B Pazopanib as a monotherapic anticancer drug

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ABSTRACT

Cancer is wide range of illnesses characterised by the expansion of abnormal cells with the ability to invade and damage healthy bodily tissue. Cancer is treated with a variety of chemotherapy or chemo medicines, Busulfan and the alkylating agent. Soft tissue sarcoma is uncommon kind which occurs in the tissues that encircle, link, and support other body structures. This includes the lining of your joints, blood vessels, muscle and blood vessels. The soft tissue sarcoma family includes 50 subtypes. Pazopanib is used to treat specific cancers (soft tissue sarcoma). Tyrosine kinase inhibitors are class of medications that includes pazopanib treatment for soft tissue sarcoma and its mode of action are the current areas of emphasis.

KEYWORDS

Pazopanib, Personalised medicine, pharmacokinetics, and soft tissue sarcoma, Angiogenesis

INTRODUCTION TO NEOPLASM

In the twenty-first century, cancer is predicted to rank as the largest cause of death, anticipated 29.5 million incident cases and 16.4 million fatalities by 2040, according to the (IARC) the International Agency for Research on Cancer, World Health Organization (2020). Globally, cancer claims more lives than TB, malaria, diabetes, and HIV/AIDS combined ^[1,2]. The major obstacles in oncology include cancer's molecular heterogeneity, an expensive, ineffective medications, and rising resistance to chemotherapy and radiation treatments ^[2,3]. In order to identify effective cancer treatment plans, more research is therefore essential ^[4].

Section A-Research paper

More and more studies have been conducted since the 1990s to support the notion that cancer stem cells exist^[5]. The main cause of the difficulty in treating and preventing tumours from returning is the involvement of cancer stem cells in the stability and maintenance of tumour heterogeneity^[5]. According to one theory, heterogeneity comes from several cell types, such as those cells that have stem cell-like properties ^[6]. There is a less evidences of its advent whether they are been actively produces or mutated in to stem cells, there is another theory which suggest that variety of tumour result from clonal evolution, and that mutant tumour cells can survive and grow ^[7]. A dominant mutant cell has the capacity to support tumour growth and resembles stem cells^[8]. The two viewpoints are not exclusive of one another but rather have internal relationships^[9]. Radiotherapy (RT) will be used to treat at least 60% of cancer patients ^[10]. Due to the rarity and variety treatment for STSs (soft tissue sarcomas) difficult^[11]. Surgery along with supplemental RT is the primary mode of therapy^[12]. DNAinteracting ionised intracellular compounds and reactive oxygen species are two ways that RT causes DNA alterations in cancer cells and healthy cells ^[13]. Numerous downstream cell death processes, including for instance, apoptosis, necrosis, and irreversible cell cycle arrest, are triggered Single-strand breaks (SSB) and double-strand breaks (DSB), two types of DNA damage. The tumour's innate radio resistance, the tumour a microenvironment's radiosensitivity, the surrounding normal tissues, however, limit the effectiveness of RT in the treatment of cancer^[14,15].

TYPES OF CANCER

Glioma

Up to 80% of brain cancers tumours of gliomas, which are the most prevalent main type of adult brain cancer ^[16]. Despite being the most common primary brain tumour, glioblastoma (GBM) is a kind of glioma that accounts for 57.3% of these tumours and has the worst prognosis: WHO grade IV ^[16,17]. There are two distinct categories for gliomas. First, roughly 90% of all GBMs are IDH wild-type tumours, or de novo primary GBMs, which are typically found in older patients (R62 years). Second, secondary GBM, which only accounts for 10% of cases and more frequently affects patients between the ages of 40 and 50, is the IDH mutant kind. Low-grade astrocytoma's give rise to IDH mutant tumours ^[17–19].

