



# **Insight into the recent progress of Ag/Au-based nanomaterials-synthesis, characterising tools and bio-applications**

Divya Arulraj<sup>1</sup>, Tapan Kumar Mistri<sup>1#</sup>

<sup>1</sup>SRM Institute of Science and Technology, Kattankulathur Campus, SRM Nagar, Potheri, Chennai-603203, India

**Running head:** Insight into the recent progress of Ag/Au-based nanomaterials

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**Address correspondence to:**

\*Tapan Kumar Mistri, Department of Chemistry, SRM Institute of Science and Technology, Kattankulathur, SRM Nagar, Chennai-603203, Tamil Nadu, India,  
Email: [tapankum@srmist.edu.in](mailto:tapankum@srmist.edu.in)

## **Abstract:**

Nanomedicine, known as a nanotechnology-based biological application, is concerned with synthesizing, characterization, and functionalizing unique the for targeted drug delivery from the pharmaceutical industry. Silver (Ag) and gold (Au) nanoparticles (NPs) are important due to the unique properties that make them useful in various fields. Ag-NPs are known for their excellent antimicrobial properties, while Au-NPs have unique electronic and optical properties that make them useful in catalysis, electronics, imaging, and sensing. Ag and Au-NPs have potential applications in nanomedicine, including drug delivery, cancer therapy, and imaging. This review covers the synthesis of Ag and Au-NPs using chemical, physical, and biological methods. Biosynthesis methods are preferred over chemical and physical methods. Characterization of the nanoparticles was done using various techniques, including X-ray diffraction, UV-VIS, FTIR, DLS, XPS, SEM, and TEM analysis. The review also discusses the nanoparticles' potential applications, including antimicrobial activity, regenerative medicine, and cell therapy. In the future, Ag and Au-NPs will have potential applications in medical treatments, environmental remediation, energy production, electronics, and advanced materials development. Ongoing research into their properties and capabilities could lead to significant advances in these fields.

## **1. Introduction:**

The development of nanotechnology has been accelerated by the introduction of nanomaterials, specifically inorganic nanoparticles (NPs) and nanorods, which possess unique properties and physicochemical characteristics that are distinct from their bulk counterparts and are size-dependent. Current nanotechnology focuses on nanostructures that are smaller than 100 nanometers in at least one dimension and that can be altered at the atomic or molecular level. Early symptom diagnosis and treatment in clinical and environmental research have huge potential thanks to the interdisciplinary study field that combines chemistry, biology, engineering and medicine.[1,2].The term “nano” is used to describe objects or structures that are extremely small, with dimensions measured in nanometers, which is one billionth of a meter[3].Metallic materials have long attracted researchers at the nanoscale level (metallic nanoparticles). They are now using them in a wide range of applications in many different industries, including engineering, agriculture, medicine, etc[4,5].When compared to macromolecules, nanoparticles (0.1-100 nm) have distinct physical and chemical properties. A recent development in nanotechnology has a wide range of medical uses. Researchers have been interested in the environmentally safe and nontoxic process of biological nanoparticle manufacturing. Due to their greater surface area, nanoparticles offer stronger interactions with microbial agents. Moreover, nanoparticles have a greater ability than conventional macro- and microparticles to enter bacterial cells and display bactericidal activity[6].These benefits make nanoparticle systems potential medicinal agents.

Both organic and inorganic functional groups have a significant impact on the surface modifications that occur on the atoms of desired metals or metal oxides.. This method not only prevents oxidation and agglomeration of nanoparticles but also enables improved functionalization[7].By connecting with the surfaces of other molecules, solids, or nanoparticles, simple organic molecules or groups provide nanoparticles with sufficient potential protection against agglomeration [8].This will enable NPs to approach a desired biological organism. If they are coated with biological molecules, they might be able to adhere to a cellular division's surface and form a link with it, offering a precise means of tracking or finding it.[9].In the biological sciences, using nanoparticles (NPs), primarily metals NPs, for therapeutic and diagnostic reasons has recently increased. The increase of NPs' application and purposes can be

attributed to factors such as their distinctly small size, tremendous strong responsiveness to living tissues, high temperature stability, and cellular mobility are all characteristics of this material.[10].The size, shape, and morphology of nanoparticles determine their biological characteristics, and these factors may be affected by the synthesis process[11].It is possible to create nanoparticles through chemical, physical, and biological processes[12].Evaporation-condensation and laser ablation are examples of physical processes, while chemical approaches call for reducing and stabilizing agents[13].On the other hand, biogenic synthesis uses algae, fungi, plants, and microbes as reducing agents and is, therefore, more environmentally benign[14].

Due to their environmental friendliness, low cost, and very simple manufacture, syntheses of nanoparticles, especially Ag and Au, have attracted significant interest[15].Nanoparticles made of metal and metal oxide have several uses in the study of biomedicine and pharmaceuticals.Ag-NPshave been widely used in therapies, particularly in the practice of customized healthcare. Ag-NPs are frequently employed in tissue regeneration, medication administration, antibacterial therapy, and diagnostic procedures.Au-NPs have been extensively utilised in a variety of biomedical activities, including cancer treatment and diagnosis, targeted medication distribution, and bioimaging[16]. They are simple to produce and can be coupled with a variety of molecules and chemicals. Compared to other metal particles, such as Ag-NPs, they have better rates of penetration and are significantly less harmful. Au-NPsare the ideal material of choice for biomedicine because of their characteristics[17].Additionally, Ag-NPs have been successfully applied as antibacterial agents in a variety of ecological applications, fabric coatings, storage of food, and biopharmaceutical sectors.[18].Ag has, in fact, been used successfully to both prevent and treat infections brought on by a number of different microbes[19].Ag-NP-based conjugates with doxorubicin, fluorouracil, peptides, and folic acid, among other chemotherapeutic anticancer medicines, have been created so far as anticancer agents and for various cancer-related uses[20].It is obvious that Ag-NPs and their conjugates have a lot of promise for use in a variety of scientific and technological industries, especially in the medicinal and medicinal fields[21].From diagnosis to treatment, including biosensors, Au-NPsin a variety of shapes like nanostar, nanorod, nanocage, and nanosphere, have been investigated in a variety of biomedical applicationsincluding the delivery of drugs and genes[22-23], phototherapyalong with overheating[24], and antimicrobial applications[25-27].According to a previous study, Au-NPs

between 10 and 30 nm in size implanted more readily into malignant tumor cells than bigger ones[28]. Surprisingly, smaller diameters of 20 nm, as opposed to 40 and 80 nm, of surface-coated Au-NPs demonstrated a greater cell uptake[29].

This review concentrated on identifying recent exposure to Ag/Au-NPs relevant synthesis, characterization, and bio-applications. The biomedical and healthcare implications of Ag-NPs have mostly focused on microbiological, fungus, antitumor, anti-oxidative, antiviral, and antihyperlipidemic applications. On the other hand, the key focus of Au-NPs was given towards biomedical applications such as regenerative medicine, cell therapy and antimicrobial activity. This review also provides an overview of recent developments in the design of hybrid nanomaterials and discusses how they are mostly used in clinical and translational research. Furthermore, the review gave insight into the future direction of the research field relevant to the bio-application of Ag/Au-NPs in regenerative medicine.

## **2. Synthesis of Ag and Au-NPs**

Researchers have been interested in Ag and Au-NPs (NPs) because of their distinctive features (for example size and form can vary based on optical, antibacterial, and electronic qualities). For the synthesis of Ag-NPs, many various synthesis techniques have been published [30]. Nanoparticle synthesis can be done physically, chemically, photochemically, or biologically. Furthermore, condensing and dispersing are the two categories under which colloidal nanoparticle solutions are prepared. Additionally, Laser ablation, cathode sputtering, and electric arc dispersion are examples of "top-down" dispersion techniques that work by destroying the material's crystal lattice. The chemical process is the basis for condensation techniques (reduction in solution, followed by nanoparticle precipitation, formation and stabilization)[31] **(Figure 1)**.

The most used processes for creating nanoparticles are chemical ones. Certain chemical techniques, however, are unable to completely eliminate the synthesis protocol's use of harmful substances[32-33]. Since noble metal nanoparticles like Au and Ag are frequently used in regions where people come into touch with them, there is an increasing need to create ecologically acceptable nanoparticle synthesis procedures that don't include hazardous chemicals. As potential environmentally benign biological techniques for creating nanoparticles utilising

microbes are alternatives to chemical and physical processes[34-36], proteins [37], and plant extract or plants [38] have been proposed. This has raised awareness of biogenic synthesis techniques. These techniques make it is feasible to make NPs that have better physiochemical properties, more consistency, and lower toxicity[39].

