with 1000 µg/ml and the LC<sub>50</sub> value was 369.735 µl/ml. Hence, the CUD concentrations ranging below the LC<sub>50</sub> value was considered for further evaluation of its effects on the growth and biochemical composition of *D.excisum*.

In cladocerans, lipids and proteins are considered to be good indicators of the nutritive state (36-37). At low food concentration, lipid reserves, mainly triacylglycerides, are metabolised while proteins are only catabolised under severe starvation (38) Under laboratory conditions, the lipid content of D. magna ranges from 128 to 197 mg/g (DW) in well-fed animals and drops to about 60 mg/g (DW) in starved, hatching or senescent specimens (38). Fig.2 presents the effect of different concentrations of CUD on the The biochemical composition of *D.excisum*. zooplankton grown with CUD concentrations of 0.01µl/ml, 0.025µl/ml and 0.05µl/ml showed a carbohydrate content of 54, 67 and 60 µl/ml respectively compared to the control organism. But with increase in concentration of CUD a decrease in the carbohydrate content was observed. A dose dependent reduction in the total protein was observed compared to control. Increase in the concentration of CUD (0.1 µl/ml) significantly reduced the level of total protein (13 µl/ml). The CUD concentration of 0.05µl/ml showed an increase in the total lipid content (13.33 µl/ml) compared to control. But the total lipid was found to be decreased with increase in the concentration of CUD. The low content of protein and lipids observed in this study in D.excisum might be attributed to the age and feed received by the zooplankton species during their growth period (40). Farkas et al., (41) reported a protein content of 223 mg/g and 346 mg/g of lipids in cladocera. Cauchie et al., (42) reported

that the protein and lipid content of cladocera ranged from 186 to 397 mg/g and 65 to 171 mg/g respectively. The protein content in Daphnia species was found to be 62.6% and *D.excisum* was reported to contain 57.3% of protein. The study by Praveena et al., (2020) showed a low protein content of 96 µg/dl in Mesocyclops leukarti. From the earlier reports it was generally found that the cultured organism had low content of biomolecules compared to wild type (42). Praveena et al., (20) reported an optimal concentration of CUD (0.5%)effectively enhanced the growth of copepod culture. Mesocyclops leukarti. The present study witnessed optimal concentration of an 0.025 µl/ml concentration of CUD had a effective role in enhancing the biochemical composition of D.excisum hemolymph.

The hemolymph of D.excisum cultured with CUD was estimated for its antibacterial activity against the gram positive and gram negative bacterial pathogens such as R.rhodochrous S.aureus, E.coli and P.aeruginosa. Table 2 and Fig.3 shows the inhibitory activity of D.excisum against pathogenic bacterial species. From the results obtained it could be concluded that gram positive bacterial species were more susceptible compared to gram negative species. Maximum zone of inhibition was observed with S.aureus (23.9±0.7) and minimum inhibitory activity was recorded against the gram negative strains  $(18.5\pm0.9)$ without any significant difference in activity among the two negative strains examined in this study. In a previous study, utilizing the hemolymph of *D.excisum*, the negative bacterial species including E.coli and K.pneumoniae were reported to be effectively inhibited compared to gram positive strains. This antagonist finding with the present study might be attributed to difference in species response towards bacterial pathogens (20).

#### Conclusion

Increasing global population and the subsequent demand for animal proteins are majorly compensated by the rapid development of aquaculture. Mass production of zooplanktons as live food in aquaculture thus becomes inevitable. The present study signifies the use of cow urine distillate as a valuable resource in the enhanced production of zooplankton, particularly cladocera, *Diaphanosoma excisum*. The species showed sustainable growth with increased biochemical composition with cow distillate under optimized conditions. Hence, the study recommends the use of cow urine distillate in the culture of *D.excisum* to be used as live feed in aquaculture.