Liver

Two common digestive gland malignancies are liver cancer and pancreatic cancer ^[20]. The sole remaining option for pancreatic cancer patients and liver cancer to receive a radical cure is surgical resection ^[21]. A late or distant metastasis prevents the majority of patients from

having surgery ^[21]. Most patients will experience local or distant metastases even after major surgery, which will ultimately result in death ^[22].

Breast cancer

Breast cancer is one of the most prevalent malignant tumours that puts women's health in danger worldwide ^[23]. The breast cancer treatment still faces a number of difficulties, Despite the advancement of diagnostic and treatment methods such surgery, endocrine therapy, immunotherapy, and adjuvant chemoradiotherapy, certain problems still exist, such as simple metastasis and a high recurrence rate ^[24-27]. Therefore, it is crucial to monitor the activity of cancer cells while they are present and stop tumour metastasis through early detection and therapy. For the detection of various malignancies, a number of techniques are available, including Reverse transcription-polymerase chain reaction (RT-PCR), computed tomography (CT), magnetic resonance imaging (MRI), and immunohistochemistry analyses ^[28,29]. Although these techniques have the ability to accurately identify cancer in vitro, they require complex preprocessing when used at the biological level, which makes real-time in-vivo detection challenging. Optical imaging is a noninvasive method for identifying cancer in situ fluorescence in real time, as opposed to conventional detection procedures ^[30]. Currently, fluorescent probes that respond to enzymic tumour biomarkers with fluorescent "off-on" signals can be used to visualize cancer cells ^[31–33]. In fluorescence-guided surgery, these probes have developed into a potent biological tool ^[34–37].

Gastrointestinal cancer

According to GLOBOCAN 2018, there will be 9.6 million cancer-related fatalities and 18.1 fresh instances of cancer worldwide in 2018 ^[38]. 2018 was predicted to see over 1,000,000 new cases of stomach cancer, 783,000 fatalities, New cases of colorectal cancer totaling 1.8 million, resulting in an anticipated 881,000 fatalities. In all, gastrointestinal (GI) tract malignancies were responsible for nearly15% of new cases and 17% of cancer-related fatalities. The next-most common cancers are colorectal and gastric cancer common cancers in China, respectively, and the third and fifth most common causes of cancer mortality ^[39]. Compared to the US and the UK, with digestive tract malignancies accounting for 36.4% of all cancer-related fatalities, China has a lower cancer incidence than the US, but a 30%–40% higher cancer fatality rate ^[40]. The most efficient GI cancer therapies are surgical resection, radiation, and medication therapy. Due to late diagnosis, GI cancer has an extremely poor prognosis., despite advances in medication research. Immunotherapy, targeted therapy, and chemotherapy are the primary types of pharmacological therapy used for GI cancer. the status

of things right now suggests that in order to increase the number of drug clinical studies for GI cancer, we urgently need to promote more of them ^[41].

Soft tissue sarcoma

About 1%–2% of all malignancies are "soft tissue sarcomas" (STS), a rare cancer originating derived from mesenchymal connective tissue. Annually, 12750 new cases are identified in the USA, and STS causes 5270 fatalities ^[42]. In the 28 EU member states, 4.71 per 100,000 people were affected by STS individuals in Europe. There were reportedly 25851 new cases ^[43]. Traditional histology and molecular genetics are combined in the diagnosis and is based on the 2013 WHO classification of soft tissue tumour ^[44]. It can be difficult to diagnose mesenchymal tumour solely based on morphology and immunohistochemical staining, despite the fact that some sarcomas exhibit distinctive such as spindle cells, epithelioid or epithelial-like cells, myxoid tumour, round cells, and pleomorphic morphology ^[45]. Furthermore, a traditional histologic diagnosis frequently lacks a clear direction for anticancer therapy ^[46]. Cytogenetic PCR, targeted sequencing, and molecular genetic analysis, such Fluorescence in situ hybridization, karyotyping, and reverse transcription, are increasingly often used more biomarkers for diagnosis and treatment decision-making in the sarcoma diagnostic work-up ^[47].