## **2.1 Preparation of Ag-NPs**

Firstly, the review would discuss the key synthesis approaches of the Ag-NPs. In the chemical reduction method for synthesising Ag-NPs, initial concentrations of AgNO<sub>3</sub>, reducing agent, AgNO<sub>3</sub> molar ratios, and stabiliser concentrations all have an impact on several metrics, including the particle size and aggregation state of Ag-NPs[40]. Various reducing agents along with suitable stabilizing agent are used for chemical reduction method such as Hydrazine hydrate and Sodium Dodecyl Sulphate (SDS); Citrate of sodium (performs both reducing and stabilising functions)[41]; ascorbic acid (reducing agent) and Cetyltrimethylammonium bromide (CTAB, stabilizing agent)[42]; Trisodium citrate (reducing agent)[43-45]; and Sodium borohydrate (reducing agent)[46-48]. Poly vinyl alcohol, Sodium citrate and Hydrogen peroxide (performs both reducing and stabilising functions)[49,50]; PVP, Ethylene glycol solution (performs both reducing and stabilising functions)[51]; Threonine (reducing agent), PVP and NaOH (stabilizing agent) [52]; L-alanine and ascorbic acid (reducing agent) [53]. There are numerous ways that have been established for the chemical preparation of Ag-NPs, including the chemical reduction approach [54], the polyol method [55], and the radiolytic process [56]. Chemical reduction is the best and easiest way to produce nanoparticles that don't aggregate, have a high yield, and require little to no pre-treatment [57].

## **2.2 Preparation of Au-NPs**

Synthesis of Au-NPs using various types of methods are discussed here. Au-NPs are also synthesised similar way to that of Ag-NPs with the help of reducing and stabilizing agents. Primarily the review found out that the chemical synthesis method such as chemical method is the key synthesis strategy for having bio-graded NPs where both reduction (using substance such as borohydrides, amino boranes and one electronic reducing substances such as electron-rich transition metal sandwich complexes) and followed by stabilization (using substances such as trisodium citrate dihydrate) are performed subsequently[58]. Turkevich

method, for reducing  $\text{HAuCl}_4$  with citrate in aqueous, is one of the most well-known methods for producing Au-NPs [59]. During time, the solution progressively changes from bright yellow to wine red. By using this technique, Au-NPs with a diameter of roughly 20 nm are produced. Citrate ions act as both stabilizing and reducing agents [59,60], Brust-Schiffrin method, this technique made it simple to create Au-NPs with regulated size and low dispersion that are thermally and air-stable. In this method,  $\text{AuCl}_4$  was present in dodecanethiol and the addition of the reducing agent causes the transition of the organic phase from orange to dark brown [61-64]. The electrochemical method can be used to create Multiwalled carbon nanotube surfaces with glassy carbon electrodes are coated in Au-NPs. By electrochemically oxidizing the anode and reducing the cathode in a straightforward two electrode cell, Au-NPs were produced [65-67]. In seeding growth method, the buried growth method was used to provide restricted size dispersion. Controlling the particle size of gold nanoparticles is achievable by adjusting the seed to metal salt ratio, which in turn enables the generation of different levels within the 540 nm range. This approach has the benefit of being a simple, efficient, and affordable procedure [68-72]. Furthermore, surfactant-assisted method had been employed to create partially functionalized Au-NPs. Various surfactants, including oleylamine, poly (N vinylpyrrolidone), and  $\text{AuCl}_4$  in 1,5-pentanediol, were used in the hot injection approach to create Au-NPs and stabilize the colloidal solution. By reducing Au salt chemically in an organic solvent while also adding a stabilizing agent, Au-NPs were created using this process [73-75].

Moreover, leaf broth was used to explain the creation of pure metallic nanoparticles of Ag and Au by reduction of  $\text{Ag}^+$  and  $\text{Au}^{+3}$  ions. [76,77]. As potential environmentally acceptable alternatives to physiochemical approaches, biological methods of nanoparticle synthesis using microorganisms, enzymes, and plants or plant extracts have been proposed [78]. By avoiding the complex process of maintaining cell cultures, using plants to synthesise nanoparticles has an advantage over other biological techniques. Significantly, this method has potential to be appropriately scaled up for the large-scale synthesis of nanoparticles. Another benefit of utilising plant extracts instead of bacteria to create nanoparticles is that they are more stable over a longer period of time [79]. Additionally, fungi are able to manufacture more nanoparticles than bacteria because they can secrete enormous amounts of proteins, which directly help to transform into better nanoparticle productivity [80]. Among chemical, physical, and biological methods of both Ag and Au-NPs synthesis, the biosynthesis method produces more NPs' yield compared to the

chemical as well as physical methods. Current research has shown that biologically produced nanoparticles are more pharmacologically active than nanoparticles made by physicochemical synthesis [81].

### **3. Characterization of Ag and Au nanoparticles using various analytical techniques:**

The prepared Ag- and Au-NPs are typically identified by a variety of analytical approaches, including scanning electron microscopy (SEM), transmission electron microscopy (TEM), atomic force microscopy (AFM), extended X-ray absorption, energy-dispersive spectroscopy (EDS), fourier transform infrared (FTIR), nanoscale infrared spectroscopy, and dynamic light scattering (DLS) (**Figure 2**). Here, the review succinctly focused on each instrument's essential operation and how it helps in NP's identification. Firstly, the review discussed about the UV-Visible spectroscopy, among the most crucial methods for the molecular characterisation of metal NPs. ultraviolet-visible spectrophotometer was often employed to monitor the production and stability of Au and Ag-NPs as they were created. As the process progressed, the Chloroauric acid ( $\text{AuCl}_4$ ) and cell-free extract were used to record the UV-visible spectra. Au-NPs exhibit a peak in the surface plasmon resonance at 540 nm, as can be seen from the spectrum [82,83]. The reaction reaches its peak intensity after 48 hours, proving that all of the  $\text{AuCl}_4$  ions have been completely reduced. Similarly, the synthesis and stability of the Ag-NPs produced by the elimination of  $\text{Ag}^+$  ions in the cell-free extract were monitored using UV-visible spectroscopy. The 450 nm band in the spectrum, which corresponds to the surface plasmon resonance of the spherical Ag-NPs, indicates that Ag nanoparticle creation occurred in the reaction mixture [84-86]. Surface plasmon absorbance bands at 525 nm for Au-NPs and 397 nm for Ag-NPs in aqueous solutions served as proof that the citrate-capped Au-NPs and Ag-NPs had been successfully synthesised [87]. The colloidal Au and Ag-NPs distinctive colours were caused by the plasmon absorption. Due to oscillations, incoming light absorbs conduction electrons on nanoparticle surfaces as well as electromagnetic radiation. Distilled water is added to the nanoparticle solution to dilute it, and the absorption maximum is set between 0.5 and 0.7 [88]. To determine particle size, one can utilise the maximal plasmon absorption wavelength in a given solution. The spectrum is made up of (max 400 nm) Ag-NPs [88]. Because  $\text{HAuCl}_4$  and the PA solution don't absorb at this wavelength, the Au-NPs absorbance spectra are detected at around 520 nm.

FTIR measurements in transmission mode with a resolution of  $8\text{ cm}^{-1}$  over the course of 100 scans. The simulation results were contrasted with the experimental Au-NPs and Ag-NPs FT-IR data. The matching metal acetates were used as model compounds in the molecular orbital computations [89]. It has been previously found and published which bands in the FT-IR spectrum of manufactured Au-NPs and Ag-NPs indicate the existence of biomolecules on the surface of NPs, such as enzymes and protein molecules [90,91]. Since the ability of proteins to bind metal is greatest when the carbonyl group is present in amino acid residues and peptides, capped Au, Ag-NPs were created to protect aggregation and stabilise the NPs in the medium. This was demonstrated by an IR spectroscopic investigation [92]. On the other hand, the aggregate nanoparticles' crystallite size is assessed by XRD analysis. The tests can be performed using a Philips diffractometer from the 'X' Pert firm that emits nanochromatic Cu K $\alpha$  (=1.54060) radiation. The size is calculated using Scherrer's formula,  $D=0.86/\cos$ , and the width of the XRD peaks. The products' XRD patterns after using only a small portion of the sample allowed for the measurement and confirmation of the nanoparticle size and structure. Also, the XRD pattern pointed to the three diffraction peaks of Ag-NPs, which corresponded to the (111), (200), and (220) planes (JCPDS No. 040783,) diffractions of face-centered cubic (FCC). The metallic Ag serves as an example of the great pureness of the face centered cubic structure with  $a=4.065$  [93]. However, the monophasic nature of pure Au with FCC symmetry was identified by the XRD arrangement of as-prepared Au-NPs, which shows three peaks at two values of 38.46, 44.65, and 64.73 and is denoted by the indices (1 1 1), (2 0 0), and (2 2 0), respectively (JCPDS No. 011174). The broad, somewhat intense peaks reveal the nanocrystalline structure of Au powder [94-96].