#### Reference

- 1. Dejen, E., Vijverberg, J., Nagelkerke, L. A., & Sibbing, F. A. (2004). Temporal and spatial distribution of microcrustacean zooplankton in relation to turbidity and other environmental factors in a large tropical lake (L. Tana, Ethiopia). *Hydrobiologia*, *513*, 39-49.
- Pinto-Coelho, R., Pinel-Alloul, B., Méthot, G., & Havens, K. E. (2005). Crustacean zooplankton in lakes and reservoirs of temperate and tropical regions: variation with trophic status. *Canadian Journal of Fisheries* and Aquatic Sciences, 62(2), 348-361.
- 3. Petr, T. (1994). Intensification of reservoir fisheries in tropical and subtropical countries. *Internationale Revue der gesamten Hydrobiologie und Hydrographie*, 79(1), 131-138.
- 4. Versichele, D., LCger, P., Lavens, P. and Sorgeloos, P., 1989. The Utilization of Anemia. Technique et Documentation. Lavoisier, Paris, France, pp. 241-260.
- 5. Pourriot, R., & Francez, A. J. (1986). *Rotifères* (Vol. 8). Association franç aise de Limnologie.
- Adeyemo, A. A., Oladosu, G. A., & Ayinla, A. O. (1994). Growth and survival of fry of African catfish species, Clarias gariepinus Burchell, Heterobranchus bidorsalis Geoffery and Heteroclarias reared on Moina dubia in comparison with other first feed sources. *Aquaculture*, 119(1), 41-45.
- Ganzon-Naret, E. S. (2014). Utilization of Moringa oleifera leaf meals as plant protein sources at different inclusion levels in fish meal based diets fed to Lates calcarifer. *Animal Biology & Animal Husbandry*, 6(2), 158-167.
- 8. Støttrup, J. G., & Norsker, N. H. (1997). Production and use of copepods in marine fish larviculture. *Aquaculture*, *155*(1-4), 231-247.
- 9. Ovie, S. I., Adeniji, H. A., & Olowe, D. I. (1993). Isolation and growth of curve characteristics of a freshwater zooplankton for feeding early larvae and fry stages of fish. *Journal of Aqua. Tropical*, *8*, 181-196.
- 10.Ovie, S. I., & Egborge, A. M. (2002). The effect of different algal densities of Scenedesmus acuminatus on the population growth of Moina micrura Kurz (Crustacea: Anomopoda, Moinidae). *Hydrobiologia*, 477, 41-45.
- 11.Sarma, S. S. S., Nandini, S., & Gulati, R. D. (2005). Life history strategies of cladocerans:

comparisons of tropical and temperate taxa. *Hydrobiologia*, 542, 315-333.

- 12.Korovchinsky, N. M. (2013). Cladocera (Crustacea: Branchiopoda) of South East Asia: history of exploration, taxon richness and notes on zoogeography. *Journal of Limnology*, 72.
- 13.Sripayatt,P.,1981.Diaphanosomaculture.ThaiFi sh.Gaz. 34,617–621.
- 14. Mavuti, K. M. (1994). Durations of development and production estimates by two crustacean zooplankton species Thermocyclops oblongatus Sars (Copepoda) and Diaphanosoma excisum Sars (Cladocera), in Lake Naivasha, Kenya. *Hydrobiologia*, 272, 185-200.
- 15. Dumont, H. J. (2014). On the diversity of the Cladocera in the tropics. *Studies on the Ecology of Tropical Zooplankton*, 27-38
- 16. Pan, J., Liu, P., Pajk, F., Dumont, H. J., & Han,
  B. P. (2021). The mitochondrial genome of Diaphanosoma excisum Sars, 1885 (Crustacea: Branchiopoda: Cladocera) from Hainan Island, China. *Mitochondrial DNA Part B*, 6(3), 1279-1280.
- 17. Korovchinsky, N. M., Walsh, E. J., & Smolak,
  R. (2017). Diaphanosoma Fischer, 1850 (Crustacea: Cladocera: Sididae) of Lake Turkana (East Africa), with the description of a new species of the genus. *Zootaxa*, 4250(1),77-89.
- 18. Pajk, F., Zhang, J., Han, B. P., & Dumont, H. J. (2018). Thermal reaction norms of a subtropical and а tropical species of their Diaphanosoma (Cladocera) explain distribution. Limnology and Oceanography, 63(3), 1204-1220.
- 19. Kotov, A., Forró, L., Korovchinsky, N. M., & Petrusek, A. (2013). World checklist of freshwater Cladocera species. World Wide Web electronic publication.
- 20.Praveena, V., Venkatalakshmi, S., Alharbi, N. S., Kadaikunnan, S., Khaled, J. M., & Govindarajan, M. (2020). Identification of a novel antibacterial protein from hemolymph of freshwater zooplankton Mesocyclops leuckarti. *Saudi Journal of Biological Sciences*, 27(9), 2390-2397.
- 21.Randall, D. G., & Naidoo, V. (2018). Urine: The liquid gold of wastewater. *Journal of Environmental Chemical Engineering*, 6(2), 2627-2635.
- 22. Maurer, M., Pronk, W., & Larsen, T. A. (2006). Treatment processes for source-separated urine. *Water research*, 40(17), 3151-3166.