TYPES OF SOFT TISSUE SARCOMA

General overview of sarcoma the two main neoplasm groups that make up the heterogeneous sarcoma tumour category are:

Bone sarcomas and soft tissue sarcomas

Soft tissue sarcomas are mesodermal in origin and typically start in the body's muscle, fat, fibrous tissue, blood vessels, or another supporting tissue. These sarcomas make up about1% of adult cancers and 7% of paediatric cancers. The most common varieties include among the cancers arising from fibrous tissue are fibrosarcoma and malignant fibrous histiocytoma., Leiomyosarcoma from smooth muscle, liposarcoma from fatty tissue, muscle-derived rhabdomyosarcoma and angiosarcoma, Blood and lymph vessel-related lymphangiosarcomas and Kaposi sarcoma, perivascular tissue-derived hemangiopericytoma, 0.2% of all new cancer diagnoses are bone sarcomas, which peak older adults (secondary sarcomas) and adolescents (initial sarcomas) linked to Irradiated bones and Paget disease). Ewing sarcoma and osteosarcoma are the two primary types of bone sarcomas. The group of Ewing sarcomas consists of a skin tumour, extraosseous Ewing tumour, PNETs, or peripheral neuroectodermal tumours neuroepitheliomas (the chest wall's PNETs). These malignancies originate from the same kind of stem cell ^[48].

Soft tissue and bone sarcomas in adults: -

The majority of uncommon malignancies fall under the category of sarcomas. There are two major groups: Bone and soft tissue sarcomas.

3.1 Soft tissue sarcomas are malignancies that can develop in many bodily parts. By the site or kind of tissue that is damaged, They are cancers of the mesenchymal (supporting) tissues. The most prevalent sarcoma, gastrointestinal stromal tumour, or GIST, damages the gastrointestinal tract's wall and is typically classified separately from other sarcomas. Adult soft tissue sarcomas account for more than 80% of sarcomas, with an incidence of 4 per 100,000 people annually in Europe.They are distinct from childhood soft tissue sarcomas., which frequently occur in rare paediatric cancers and have various characteristics,Guidelines and treatment procedures. For more information, Consult the SIOPE Strategic Plan and the European Society for Paediatric Oncology (SIOPE, https://www.siope.eu)^[49].

3.2 Sarcomas of the bone are the most common types of cancer. About 15% of sarcomas in Europe are these, which are less common than adult soft tissue sarcomas. The most prevalent kinds osteosarcoma and Ewing sarcoma, which are featured in this publication because the treatment strategies used are similar to those employed for adults. Teenagers and young adults are the most susceptible to these types. The most frequent adult bone sarcoma is chondrosarco Undifferentiated pleomorphic Sarcomas of the Bone. A few other bone sarcomas are chordomas, giant cell tumour of the bone, and undifferentiated pleomorphic sarcomas, The European Standards of Care for Children with Cancer also apply ^[50].



Figure1.Molecular mechanism of action of pazopanib.

DIAGNOSIS:

Due to the rarity of sarcomas, the wide variety of forms, and the frequently ambiguous symptoms, most primary care physicians only occasionally encounter a patient who has sarcoma. A benign diagnosis may also outnumber a sarcoma diagnosis by a factor of 100. Delays in referrals and diagnosis may occur from this.

Making the proper diagnosis of sarcomas requires the expertise of radiologists and pathologists who specialise in sarcomas, although they often work in a few numbers of centers. Non-expert surgical biopsies may result in issues, problems with following therapies, and even the spread of the tumour. A 2012 study ^[51] is cited. The ECCO expert panel emphasises that only sarcoma centres or paediatric cancer centres with experience treating sarcomas should be used for diagnosis. At second reading, it was discovered that more than 40% of initial histological diagnoses had been altered, possibly influencing different treatment options ^[52].