Transmission electron microscopic (TEM) studies is one of the key techniques used for NPs characterisation. In this method, on the carbon-coated copper grids for the TEM approach, a few prepared NPs were inserted. [97]. The TEM apparatus is used to capture micrographs of a drop of NPs while it is operating at, for instance, 200 kv. Grid TEM is dry. An ultra-thin specimen is passed through by a stream of photons, which interact with the object as they do so. An image is produced from the interaction of the electrons as they move through the specimen. An imaging equipment is used to enlarge and concentrate the image [97]. The photos proved that the scattered condition of Au and Ag-NPs was preceded by the ideal starting materials concentration, volume, optimal pH, and incubation time for each reducing agent. [98]. Scanning electron microscope and



EDX analysis, this method is employed to investigate the dimensions and shape of the NPs. For SEM analysis, using a pipet, a small sample was taken and applied to a stub. The stub is composed of copper and has a diameter of around 1 mm. The stub contains double-sided carbon material attached to one side of it. After the sample is put on the carbon material, the stub is fastened to a holder. About seven products can fit in the holder [99]. Using a high-resolution FE-SEM, the Au and Ag-NPs were further analysed. Many spherical, hexagonal, and cubical nanoparticles are generated as a result of the reduction of Au ions, as shown by scanning electron micrographs of the saponin from *T. decandra* was reduced with  $\text{HAuCl}_4$  to produce Au-NPs. These Au-NPs have relative diameters between 37.7 and 79.9 nm revealed that tea polyphenols serve as significant reducing agents [100]. SEM images of spherical NPs with sizes ranging from 17.9 nm to 59.6 nm have led scientists to believe that reducing sugars may have played a role in the converting of  $\text{AgNO}_3$  to Ag-NPs [101]. The Hitachi S-3400N FE-Thermal SEM's EDX attachment was used to carry out the EDX analysis. The reduction of chloroaurate ions was investigated in the current work,  $(\text{AuCl}_4)^-$ , led to the formation of nanotriangles. EDX spectroscopy was used to further demonstrate that there are Au atoms present in the Au-NPs. The optical absorption peak, which is typical for Au nanocrystallite absorption caused by surface plasmon resonance, was measured at 2.2 keV. Strong Au atom signals were visible in the current EDX spectroscopy image of *T. decandra*'s Au-NPs at energies of 0.30, 2, 2.2, 2.4, and 9.7 keV [102].

Furthermore, EDX spectroscopy was used to demonstrate the presence of Ag atoms in the Ag-NPs. According to surface plasmon resonance, the optical absorption peak, which is typical for the absorption of Ag nanocrystallite, was observed at 3 keV. Individual, spherical Ag-NPs made from alfalfa in a previous work displayed absorption peaks in the 2.5–4 keV range [102,103]. Additionally, DLS technique, NPs zeta potential ( $\xi$ ) provides a crucial insight into their surface charge and stability. The DLS analysis's calculated negative zeta potential ( $\xi$ ) values for b-Au-NPs and b-Ag-NPs (= 10 to 16 mV) demonstrate the stability of these nanoparticles over an extended period of time in solution. These nanoparticles are more stable and have a higher dispersion due to the solution has a repelling negative-negative force [104,105]. X-ray photoelectron spectroscopy (XPS), atomic compositions determined by XPS were compared to simulations produced by various models using experimentally determined values [106]. In addition to helping identify the elements present in the sample and determine their oxidation

states, XPS is a significant surface-sensitive analytical technique. The produced Ag nanoparticles revealed 368.3 and 374.3 eV are the binding energy peaks, which are equivalent to Ag 3d<sub>5/2</sub> and Ag 3d<sub>3/2</sub>'s spin-orbit splitting components, respectively. These binding energy peaks are typical of Ag in the zero-oxidation state. At binding energies of 84.3 and 87.9 eV, two spin-orbit splitting components from the high-resolution narrow scan for Au 4f could be extracted, which may be attributed to Au 4f<sub>7/2</sub> and Au 4f<sub>5/2</sub>, dual spin-orbit splitting modes that are unique to Au in metallic form. There is good consistency between the maximum of the measured binding energy for Ag and Au and earlier literature reports [107-109].

#### **4. Biological application of Ag and Au-NPs**

The numerous applications of nanoparticles in the pharmaceutical area were discussed (**Figure 3**). The vast applicability of Ag and Au-NPs is owing to the nanoparticle's new features, which aid in applications such as superior catalysis, good biocompatibility, huge surface area, and conductivity. When nanoparticles are coupled within the fusion of Au/NPs and Au-NPs/MPA with biomolecules (mercaptopropionic acid), biosensors with a large linear range between 0.25 mM and 1.25 mM glucose concentration with a detection limit of 0.025 mM are extensively used [110]. The antimicrobial activity of the produced Au-NPs and Ag-NPs was tested against eight different *S. aureus*, *E. faecalis*, *E. coli*, *P. aeruginosa*, *P. vulgaris*, *B. subtilis*, *Y. enterocolitica*, *K. pneumoniae*, and *C. albicans* are among the several bacteria that have been found. The antibacterial activity of Au-NPs and Ag-NPs against *Y. enterocolitica*, *P. vulgaris*, *E. coli*, *S. aureus*, and *S. faecalis* was very high. Au-NPs inhibited zones ranging from 8.2 mm to 11.5 mm, whereas Ag equivalents inhibited zones ranging from 7.8 mm to 20.3 mm [111]. The nanoparticles exhibit promising antibacterial efficacy against both test microorganisms. Ag-NPs are more biocidal than Au-NPs [112]. In comparison to the gram-negative bacterium *E. coli*, the gram-positive bacterium *S. aureus* was found to be less susceptible to nanoparticles, which may be connected to variations in the cell wall structure. The simplicity, low toxicity of the chemicals used, and creation of practically tiny NPs with acceptable monodispersity and high purity of the metal NPs are the benefits of this technique [113-115].

Furthermore, the antibacterial activity of Ag-NPs the development of pits in the bacterial cell wall was found to be closely related to gram-negative bacteria. As a result, the bacterial membrane developed an accumulation of Ag nanoparticles, which significantly increased

permeability and led to cell death.[116].Au reacts more readily with soft bases including sulphur or phosphorus. As a result, Ag-NPs are primarily attracted to DNA molecules that contain phosphorus as well as proteins that include sulphur that are located in membranes or inside of cells [116]. Furthermore, thiol groups in enzymes like NADH dehydrogenases are bound by Au ions, disrupting the respiratory chain and allowing active oxygen species to be released, resulting in oxidative stress, substantial cell death as a result of harm to the cell's structure [117]. Immobilization and labelling of biomolecules on nanoparticles to produce hybrid molecules has been reported using various methods including specific recognition, covalent coupling, physical adsorption, and electrostatic binding [118]. Among other things, Au-NPs are used in protein, nucleic acid, and targeted transport as well as in vivo gene therapy [119]. Nanoparticles combined with antibodies directed against certain cancer cell surface receptors are used to specifically engage with malignant cells. Functionalized NPs are employed for aimed cell entrance. It has been demonstrated that Au-NPs stabilised by phthalocyanine are a reliable delivery vehicle for photodynamic therapy [120]. To permeate the biological membrane and target the nucleus, functional nanoparticles of Au-NPs with a size of 20 nm were coupled to several cellular targeting peptides. Several nanoparticles have also been used as targeted drug delivery and biomarker agents in cancer detection and treatment. Tumor cells or bacteria can be successfully destroyed by Au-NPs functionalized with specific biomolecules [121]. The high surface-to-volume ratio of Au-NPs allows for the transport of a large number of medicament molecules [122]. Ag nanoparticle application in drug analysis, the placement of Ag-NPs onto supporting is essential for further research into Ag-NP properties. The incorporation of Ag-NPs into graphene oxide sheets, for example, indicates that the antibacterial effectiveness of Ag-graphene oxide nanohybrids was improved over Ag-NPs alone. On rat skin, the Ag/GO hybrids are non-toxic [123]. Additional studies [124] found that Ag/GO nanohybrids have outstanding antibacterial activity. The *Butea monosperma* (BM) leaf extract exhibits a very small cytotoxic effect, whereas the biosynthesized Au and Ag-NPs do not.

However, the *Butea Monosperma* (BM) leaf extract aids in the synthesis of b-Au-500 and b-Ag-750, which could be helpful for the delivery of a variety of medications used in the treatment of tumors. Methanolic/aqueous *B. monosperma* flower extract exhibits both cellular immune responses and anticancer action, according to published literatures [125,126]. Nevertheless, aqueous BM leaf extract cytotoxicity has not been reported. In both the DPPH(2, 2-

diphenyl-1-picrylhydrazyl) and PMA (Phosphomolybdenum assay) assays, the free radicals were decreased by the RECO-GNPs and RECO-SNPs' strong antioxidant properties. The nanoparticle was demonstrated to be cytotoxic to HEP G2 cancer cells using the MTT assay. Gram positive and Gram-negative bacteria (*Proteus vulgaris*) (*Micrococcus luteus*) bacteria were both found to be inhibited by GNPs and SNPs, with Gram negative bacteria showing the strongest antibacterial action [127]. Botanical materials are a rich source of antioxidants. As a result, nanoparticles made from plant sources also have antioxidant action. When compared to nanoparticles created using chemical methods, those created using green methodology had stronger antioxidant capacities. Antioxidant activity of plant-mediated produced Ag/Au nanoparticles has been established [128]. An essential part of the inflammatory process is wound healing, which requires the cooperation of vascular and immune cells to restore tissue integrity [129,130]. For instance, it was discovered that an ointment combining *Falcaria vulgaris* and Au nanoparticles facilitated fibroblast migration and accelerated wound healing [131]. Dermal wound healing was accelerated by Au-NPs created using an extract from *Gundelia tournefortii* L. leaves [132]. In a different work, Ag nanoparticles were produced on a "hydroxyapatite" surface to modify the surface and enhance "hydroxyapatite" adherence. The nanoparticles were stabilised and reduced using polydopamine [133]. The flower extract of *Bauhinia acuminata* was used by others as a reducing and stabilising agent while creating Ag nanoparticles. Mesenchymal stem cell proliferation and osteogenic differentiation were increased by Ag NPs, which aided in the treatment of bone fractures [134]. In order to transfer experimental drugs and genes to a human fibrosarcoma cell line, biocomposites containing Au-NPs made from plant extracts have been used [135].