- 23. Viskari, E. L., Grobler, G., Karimäki, K., Gorbatova, A., Vilpas, R., & Lehtoranta, S. (2018). Nitrogen recovery with source separation of human urine—preliminary results of its fertiliser potential and use in agriculture. *Frontiers in Sustainable Food Systems*, 2, 32.
- 24. Mohanty, I., Senapati, M. R., Jena, D., & Palai, S. (2014). Diversified uses of cow urine. *Int J Pharm Pharm Sci*, 6(3), 20-2. Randhawa, G. (2010). Cow urine distillate as bioenhancer. *Journal of Ayurveda and integrative medicine*, 1(4), 240.
- 25. Randhawa, G. (2010). Cow urine distillate as bioenhancer. *Journal of Ayurveda and integrative medicine*, 1(4), 240.
- 26.Davis, C. C. (1955). Plankton and industrial pollution in Cleveland 6arbour. *Sewage and Industrial Wastes*, 835-850.
- 27. Manickam, N., Bhavan, P. S., Santhanam, P., Muralisankar. Т., Srinivasan, V., Radhakrishnan, S., ... & Ali, A. J. (2014). Seasonal variations of zooplankton diversity in reservoir perennial at Thoppaiyar, Dharmapuri District, South India. Austin Journal of Aquaculture and Marine Biology, 1(1), 1-7.
- 28.Nandakumar, R., Prasath, B. B., Santhanam, P., Ananth, S., Jayalakshmi, T., Kumar, S. D., & Devi, A. S. (2015). Optimization of culture conditions for marine copepod Macrosetella gracilis (Dana, 1847) with emphasis on salinity and algal diets.
- 29.Lynne M. Witty, (2004). Practical Guide to Identifying Freshwater Crustacean Zooplankton. Cooperative Freshwater Ecology Unit, 2nd edition.
- 30.Sattanathan, G., & Venkatlakshmi, S. (2015). Efficacy of different breeds of cow urine distillate on growth and food utilization Of Indian Major Carp, Labeo Rohita (Hamilton) Fingerlings. *History*, 14(46), 169-185.
- 31. Arunkumar Sathasivam, M. Muthuselvam and Rajasekaran Rajendran, 2010. Antimicrobial Activities of Cow Urine Distillate Against Some Clinical Pathogens, Global Journal of Pharmacology4 (1): 41-44.
- 32. Roe, J. H. (1955). The determination of sugar in blood and spinal fluid with anthrone reagent. *Journal of Biological chemistry*, 212, 335-343.