In conclusion, a patient who is not diagnosed at a sarcoma center may suffer serious repercussions, such as missing the possibility to receive a prompt diagnosis of a condition that may be treatable and avoiding more invasive surgery ^[53].

TREATMENT SOFT TISSUE SARCOMA INVOLVES:

SR NO	GENE	SNP	CANCER TYPE	THERAPY	TOXICITY	REFERENCE
1	SLC22A 16	rs714368 rs6907567 rs723685	ASTS	Doxorubicin	decreased the frequency of grade 3–4 AE (rs723685)	Seddon 2017
2	ABCB1	rs1128503	ASTS	Trabectedin	decreased risk of severe hepatic cytolysis	Maillard 2020 Maillard 2020
		rs2032582	ASTS	Trabectedin	decreased risk of overall hepatotoxicity	
3	ABCC2	rs717620 rs8187707 rs8187710	LPS	Trabectedin	irreversible hepatotoxicity	Laurenty 2013 Maillard 2020
		rs2273697	ASTS	Trabectedin	increased risk of hepatic cytolysis	Maillard 2020

Table 1: Available treatment for soft tissue sarcoma :

		rs17222723	ASTS	Trabectedin	decreased risk of hepatic cytolysis	
4	ABCC3	rs2072365	ASTS	Trabectedin	higher potential for severe cytolysis and total hepatotoxicity	Maillard 2020
5	ABCC4	rs9516519	ASTS	Trabectedin	overall hepatotoxicity risk is reduced	Maillard 2020
6	ABCG2	rs7699188	ASTS	Trabectedin	Hepatic cytolysis is more likely to occur.	Maillard 2020
7	СҮРЗА5	rs779746	ASTS	Trabectedin	grade 3/4 hepatic incident is more likely to occur.	Maillard 2020
8	ITGA	rs1126643	STS	Apatinib	spontaneous pneumothorax and surgical wound complications	Bao Q 2019

Abbreviations: LPS = liposarcoma; ASTS = advanced soft tissue sarcoma; AE = adverse event; SNP = single nucleotide polymorphism.

PAZOPANIB:

In order to treat advanced renal cell carcinoma, the medication pazopanib (Votrient®) is prescribed. Vascular endothelial growth factor receptors (VEGFR-1, VEGFR-2), and VEGFR-3), platelet-derived growth factor receptor and stem cell growth factor receptor (c-Kit) are the targets of this drug's tyrosine kinase inhibitor class i.e pazopanib as shown in figure 1 ^[59]. According to the most recent dosing recommendations, a starting dose of 800 mg once daily is indicated for treatment ^[60]. Because pazopanib is taken by up to 40–50% of patients with renal cancer, there have been worries about hepatotoxicity with this medication. in clinical trials had elevated blood transaminases and bilirubin ^[61–64]. the US Food and Drug Administration (FDA) mandated a black box alert in 2012 ^[65,66]. The anticancer and antiangiogenic efficacy of pazopanib in preclinical tests using mice xenograft models with multiple

myeloma cells models is reliant on concentration, requiring a steady-state plasma concentration of >40 mol/l (= 17.5 mg/L) ^[67,68]. Pazopanib's effectiveness in treating patients with metastatic RCC was connected pazopanib C trough of 15 mg/L in a phase I dose-escalating experiment, The patients received dosages ranging from 50 mg three times weekly to 2000 mg OD and 300-400 mg BID ^[69]. Patients received 800 mg or 300 mg BID, who showed a clinical response. Although MTD (maximum tolerable dose) was not reached, At the suggested dose of 800 mg OD, the exposure to pazopanib did not increase with predefined dose decreases in case of unacceptable toxicity. Active metabolites of pazopanib account for further 6% of all drug exposure ^[70].