## **5. Conclusion**

In conclusion, the preparation of Ag-NPs and Au-NPs had been the subject of extensive research due to their special qualities and the potential for many other uses. Various synthesis methods have been developed, including chemical reduction, green synthesis, and biological synthesis.

Each method had its advantages and disadvantages in terms of efficiency, cost, and environmental impact. Ag-NPs have antibacterial properties, making them effective against a broad spectrum of bacteria. They are also biocompatible and can be used as efficient catalysts. However, the release of Ag-NPs into the environment can have a harmful impact on aquatic

organisms, and high doses of Ag nanoparticles can be toxic to living cells. Au-NPs are biocompatible and exhibit unique optical properties that make them useful in biomedical imaging and sensing applications. They can also be functionalized with drugs for targeted drug delivery. However, Au nanoparticles are more expensive than other metal nanoparticles, and their production process and potential release into the environment can have an impact. Au nanoparticles can also be unstable in certain conditions, limiting their use in some applications.

However, although Au and Ag nanoparticles have promising potential in areas such as medical and environmental applications, it is important to carefully consider the potential impact on the environment and the safety of human exposure. Further research is needed to fully understand the implications of using these nanoparticles and to develop sustainable and safe synthesis methods.

Traditional approaches (physical and chemical procedures) for the creation of nanoparticles call for hazardous substances that have negative side effects. Biosynthesis, on the other hand, results in nanoparticles with more clearly defined sizes and morphologies. Humans also benefit from biosynthetic routes in other ways, such as lowering the use of organic solvent-free media, the consumption of hazardous materials and waste, the formation of waste, the use of ambient temperature and pressure for synthesis to considerably reduce energy consumption, and providing inexpensive, one-step, eco-friendly processes and higher efficiency. Using plants to create biogenic synthesis could use to create non-spherical metallic nanostructures with regulated forms and sizes that may be less harmful than those created using standard chemical solvents. Depending on the inorganic material nano compounds and the type of plant medium utilized (extract vs solution), Moreover, the presence of biologically efficient compounds such as amino acids, proteins, flavonoids, and carboxylic acids gives biosynthesized metal nanoparticles stronger antioxidant capabilities than those made by conventional methods.

Overall, the potential future applications of Ag and Au nanoparticles are vast and diverse. As research continues, new applications and uses for these nanoparticles are likely to emerge, making them an important and versatile area of study. However, it is important to carefully consider the potential environmental and health impacts of their use, and to develop safe and sustainable production methods.

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**Conflict of interest:**

The authors declare that there is no conflict of interest in the review.

**Author contributions:**

All the authors equally contributed to writing the manuscript.

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**References:**

1. Ahmad, F., Salem-Bekhit, M.M., Khan, F., Alshehri, S., Khan, A., Ghoneim, M.M., Wu, H.F., Taha, E.I. and Elbagory, I., 2022. Unique properties of surface-functionalized nanoparticles for bio-application: Functionalization mechanisms and importance in application. *Nanomaterials*, 12(8), p.1333.
2. Tarafdar, J.C., Sharma, S. and Raliya, R., 2013. Nanotechnology: Interdisciplinary science of applications. *African Journal of Biotechnology*, 12(3).
3. What Does Nano Mean? Swiss Nanoscience Institute. Available online: <https://nanoscience.ch/en/about-us/nanosciences/whatdoes-nano-mean/> (accessed on 9 February 2022).
4. Dikshit, P.K., Kumar, J., Das, A.K., Sadhu, S., Sharma, S., Singh, S., Gupta, P.K. and Kim, B.S., 2021. Green synthesis of metallic nanoparticles: Applications and limitations. *Catalysts*, 11(8), p.902.
5. Hemanth, N.R., Mohili, R.D., Patel, M., Jadhav, A.H., Lee, K. and Chaudhari, N.K., 2022. Metallic nanosponges for energy storage and conversion applications. *Journal of Materials Chemistry A*, 10(27), pp.14221-14246.
6. Tran, P.A. and Webster, T.J., 2011. Selenium nanoparticles inhibit Staphylococcus aureus growth. *International journal of nanomedicine*, pp.1553-1558.

7. Sun, S.N., Wei, C., Zhu, Z.Z., Hou, Y.L., Venkatraman, S.S. and Xu, Z.C., 2014. Magnetic iron oxide nanoparticles: Synthesis and surface coating techniques for biomedical applications. *Chinese Physics B*, 23(3), p.037503.
8. Neouze, M.A. and Schubert, U., 2008. Surface modification and functionalization of metal and metal oxide nanoparticles by organic ligands. *Monatshefte für Chemie-Chemical Monthly*, 139, pp.183-195.
9. McNamara, K. and Tofail, S.A., 2015. Nanosystems: the use of nanoalloys, metallic, bimetallic, and magnetic nanoparticles in biomedical applications. *Physical chemistry chemical physics*, 17(42), pp.27981-27995.
10. Ferdous, Z. and Nemmar, A., 2020. Health impact of silver nanoparticles: a review of the biodistribution and toxicity following various routes of exposure. *International journal of molecular sciences*, 21(7), p.2375.
11. Lee, S.H. and Jun, B.H., 2019. Silver nanoparticles: synthesis and application for nanomedicine. *International journal of molecular sciences*, 20(4), p.865.
12. Thakkar, K.N., Mhatre, S.S. and Parikh, R.Y., 2010. Biological synthesis of metallic nanoparticles. *Nanomedicine: nanotechnology, biology and medicine*, 6(2), pp.257-262.
13. Hu, D., Ogawa, K., Kajiyama, M. and Enomae, T., 2020. Characterization of self-assembled silver nanoparticle ink based on nanoemulsion method. *Royal Society Open Science*, 7(5), p.200296.
14. Zhang, H., Zhou, H., Bai, J., Li, Y., Yang, J., Ma, Q. and Qu, Y., 2019. Biosynthesis of selenium nanoparticles mediated by fungus *Mariannaea* sp. HJ and their characterization. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 571, pp.9-16.
15. Adil, S.F., Assal, M.E., Khan, M., Al-Warthan, A., Siddiqui, M.R.H. and Liz-Marzán, L.M., 2015. Biogenic synthesis of metallic nanoparticles and prospects toward green chemistry. *Dalton Transactions*, 44(21), pp.9709-9717.
16. Lee, K.X., Shameli, K., Yew, Y.P., Teow, S.Y., Jahangirian, H., Rafiee-Moghaddam, R. and Webster, T.J., 2020. Recent developments in the facile bio-synthesis of gold nanoparticles (Au-NPss) and their biomedical applications. *International journal of nanomedicine*, pp.275-300.
17. Malaikozhundan, B., Vinodhini, J., Manivannan, N., Boopathi, T. and Vijayakumar, S., 2022. Bioapplications of nanoparticles. In *Nano-Bioremediation: Fundamentals and Applications* (pp. 213-239). Elsevier.
18. Sampath, G., Govarthanam, M., Rameshkumar, N., Vo, D.V.N., Krishnan, M., Sivasankar, P. and Kayalvizhi, N., 2021. Eco-friendly biosynthesis metallic silver nanoparticles using *Aegle marmelos* (Indian bael) and its clinical and environmental applications. *Applied Nanoscience*, pp.1-12.
19. Bruna, T., Maldonado-Bravo, F., Jara, P. and Caro, N., 2021. Silver nanoparticles and their antibacterial applications. *International Journal of Molecular Sciences*, 22(13), p.7202.
20. Rana, K.; Pandey, S.K.; Chauhan, S.; Preet, S. Anticancer therapeutic potential of 5-fluorouracil and nisin co-loaded chitosan coated Ag-NPs against murine skin cancer. *Int. J. Pharm.* 2022, 620, 121744. [CrossRef]