- 33.Folch, J., Lees, M., and Sloane-Stanley, G.H. (1957) A Simple Method for the Isolation and Purification of Total Lipides from Animal Tissues, J. Biol. Chem. 226, 497–509.
- 34.Barnes, H., & Blackstock, J. (1973). Estimation of lipids in marine animals and tissues: detailed investigation of the sulphophosphovanilun method for 'total'lipids. *Journal of experimental marine biology and ecology*, *12*(1), 103-118.
- 35.Ruangpan, L., Tendencia. & E. (2004). Laboratory manual of standardized methods for antimicrobial sensitivity tests for bacteria isolated from aquatic animals and environment. Aquaculture Department, Southeast Fisheries Development Asian Center..
- 36. Guisande, C., & Gliwicz, Z. M. (1992). Egg size and clutch size in two Daphnia species grown at different food levels. *Journal of Plankton Research*, *14*(7), 997-1007.
- 37.Tessier AJ, Goulden CE (1982) Estimating food limitation in cladoceran populations. Limnol Oceanogr 27:707-727
- 38.Elendt, B. P. (1989). Effects of starvation on growth, reproduction, survival and biochemical composition of Daphnia magna. *Archiv für Hydrobiologie*, 415-433.
- 39.Safi, V., Darshan, A., Gogoi, B., Kumar, R., Saikia, R., & Das, D. N. (2016). Effect of different levels of poultry droppings on growth performance of Indian major carps in the foothills of Arunachal Pradesh, India. *International Journal of Fisheries and Aquatic Studies*, 4(2), 56-63.
- 40.Budhin Gogoi, Vivekanand Safi and Debangshu Narayan Das (2016). The cladoceran as live feed for fish culture. A brief Animal. review. Research Journal of Veterinary and Fishery Sciences, 4(3), 7-12.
- 41. Farkas, G. L., & Kiraly, Z. (1958). Enzymological aspects of plant diseases. I. Oxidative enzymes. *Phytopath. Z.*, *31*, 251-272.
- 42. Cauchie, H. M., Hoffmann, L. U. C. I. E. N., & Thome, J. P. (1999). Ingestion rates of Daphnia magna Straus (Crustacea: Branchiopoda: Anomopoda) on bacterioplankton and phytoplankton in an aerated waste stabilisation pond. *Belgian journal of zoology*, *129*(1), 285-303.



Fig. 1 . Survivability of different Cladocera species in culture



Hours_	Percentage Mortality/ CUD (µl/ml)					LC 50 (LCL-	LC 90 (LCL-	Regression
	200	400	600	800	1000	UCL)	UCL)	Equation
6	10.00±10.00	26.67±5.77	36.67±5.77	50.00±20.20	70.00±10.00	598.494 (501.584- 722.841)	1614.154 (1189.944- 2898.294)	Y= 6.667X+.087
12	20.00±10.00	40.00±10.00	50.00±10.00	60.00±10.00	76.67±5.77	517.515 (397.860- 663.346)	1867.949 (1222.073- 5053.396)	Y= 5.667X+.075
18	23.33±5.77	46.67±5.77	56.67±5.77	66.67±5.77	86.67±5.77	482.741 (380.860- 594.812)	2175.428 (1425.142- 3345.285)	Y= 14.667X+.063
24	36.67±5.77	56.33±5.77	56.67±5.77	73.33±5.77	96.67±5.77	369.735 (235.134- 483.094)	2465.242 (1416.634- 10724.634)	Y= 26.667X+.053





Table 2 Antibacterial activity of D.excisum hemolymph

SUNA	Destario	Zone of Inhibition (mm)				
51.INU	Dacteria	Antibiotic	Hemolymph			
1	R.rhodochrous	20.6±0.5	14.5±0.04			
2	S.aureus	24.0±0.3	23.9±0.7			
3	E.coli	21.3±0.6	20.6±0.5			
4	P.aeruginosa	20.4±0.4	18.5±0.4			

#### Fig. 3 Antibacterial activity of D.excisum hemolymph