Clinical response thresholds

Exposure-response relationships include

There are given clinical research on the relationship between pazopanib's exposureeffectiveness. pazopanib threshold C trough >20.5 mg/L was identified by Suttle et al as being associated showed a substantial increase in the median PFS in RCC patients ^[71]. Patients below this cutoff demonstrated similar efficacy to placebo. This threshold was independently confirmed by Verheijen et al ^[72], and it roughly corresponds to the results of the preclinical/early-phase trials. Although there were variations in reaction at the same threshold for STS patients, The difference was not statistically significant. This may be because there were fewer patients and a smaller impact size in STS patients as comparison to m RCC ^[88], so even though it was less durable, the same threshold may still be appropriate for STS patients. pazopanib trough levels have been linked to both survival and response rates (as determined by the RECIST criteria); 11 of the remaining 24 RCC patients out of 27 patients had an OR, but of the remaining 24 patients, none of the three individuals with a pazopanib C trough of 20.5 mg/L out of 27 RCC patients did ^[74].

Relation between toxicity and exposure

It has also been proven that exposure and toxicity are related ^[84,88,73,74], demonstrating that an increase in pazopanib C trough is linked to an increase in the frequency of adverse events ^[84,88]. In two investigations (n = 205), it was determined that individuals the highest levels of pazopanib C trough > 46 mg/L rate of adverse events (AEs), particularly those with hand-foot syndrome and hypertension (all grades) ^[43,84]. Recently, Noda et al (n = 27) determined a somewhat equivalent upper threshold of 50.3 mg/L for grade 3 toxicity. The results were most convincing for fatigue, anorexia, and hypertension.

CONCLUSION:

We advise setting a target exposure threshold of >20.5 mg/L for pazopanib C trough because multiple clinical trials have shown a correlation between pazopanib C trough >20.5 mg/L and a notable increase in median PFS. Pazopanib C trough values >46 mg/L patients report experiencing more toxicity.

A pazopanib safety and efficacy study

Due to their apparent inability to tolerate the treatment's negative effects or to get a significant benefit from it. Therefore, there is no agreed-upon standard of care, and there are no definitive rules for the to take care of patients with low PS. PFS (progression-free survival) and overall survival rates have significantly improved in first-line therapy trials for a RCC (OS). Tyrosine kinase inhibitors (TKIs) targeting 1-4 VEGFRs and, more recently, testing of either two immune checkpoint inhibitors (ICI) or a mix of one ICI and a TKI. Most of these trials, however, excluded a RCC ECOG PS 2 patients, Karnofsky PS 70% patients, and just 1included patient using ECOG PS 0-23. Therefore, there is a lack of information on the therapies' tolerance and effectiveness for a RCC patients with low PS. The TKI pazopanib targets the signalling pathways involved in the formation of tumours as well as VEGFR-1, -2, and -3. 10 The recommended treatment for an RCC at the time was sunitinib, a VEGFRtargeting TKI. was shown to have a shorter PFS than pazopanib in a significant phase III trial in patients with a RCC and PS 0-1^[86]. The same trial also showed that pazopanib had improved health-related quality of life (QoL) ratings and was better tolerated than sunitinib. Poor PS is seen in between 13% and 29% of all a RCC patients ^[87,88]. To close this knowledge gap and establish a higher standard of care for these patients, the Pazo2 study was developed. Pazo2's objective was to evaluate the drug's effectiveness and tolerability. The first-line use of Pazopanib therapy for a RCC patients with ECOG PS2.