21. Wahab, M.A.; Li, L.; Li, H.; Abdala, A. Ag nanoparticle-based nanocomposites for combating infectious pathogens: Recent advances and future prospects. *Nanomaterials* 2021, 11, 581. [CrossRef]
22. Tian, L., Qian, K., Qi, J., Liu, Q., Yao, C., Song, W. and Wang, Y., 2018. Gold nanoparticles superlattices assembly for electrochemical biosensor detection of microRNA-21. *Biosensors and Bioelectronics*, 99, pp.564-570.
23. Xiong, Z., Alves, C.S., Wang, J., Li, A., Liu, J., Shen, M., Rodrigues, J., Tomás, H. and Shi, X., 2019. Zwitterion-functionalized dendrimer-entrapped gold nanoparticles for serum-enhanced gene delivery to inhibit cancer cell metastasis. *Acta Biomaterialia*, 99, pp.320-329.
24. Cárcamo-Martínez, Á., Domínguez-Robles, J., Mallon, B., Raman, M.T., Cordeiro, A.S., Bell, S.E., Larrañeta, E. and Donnelly, R.F., 2020. Potential of polymeric films loaded with gold nanorods for local hyperthermia applications. *Nanomaterials*, 10(3), p.582.
25. Li, X., Xing, L., Zheng, K., Wei, P., Du, L., Shen, M. and Shi, X., 2017. Formation of gold nanostar-coated hollow mesoporous silica for tumor multimodality imaging and photothermal therapy. *ACS applied materials & interfaces*, 9(7), pp.5817-5827.
26. Mehravani, B., Ribeiro, A.I. and Zille, A., 2021. Gold nanoparticles synthesis and antimicrobial effect on fibrous materials. *Nanomaterials*, 11(5), p.1067.
27. Qiu, J., Liu, Y. and Xia, Y., 2021. Radiolabeling of gold nanocages for potential applications in tracking, diagnosis, and image-guided therapy. *Advanced healthcare materials*, 10(15), p.2002031.
28. Brigger, I., Dubernet, C. and Couvreur, P., 2012. Nanoparticles in cancer therapy and diagnosis. *Advanced drug delivery reviews*, 64, pp.24-36.
29. Lipka, J., Semmler-Behnke, M., Sperling, R.A., Wenk, A., Takenaka, S., Schleh, C., Kissel, T., Parak, W.J. and Kreyling, W.G., 2010. Biodistribution of PEG-modified gold nanoparticles following intratracheal instillation and intravenous injection. *Biomaterials*, 31(25), pp.6574-6581.
30. Iravani, S., Korbekandi, H., Mirmohammadi, S.V. and Zolfaghari, B., 2014. Synthesis of silver nanoparticles: chemical, physical and biological methods. *Research in pharmaceutical sciences*, 9(6), p.385.
31. Slepíčka, P., SlepíčkováKasálková, N., Siegel, J., Kolská, Z. and Švorčík, V., 2019. Methods of gold and silver nanoparticles preparation. *Materials*, 13(1), p.1.
32. Song, J.Y. and Kim, B.S., 2009. Rapid biological synthesis of silver nanoparticles using plant leaf extracts. *Bioprocess and biosystems engineering*, 32, pp.79-84.
33. Zhang, X.F., Liu, Z.G., Shen, W. and Gurunathan, S., 2016. Silver nanoparticles: synthesis, characterization, properties, applications, and therapeutic approaches. *International journal of molecular sciences*, 17(9), p.1534.
34. Klaus, T., Joerger, R., Olsson, E. and Granqvist, C.G., 1999. Silver-based crystalline nanoparticles, microbially fabricated. *Proceedings of the National Academy of Sciences*, 96(24), pp.13611-13614.
35. Konishi, Y., Ohno, K., Saitoh, N., Nomura, T., Nagamine, S., Hishida, H., Takahashi, Y. and Uruga, T., 2007. Bioreductive deposition of platinum nanoparticles on the bacterium *Shewanella* algae. *Journal of biotechnology*, 128(3), pp.648-653.



36. Nair, B. and Pradeep, T., 2002. Coalescence of nanoclusters and formation of submicron crystallites assisted by Lactobacillus strains. *Crystal growth & design*, 2(4), pp.293-298.
37. Willner, I., Baron, R. and Willner, B., 2006. Growing metal nanoparticles by enzymes. *Advanced Materials*, 18(9), pp.1109-1120.
38. Shankar, S.S., Rai, A., Ahmad, A. and Sastry, M., 2004. Rapid synthesis of Au, Ag, and bimetallic Au core–Ag shell nanoparticles using Neem (*Azadirachta indica*) leaf broth. *Journal of colloid and interface science*, 275(2), pp.496-502.
39. Graily-Moradi, F., MaadaniMallak, A. and Ghorbanpour, M., 2020. Biogenic synthesis of gold nanoparticles and their potential application in agriculture. *Biogenic nano-particles and their use in agro-ecosystems*, pp.187-204.
40. Gudikandula, Krishna, and SingaraCharyaMaringanti. "Synthesis of silver nanoparticles by chemical and biological methods and their antimicrobial properties." *Journal of experimental nanoscience* 11, no. 9 (2016): 714-721.
41. Guzmán, M.G., Dille, J. and Godet, S., 2009. Synthesis of silver nanoparticles by chemical reduction method and their antibacterial activity. *Int J Chem BiomolEng*, 2(3), pp.104-111.
42. Jose, M. and Sakthivel, M., 2014. Synthesis and characterization of silver nanospheres in mixed surfactant solution. *Materials Letters*, 117, pp.78-81.
43. Khatoon, U.T., Rao, G.N., Mohan, K.M., Ramanaviciene, A. and Ramanavicius, A., 2017. Antibacterial and antifungal activity of silver nanospheres synthesized by tri-sodium citrate assisted chemical approach. *Vacuum*, 146, pp.259-265.
44. Fatimah, I., Hidayat, H., Nugroho, B. and Husein, S., 2023. Green synthesis of silver nanoparticles using Datura metel flower extract assisted by ultrasound method and its antibacterial activity. *Recent Patents on Nanotechnology*, 17(1), pp.68-73.
45. Begum, I., Shamim, S., Ameen, F., Hussain, Z., Bhat, S.A., Qadri, T. and Hussain, M., 2022. A combinatorial approach towards antibacterial and antioxidant activity using tartaric acid capped silver nanoparticles. *Processes*, 10(4), p.716.
46. Priyadarshi, R. and Negi, Y.S., 2019. Poly (vinyl pyrrolidone)-mediated synthesis of silver nanowires decorated with silver nanospheres and their antimicrobial activity. *Bulletin of Materials Science*, 42, pp.1-7.
47. Quintero-Quiroz, C., Acevedo, N., Zapata-Giraldo, J., Botero, L.E., Quintero, J., Zárate-Triviño, D., Saldarriaga, J. and Pérez, V.Z., 2019. Optimization of silver nanoparticle synthesis by chemical reduction and evaluation of its antimicrobial and toxic activity. *Biomaterials research*, 23(1), pp.1-15.
48. Alharbi, N.S., Alsubhi, N.S. and Felimban, A.I., 2022. Green synthesis of silver nanoparticles using medicinal plants: Characterization and application. *Journal of Radiation Research and Applied Sciences*, 15(3), pp.109-124.
49. Al-Ghamdi, H.S. and Mahmoud, W.E., 2013. One pot synthesis of multi-plasmonic shapes of silver nanoparticles. *Materials Letters*, 105, pp.62-64.
50. Al-Mashhadani, T.A. and Al-Maliki, F.J., 2023. Effect of Silver Colloidal Concentration on Morphology of Silver Nanostructures Prepared by Chemical Reduction Method. *Iraqi Journal of Applied Physics Letters*, 6(1).

51. Song, Y.J., Wang, M., Zhang, X.Y., Wu, J.Y. and Zhang, T., 2014. Investigation on the role of the molecular weight of polyvinyl pyrrolidone in the shape control of high-yield silver nanospheres and nanowires. *Nanoscale research letters*, 9, pp.1-8.
52. Jayaprakash, N., Vijaya, J.J. and Kennedy, L.J., 2015. Microwave-Assisted Rapid Facile Synthesis, Characterization, and Their Antibacterial Activity of PVP Capped Silver Nanospheres. *Synthesis and Reactivity in Inorganic, Metal-Organic, and Nano-Metal Chemistry*, 45(10), pp.1533-1538.
53. Naaz, F., Farooq, U., Khan, M.M. and Ahmad, T., 2020. Multifunctional efficacy of environmentally benign silver nanospheres for organic transformation, photocatalysis, and water remediation. *ACS omega*, 5(40), pp.26063-26076.
54. Chou, K.S., Lu, Y.C. and Lee, H.H., 2005. Effect of alkaline ion on the mechanism and kinetics of chemical reduction of silver. *Materials Chemistry and Physics*, 94(2-3), pp.429-433.
55. Lin, W.C. and Yang, M.C., 2005. Novel Silver/Poly (vinyl alcohol) Nanocomposites for Surface- Enhanced Raman Scattering- Active Substrates. *Macromolecular rapid communications*, 26(24), pp.1942-1947.
56. Shin, H.S., Yang, H.J., Kim, S.B. and Lee, M.S., 2004. Mechanism of growth of colloidal silver nanoparticles stabilized by polyvinyl pyrrolidone in  $\gamma$ -irradiated silver nitrate solution. *Journal of colloid and interface science*, 274(1), pp.89-94.
57. Do Kim, K., Han, D.N. and Kim, H.T., 2004. Optimization of experimental conditions based on the Taguchi robust design for the formation of nano-sized silver particles by chemical reduction method. *Chemical Engineering Journal*, 104(1-3), pp.55-61.
58. Zhao, P., Li, N. and Astruc, D., 2013. State of the art in gold nanoparticle synthesis. *Coordination Chemistry Reviews*, 257(3-4), pp.638-665.
59. Hu, M., Chen, J., Li, Z.Y., Au, L., Hartland, G.V., Li, X., Marquez, M. and Xia, Y., 2006. Gold nanostructures: engineering their plasmonic properties for biomedical applications. *Chemical Society Reviews*, 35(11), pp.1084-1094.
60. Dong, J., Carpinone, P.L., Pyrgiotakis, G., Demokritou, P. and Moudgil, B.M., 2020. Synthesis of precision gold nanoparticles using Turkevich method. *KONA Powder and Particle Journal*, 37, pp.224-232.
61. Zhu, L., Zhang, C., Guo, C., Wang, X., Sun, P., Zhou, D., Chen, W. and Xue, G., 2013. New insight into intermediate precursors of Brust-Schiffrin gold nanoparticles synthesis. *The Journal of Physical Chemistry C*, 117(21), pp.11399-11404.
62. Booth, S.G., Uehara, A., Chang, S.Y., La Fontaine, C., Fujii, T., Okamoto, Y., Imai, T., Schroeder, S.L. and Dryfe, R.A.W., 2017. The significance of bromide in the Brust-Schiffrin synthesis of thiol protected gold nanoparticles. *Chemical Science*, 8(12), pp.7954-7962.
63. Liu, X., Worden, J.G., Huo, Q. and Brennan, J.P., 2006. Kinetic study of gold nanoparticle growth in solution by Brust-Schiffrin reaction. *Journal of nanoscience and nanotechnology*, 6(4), pp.1054-1059.
64. Hamamoto, M. and Yagyu, H., 2017, July. Two-phase Brust-Schiffrin synthesis of gold nanoparticles dispersion in organic solvent on glass microfluidic device. In *2017 IEEE 17th International Conference on Nanotechnology (IEEE-NANO)* (pp. 632-635). IEEE.