CONCLUSION:

The long-standing a persistent need for effective novel treatments for these uncommon diseases has been addressed by pazopanib's successful clinical development as a treatment for advanced STS. However, based on formally documented clinical trial data, only a small proportion of the STS population's patients will benefit from treatment, and in many cases, the benefit's duration would be brief. Until yet, baseline clinicopathological factors that improve pazopanib benefit have not been identified in data from prospective pazopanib studies' subgroup analyses. Additionally, the scant Translational research that have looked at circulating or tumor-based biomarkers have not yet provided meaningful and reliable potential biomarkers. Numerous biomolecular profiling data points point to the presence of intrinsic biological subgroups within particular STS histocytes. Additionally, it has been

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shown that biological characteristics including enhanced chromosomal instability occur in a percentage of tumours across different Similar clinical signs are associated with STS subtypes ^[90–92]. An interesting direction for biomarker research is the evaluation of differing treatment results between such physiologically characterised pazopanib-treated cohorts' STS subgroups, improved discriminatory trial endpoints and assistance in the the identification and validation of potential imaging surrogate markers for survival could lead to the early diagnosis of clinical effects. The exact mechanism(s) by which When used in STS, pazopanib has an anticancer effect. changes across and within various STS subtypes are currently unknown. Furthermore, pazopanib resistance is frequently developed, even in individuals who initially showed a clear benefit from treatment. for pazopanib combination regimens, patient screening, and the development of novel combination regimens cytotoxic medications, more small molecule inhibitors, or new immunotherapeutic strategies, a deeper understanding understanding the pharmacological action mechanisms and primary and secondary drug resistance is important. To understand the therapeutic and resistance mechanisms of pazopanib in contexts of certain diseases, more preclinical and translational research is needed.

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REFERENCE:

1.Moten A, et al: Redefining global health priorities: **improving cancer care in developing settings.** J. Glob. Health 2014, 4 (1): 010304.

2.Bhuyan DJ, et al: Synergistic effects of Chinese herbal medicine and biological networks. Approaching Complex Diseases. Springer 2020, pp. 393–436.

3. Zhang Y, Liang Y, He, C: Anticancer activities and mechanisms of heat-clearing and detoxicating traditional Chinese herbal medicine. Chin. Med 2071, 12, 20

4.Jaye K, Li CG, Bhuyan DJ: **The complex interplay of gut microbiota with the five most common cancer types**: From carcinogenesis to therapeutics to prognoses. Critical Reviews in Oncology/ Hematology 2021 Sep 1,165:103429.

5.Zhu K, Xie V, Huang, S: **Epigenetic regulation of cancer stem cell and tumorigenesis.** Adv. Cancer Res 2020,148: 1–26. 6.Athanasiadis EI, Botthof JG, Andres H, Ferreira L, Lio P, Cvejic, A: **Single-cell RNA-sequencing uncovers transcriptional states and fate decisions in haematopoiesis.** Nat. Commun 2017, 8: 2045.

7.Makena MR, Ranjan A, Thirumala V, Reddy AP: Cancer stem cells: Road to therapeutic resistance and strategies to overcome resistance. Biochim. Biophys. Acta Mol. Basis Dis 2020, 1866 (4): 165339. <u>https://doi.org/10.1016/j.bbadis.2018.11.015</u>.
8.Duffey DC, Chen Z, Dong G, et al: Expression of a dominant-negative mutant

inhibitor-kappa alpha of nuclear factor-kappa B in human head and neck squamous cell carcinoma inhibits survival, proinflammatory cytokine expression, and tumor growth in vivo. Cancer Res 1999, 59 (14): 3468–3474

9.Xia P, Liu DH: **Cancer stem cell markers for liver cancer and pancreatic cancer.** Stem Cell Research. 2022 Feb 4:102701.

10. Chen HHW, Kuo MT: Improving radiotherapy in cancer treatment: Promises and challenges. Oncotarget 2017,8:62742–58. https://doi.org/10.18632/ oncotarget.18409.

11. Mangoni M, Sottili M, Salvatore G, Campanacci D, Scoccianti G, Beltrami G, et al: **soft tissue sarcomas: new opportunity of treatment with PARP inhibitors?** Radiol Medica 2019,124:282–9. <u>https://doi.org/10.1007/s11547-018-0877-4</u>.