65. Ma, H., Yin, B., Wang, S., Jiao, Y., Pan, W., Huang, S., Chen, S. and Meng, F., 2004. Synthesis of silver and gold nanoparticles by a novel electrochemical method. *ChemPhysChem*, 5(1), pp.68-75.
66. Chandra, P., Singh, J., Singh, A., Srivastava, A., Goyal, R.N. and Shim, Y.B., 2013. Gold nanoparticles and nanocomposites in clinical diagnostics using electrochemical methods. *Journal of Nanoparticles*, 2013.
67. Zou, C.E., Yang, B., Bin, D., Wang, J., Li, S., Yang, P., Wang, C., Shiraishi, Y. and Du, Y., 2017. Electrochemical synthesis of gold nanoparticles decorated flower-like graphene for high sensitivity detection of nitrite. *Journal of colloid and interface science*, 488, pp.135-141.
68. Dong, S., Tang, C., Zhou, H. and Zhao, H., 2004. Photochemical synthesis of gold nanoparticles by the sunlight radiation using a seeding approach. *Gold bulletin*, 37(3-4), pp.187-195.
69. Ziegler, C. and Eychmuller, A., 2011. Seeded growth synthesis of uniform gold nanoparticles with diameters of 15– 300 nm. *The Journal of Physical Chemistry C*, 115(11), pp.4502-4506.
70. Jenkins, J.A., Wax, T.J. and Zhao, J., 2017. Seed-mediated synthesis of gold nanoparticles of controlled sizes to demonstrate the impact of size on optical properties. *Journal of Chemical Education*, 94(8), pp.1090-1093.
71. Rioux, D. and Meunier, M., 2015. Seeded growth synthesis of composition and size-controlled gold–silver alloy nanoparticles. *The Journal of Physical Chemistry C*, 119(23), pp.13160-13168.
72. Bastús, N.G., Comenge, J. and Puntès, V., 2011. Kinetically controlled seeded growth synthesis of citrate-stabilized gold nanoparticles of up to 200 nm: size focusing versus Ostwald ripening. *Langmuir*, 27(17), pp.11098-11105.
73. Xiao, J. and Qi, L., 2011. Surfactant-assisted, shape-controlled synthesis of gold nanocrystals. *Nanoscale*, 3(4), pp.1383-1396.
74. Sanchez-Gaytan, B.L., Qian, Z., Hastings, S.P., Reca, M.L., Fakhraei, Z. and Park, S.J., 2013. Controlling the topography and surface plasmon resonance of gold nanoshells by a templated surfactant-assisted seed growth method. *The Journal of Physical Chemistry C*, 117(17), pp.8916-8923.
75. Smith, D.K. and Korgel, B.A., 2008. The importance of the CTAB surfactant on the colloidal seed-mediated synthesis of gold nanorods. *Langmuir*, 24(3), pp.644-649.
76. Hammami, I. and Alabdallah, N.M., 2021. Gold nanoparticles: Synthesis properties and applications. *Journal of king Saud university-science*, 33(7), p.101560.
77. Vanlalveni, C., Lallianrawna, S., Biswas, A., Selvaraj, M., Changmai, B. and Rokhum, S.L., 2021. Green synthesis of silver nanoparticles using plant extracts and their antimicrobial activities: A review of recent literature. *RSC advances*, 11(5), pp.2804-2837.
78. Huang, J., Li, Q., Sun, D., Lu, Y., Su, Y., Yang, X., Wang, H., Wang, Y., Shao, W., He, N. and Hong, J., 2007. Biosynthesis of silver and gold nanoparticles by novel sundried *Cinnamomum camphora* leaf. *Nanotechnology*, 18(10), p.105104.

79. Shahverdi, A.R., Minaeian, S., Shahverdi, H.R., Jamalifar, H. and Nohi, A.A., 2007. Rapid synthesis of silver nanoparticles using culture supernatants of Enterobacteria: a novel biological approach. *Process Biochemistry*, 42(5), pp.919-923.
80. Shahverdi, A.R., Minaeian, S., Shahverdi, H.R., Jamalifar, H. and Nohi, A.A., 2007. Rapid synthesis of silver nanoparticles using culture supernatants of Enterobacteria: a novel biological approach. *Process Biochemistry*, 42(5), pp.919-923.
81. Singh, P., Kim, Y.J., Zhang, D. and Yang, D.C., 2016. Biological synthesis of nanoparticles from plants and microorganisms. *Trends in biotechnology*, 34(7), pp.588-599.
82. Ahmad, T., Wani, I.A., Lone, I.H., Ganguly, A., Manzoor, N., Ahmad, A., Ahmed, J. and Al-Shihri, A.S., 2013. Antifungal activity of Au-NPs prepared by solvothermal method. *Materials Research Bulletin*, 48(1), pp.12-20.
83. Wani, I.A. and Ahmad, T., 2013. Size and shape dependant antifungal activity of gold nanoparticles: a case study of Candida. *Colloids and surfaces B: Biointerfaces*, 101, pp.162-170.
84. Wani, I.A., Khatoon, S., Ganguly, A., Ahmed, J., Ganguli, A.K. and Ahmad, T., 2010. Silver nanoparticles: Large scale solvothermal synthesis and optical properties. *Materials Research Bulletin*, 45(8), pp.1033-1038.
85. Wani, I.A., Ganguly, A., Ahmed, J. and Ahmad, T., 2011. Silver nanoparticles: ultrasonic wave assisted synthesis, optical characterization and surface area studies. *Materials Letters*, 65(3), pp.520-522.
86. Wani, I.A., Khatoon, S., Ganguly, A., Ahmed, J. and Ahmad, T., 2013. Structural characterization and antimicrobial properties of Ag-NPs prepared by inverse microemulsion method. *Colloids and Surfaces B: Biointerfaces*, 101, pp.243-250.
87. Wulandari, P., Nagahiro, T., Fukada, N., Kimura, Y., Niwano, M. and Tamada, K., 2015. Characterization of citrates on gold and silver nanoparticles. *Journal of colloid and interface science*, 438, pp.244-248.
88. Mulfinger, L., Solomon, S.D., Bahadory, M., Jeyarajasingam, A.V., Rutkowsky, S.A. and Boritz, C., 2007. Synthesis and study of silver nanoparticles. *Journal of chemical education*, 84(2), p.322.
89. Wulandari, P., Nagahiro, T., Michioka, K., Tamada, K., Ishibashi, K.I., Kimura, Y. and Niwano, M., 2008. Coordination of carboxylate on metal nanoparticles characterized by Fourier transform infrared spectroscopy. *Chemistry letters*, 37(8), pp.888-889.
90. Shaligram, N.S., Bule, M., Bhambure, R., Singhal, R.S., Singh, S.K., Szakacs, G. and Pandey, A., 2009. Biosynthesis of silver nanoparticles using aqueous extract from the compactin producing fungal strain. *Process biochemistry*, 44(8), pp.939-943.
91. Gajbhiye, M., Kesharwani, J., Ingle, A., Gade, A. and Rai, M., 2009. Fungus-mediated synthesis of silver nanoparticles and their activity against pathogenic fungi in combination with fluconazole. *Nanomedicine: Nanotechnology, Biology and Medicine*, 5(4), pp.382-386.
92. Gole, A., Dash, C., Ramakrishnan, V., Sainkar, S.R., Mandale, A.B., Rao, M. and Sastry, M., 2001. Pepsin- gold colloid conjugates: preparation, characterization, and enzymatic activity. *Langmuir*, 17(5), pp.1674-1679.
93. Alagumuthu, G. and Kirubha, R., 2012. Synthesis and characterisation of Ag-NPs in different medium. *Open Journal of Synthesis Theory and Applications*, 1(2), pp.13-17.

94. Ahmad, T., Wani, I.A., Manzoor, N., Ahmed, J. and Asiri, A.M., 2013. Biosynthesis, structural characterization and antimicrobial activity of gold and silver nanoparticles. *Colloids and Surfaces B: Biointerfaces*, 107, pp.227-234.
95. Patra, S., Mukherjee, S., Barui, A.K., Ganguly, A., Sreedhar, B. and Patra, C.R., 2015. Green synthesis, characterization of Au and Ag-NPs and their potential application for cancer therapeutics. *Materials Science and Engineering: C*, 53, pp.298-309
96. Balaji, D.S., Basavaraja, S., Deshpande, R., Mahesh, D.B., Prabhakar, B.K. and Venkataraman, A., 2009. Extracellular biosynthesis of functionalized silver nanoparticles by strains of *Cladosporium cladosporioides* fungus. *Colloids and surfaces B: biointerfaces*, 68(1), pp.88-92.
97. Krishnamoorthy, P. and Jayalakshmi, T., 2012. Preparation, characterization and synthesis of Ag-NPs by using phyllanthusniruri for the antimicrobial activity and cytotoxic effects. *J. Chem. Pharm. Res*, 4(11), pp.4783-4794.
98. Reidy, B., Haase, A., Luch, A., Dawson, K.A. and Lynch, I., 2013. Mechanisms of silver nanoparticle release, transformation and toxicity: a critical review of current knowledge and recommendations for future studies and applications. *Materials*, 6(6), pp.2295-2350.
99. Priya, T.S. and Balasubramanian, V., 2014. Enzyme mediated synthesis of silver nanoparticles using marine actinomycetes and their characterization. *Biosciences Biotechnology Research Asia*, 11, pp.159-165.
100. Begum, N.A., Mondal, S., Basu, S., Laskar, R.A. and Mandal, D., 2009. Biogenic synthesis of Au and Ag nanoparticles using aqueous solutions of Black Tea leaf extracts. *Colloids and surfaces B: Biointerfaces*, 71(1), pp.113-118.
101. Shankar, S.S., Rai, A., Ahmad, A. and Sastry, M., 2005. Controlling the optical properties of lemongrass extract synthesized gold nanotriangles and potential application in infrared-absorbing optical coatings. *Chemistry of Materials*, 17(3), pp.566-572.
102. Gardea-Torresdey, J.L., Gomez, E., Peralta-Videa, J.R., Parsons, J.G., Troiani, H. and Jose-Yacaman, M., 2003. Alfalfa sprouts: a natural source for the synthesis of silver nanoparticles. *Langmuir*, 19(4), pp.1357-1361.
103. Ali, M., Kim, B., Belfield, K.D., Norman, D., Brennan, M. and Ali, G.S., 2016. Green synthesis and characterization of silver nanoparticles using *Artemisia absinthium* aqueous extract—a comprehensive study. *Materials Science and Engineering: C*, 58, pp.359-365.

104. Shukla, R., Nune, S.K., Chanda, N., Katti, K., Mekapothula, S., Kulkarni, R.R., Welshons, W.V., Kannan, R. and Katti, K.V., 2008. Soybeans as a phytochemical reservoir for the production and stabilization of biocompatible gold nanoparticles. *Small*, 4(9), pp.1425-1436.
105. Göl, F., Aygün, A., Seyrankaya, A., Gür, T., Yenikaya, C. and Şen, F., 2020. Green synthesis and characterization of *Camellia sinensis* mediated silver nanoparticles for antibacterial ceramic applications. *Materials Chemistry and Physics*, 250, p.123037.
106. Yung-Chen, W., 2016. Quantifying the Impact of Nanoparticle Coatings and Nonuniformities on XPS Analysis: Gold/Silver Core–Shell Nanoparticles.
107. Rajasekharreddy, P., Usha Rani, P. and Sreedhar, B., 2010. Qualitative assessment of silver and gold nanoparticle synthesis in various plants: a photobiological approach. *Journal of Nanoparticle Research*, 12, pp.1711-1721.
108. Ahmad, A., Senapati, S., Khan, M.I., Kumar, R. and Sastry, M., 2005. Extra-/intracellular biosynthesis of gold nanoparticles by an alkalotolerant fungus, *Trichothecium* sp. *Journal of Biomedical Nanotechnology*, 1(1), pp.47-53.
109. Huang, Y.F., Huang, K.M. and Chang, H.T., 2006. Synthesis and characterization of Au core–Au–Ag shell nanoparticles from gold seeds: Impacts of glycine concentration and pH. *Journal of colloid and interface science*, 301(1), pp.145-154.
110. Rastogi, L., Kora, A.J. and Arunachalam, J., 2012. Highly stable, protein capped gold nanoparticles as effective drug delivery vehicles for amino-glycosidic antibiotics. *Materials Science and Engineering: C*, 32(6), pp.1571-1577.
111. Ghosh, S., Patil, S., Ahire, M., Kitture, R., Kale, S., Pardesi, K., Cameotra, S.S., Bellare, J., Dhavale, D.D., Jabgunde, A. and Chopade, B.A., 2012. Synthesis of silver nanoparticles using *Dioscorea bulbifera* tuber extract and evaluation of its synergistic potential in combination with antimicrobial agents. *International journal of nanomedicine*, pp.483-496.
112. Kim, J.S., Kuk, E., Yu, K.N., Kim, J.H., Park, S.J., Lee, H.J., Kim, S.H., Park, Y.K., Park, Y.H., Hwang, C.Y. and Kim, Y.K., 2007. Antimicrobial effects of silver nanoparticles. *Nanomedicine: Nanotechnology, biology and medicine*, 3(1), pp.95-101.
113. Morones, J.R., Elechiguerra, J.L., Camacho, A., Holt, K., Kouri, J.B., Ramírez, J.T. and Yacaman, M.J., 2005. The bactericidal effect of silver nanoparticles. *Nanotechnology*, 16(10), p.2346.

114. Rai, A., Prabhune, A. and Perry, C.C., 2010. Antibiotic mediated synthesis of gold nanoparticles with potent antimicrobial activity and their application in antimicrobial coatings. *Journal of Materials Chemistry*, 20(32), pp.6789-6798.
115. Burygin, G.L., Khlebtsov, B.N., Shantrokha, A.N., Dykman, L.A., Bogatyrev, V.A. and Khlebtsov, N.G., 2009. On the enhanced antibacterial activity of antibiotics mixed with gold nanoparticles. *Nanoscale research letters*, 4, pp.794-801.
116. Sondi, I. and Salopek-Sondi, B., 2004. Silver nanoparticles as antimicrobial agent: a case study on E. coli as a model for Gram-negative bacteria. *Journal of colloid and interface science*, 275(1), pp.177-182.
117. Amutha, R., Arumugam, P. and Berchmans, S., 2011. Synthesis of gold nanoparticles: an ecofriendly approach using Hansenulaanomala. *ACS applied materials & interfaces*, 3(5), pp.1418-1425.
118. Daraee, H., Eatemadi, A., Abbasi, E., Aval, S.F., Kouhi, M. and Akbarzadeh, A., 2016. Application of Au Nps in Biomedical and Drug Delivery, *Artif. Cells. Nanomed. Biotechnol*, 44, pp.410-422.
119. Raghvendra, R., Kanthadeivi, A., Sathesh, K.A. and Aarrthy, M.A., 2014. Diagnostics and therapeutic application of gold nanoparticles. *Medicine*, 2, p.4.
120. Doubrovsky, V.A., Yanina, I.Y. and Tuchin, V.V., 2011, April. Inhomogeneity of photo-induced fat cell lipolysis. In *Saratov Fall Meeting 2010: Optical Technologies in Biophysics and Medicine XII* (Vol. 7999, pp. 145-153). SPIE.
121. Duncan, B., Kim, C. and Rotello, V.M., 2010. Gold nanoparticle platforms as drug and biomacromolecule delivery systems. *Journal of controlled release*, 148(1), pp.122-127.
122. Grace, A.N. and Pandian, K., 2007. Antibacterial efficacy of aminoglycosidic antibiotics protected gold nanoparticles—A brief study. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 297(1-3), pp.63-70.
123. Xu, W.P., Zhang, L.C., Li, J.P., Lu, Y., Li, H.H., Ma, Y.N., Wang, W.D. and Yu, S.H., 2011. Facile synthesis of silver@ graphene oxide nanocomposites and their enhanced antibacterial properties. *Journal of Materials Chemistry*, 21(12), pp.4593-4597.
124. Das, M.R., Sarma, R.K., Borah, S.C., Kumari, R., Saikia, R., Deshmukh, A.B., Shelke, M.V., Sengupta, P., Szunerits, S. and Boukherroub, R., 2013. The synthesis of citrate-

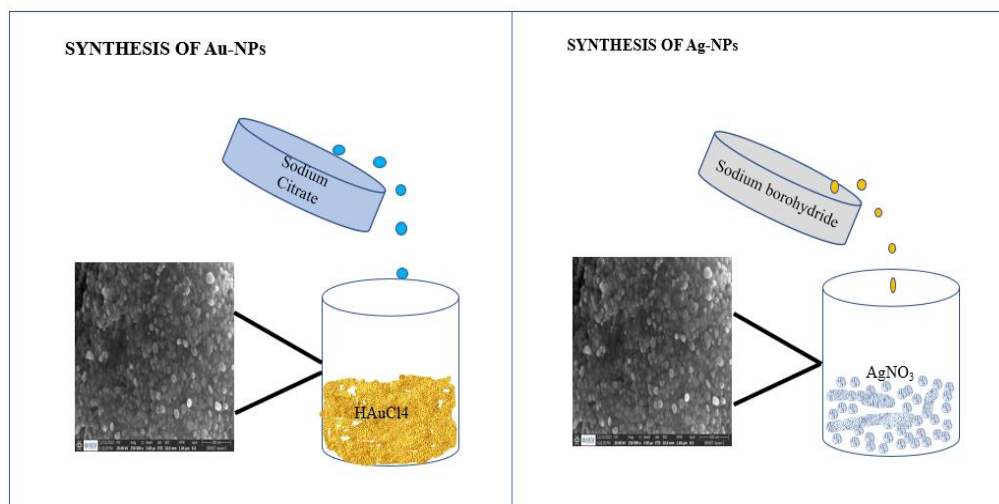
- modified silver nanoparticles in an aqueous suspension of graphene oxide nanosheets and their antibacterial activity. *Colloids and Surfaces B: Biointerfaces*, 105, pp.128-136.
125. Choedon, T., Shukla, S.K. and Kumar, V., 2010. Chemopreventive and anti-cancer properties of the aqueous extract of flowers of *Butea monosperma*. *Journal of ethnopharmacology*, 129(2), pp.208-213.
126. Rasheed, Z., Akhtar, N., Khan, A., Khan, K.A. and Haqqi, T.M., 2010. Butrin, isobutrin, and butein from medicinal plant *Butea monosperma* selectively inhibit nuclear factor- $\kappa$ B in activated human mast cells: Suppression of tumor necrosis factor- $\alpha$ , interleukin (IL)-6, and IL-8. *Journal of Pharmacology and Experimental Therapeutics*, 333(2), pp.354-363.
127. Dhayalan, M., Denison, M.I.J., Ayyar, M., Gandhi, N.N., Krishnan, K. and Abdulhadi, B., 2018. Biogenic synthesis, characterization of gold and silver nanoparticles from *Coleus forskohlii* and their clinical importance. *Journal of Photochemistry and Photobiology B: Biology*, 183, pp.251-257.
128. Zayed, M.F., Mahfoze, R.A., El-Kousy, S.M. and Al-Ashkar, E.A., 2020. In-vitro antioxidant and antimicrobial activities of metal nanoparticles biosynthesized using optimized *Pimpinella anisum* extract. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 585, p.124167.
129. Manikandan, R., Anjali, R., Beulaja, M., Prabhu, N.M., Koodalingam, A., Saiprasad, G., Chitra, P. and Arumugam, M., 2019. Synthesis, characterization, anti-proliferative and wound healing activities of silver nanoparticles synthesized from *Caulerpa scalpelliformis*. *Process biochemistry*, 79, pp.135-141.
130. Tahvilian, R., Zangeneh, M.M., Falahi, H., Sadrjavadi, K., Jalalvand, A.R. and Zangeneh, A., 2019. Green synthesis and chemical characterization of copper nanoparticles using *Allium saralicum* leaves and assessment of their cytotoxicity, antioxidant, antimicrobial, and cutaneous wound healing properties. *Applied organometallic chemistry*, 33(12), p.e5234.
131. Zangeneh, M.M., Saneei, S., Zangeneh, A., Tushmalani, R., Haddadi, A., Almasi, M. and Amiri- Paryan, A., 2019. Preparation, characterization, and evaluation of cytotoxicity, antioxidant, cutaneous wound healing, antibacterial, and antifungal effects of Au-NPs using the aqueous extract of *Falcaria vulgaris* leaves. *Applied Organometallic Chemistry*, 33(11), p.e5216.
132. Zhaleh, M., Zangeneh, A., Goorani, S., Seydi, N., Zangeneh, M.M., Tahvilian, R. and Pirabbasi, E., 2019. In vitro and in vivo evaluation of cytotoxicity, antioxidant, antibacterial, antifungal, and cutaneous wound healing properties of gold nanoparticles produced via a



green chemistry synthesis using *Gundelia tournefortii* L. as a capping and reducing agent. *Applied organometallic chemistry*, 33(9), p.e5015.

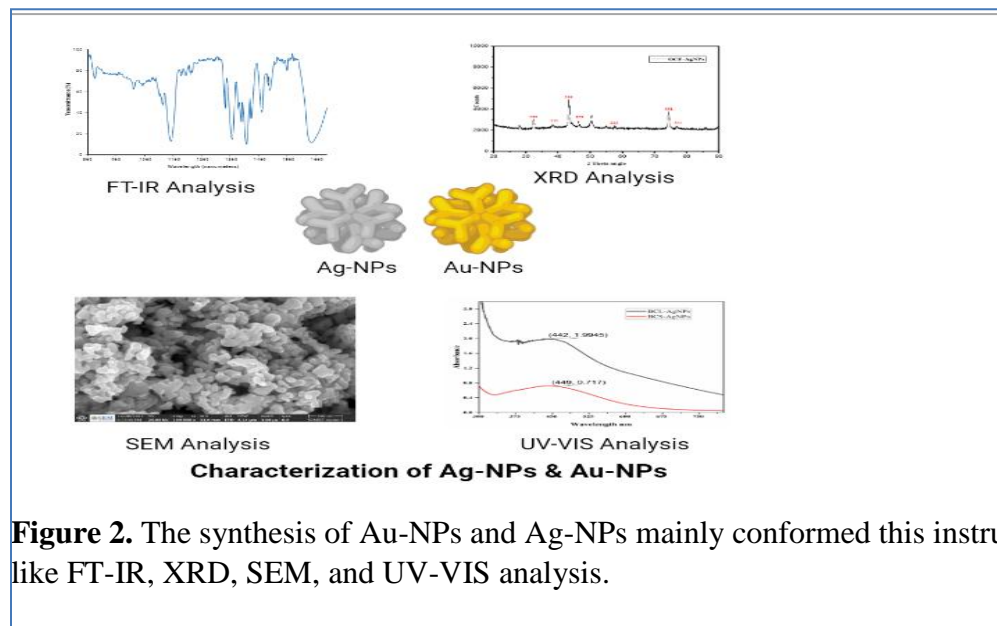
133. Das, T.K., Ganguly, S., Bhawal, P., Mondal, S. and Das, N.C., 2018. A facile green synthesis of silver nanoparticle-decorated hydroxyapatite for efficient catalytic activity towards 4-nitrophenol reduction. *Research on Chemical Intermediates*, 44, pp.1189-1208.
134. Hu, D., Gu, X., Si, W., Qin, W., Jiao, J. and Hao, Y., 2019. Biosynthesis of Silver nanoparticles using *Bauhinia acuminata* flower extract and their effect to promote osteogenesis of MSCs and improve meniscus injury healing. *Journal of Photochemistry and Photobiology B: Biology*, 197, p.111536.
135. Karuppaiya, P., Satheshkumar, E., Chao, W.T., Kao, L.Y., Chen, E.C.F. and Tsay, H.S., 2013. Anti-metastatic activity of biologically synthesized gold nanoparticles on human fibrosarcoma cell line HT-1080. *Colloids and Surfaces B: Biointerfaces*, 110, pp.163-170.

**Figure 1**



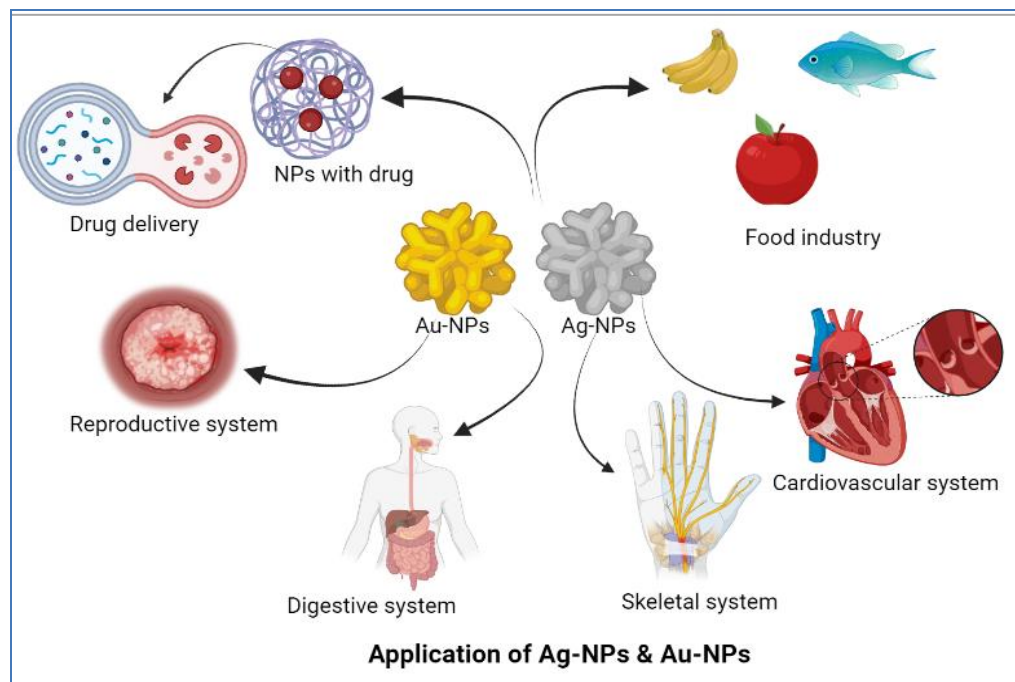
**Figure 1.** The synthesis of Au-NPs and Ag-NPs typically involves reducing metal salts in the presence of a reducing agent. The reducing agent helps to convert the metal ions into nanoparticles.

**Figure 2**



**Figure 2.** The synthesis of Au-NPs and Ag-NPs mainly conformed this instrumental analysis like FT-IR, XRD, SEM, and UV-VIS analysis.

**Figure 3**



**Figure3.** Au-NPs and Ag-NPs were used for many applications such as food industry, drug delivery, reproductive system, digestive system, skeletal system, cardiovascular system and many more.