12. Wong P, Houghton P, Kirsch DG, Finkelstein SE, Monjazeb AM, Xu-Welliver M, et al: **Combining targeted agents with modern radiotherapy in soft tissue sarcomas.** J Natl Cancer Inst 2014,106:16–8. https://doi.org/10.1093/jnci/ dju329.

13. Wang H, Mu X, He H, Zhang XD: **Cancer radiosensitizers**: Trends Pharmocol Sci 2018,39:24–48. <u>https://doi.org/10.1016/j.tips.2017.11.003</u>.

14. Tian J, Doi H, Saar M, Santos J, Li X, Peehl DM, et al: **Radioprotection and cell cycle arrest of intestinal epithelial cells by darinaparsin, a tumour radiosensitizer.** Int J Radiat Oncol Biol Phys 2013,87:1179–85. https://doi.org/10.1016/j.ijrobp.2013.08.051.

15. Bavoux M, Kamio Y, Vigneux-Foley E, Lafontaine J, Najyb O, Refet-Mollof E, Carrier JF, Gervais T, Wong P. X-ray on chip: **Quantifying therapeutic synergies between radiotherapy and anticancer drugs using soft tissue sarcoma tumour spheroids.** Radiotherapy and Oncology. 2021 Apr 1,157:175-81.

16. Ostrom QT, Cioffi G, Gittleman H, Patil N, Waite K, Kruchko C, and Barnholtz-Sloan, JS: CBTRUS statistical report: **primary brain and other central nervous system tumours diagnosed in the United States in 2012-2016.** Neuro. Oncol2019, 21: v1–v100.

17. Louis DN, Perry A, Reifenberger G, Von Deimling A, Figarella-Branger D, Cavenee, WK, Ohgaki H, Wiestler OD, Kleihues P, and Ellison DW: The 2016 world health

organization classification of tumours of the central nervous system: a summary. Acta Neuropathology 2016, 131, 803–820

18. Patterson J, Wongsurawat T, and Rodriguez A: A glioblastoma genomics primer for clinicians. Med. Res. March 2020, 8.

19.Rajabi A, Kayedi M, Rahimi S, Dashti F, Mirazimi SM, Homayoonfal M, Mahdian SM, Hamblin MR, Tamtaji OR, Afrasiabi A, Jafari A: **Non-coding RNAs and glioma: Focus on cancer stem cells.** Molecular Therapy-Oncolytics. 2022 Sep 17.

20. Siegel RL, Miller KD, Jemal A: Cancer statistics. CA Cancer J Clin 2020, 70(1): 7-30.

21.Ni J, Zhang L: Cancer cachexia: **definition**, **staging**, **and emerging treatments**. Cancer Manage. Res 2020, 12: 5597–5605

22. Xia P, Liu DH: Cancer stem cell markers for liver cancer and pancreatic cancer. Stem Cell Research. 2022 Feb 4:102701.

23.Meidanchi A: Mg(1-x) CuxFe2O4 superparamagnetic nanoparticles as nano radiosensitizer agents in radiotherapy of MCF-7 human breast cancer cells, Nanotechnology. 31 (2020) 325706.

24. Barua D, Gupta A, Gupta S: **Targeting the IRE1-XBP1 axis to overcome endocrine resistance in breast cancer: Opportunities and challenges**, Cancer Lett. 486 (2020) 29–37. 25.Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison JS, Clarke MF: **Prospective identification of tumorigenic breast cancer cells,** Proc. Natl. Acad. Sci. 100 (2003) 3983 LP – 3988.

26. Miller KD, Siegel RL, Lin CC, Mariotto AB, Kramer JL, Rowland JH, Stein KD, Alteri R, Jemal A: **Cancer treatment and survivorship statistics,** 2016, CA. Cancer J. Clin 2016, 66:271–289.

27. Partridge AH, Hughes ME, Warner ET, Ottesen RA, Wong YN, Edge SB, Theriault RL, Blayney DW, Niland JC, Winer EP, Weeks JC: **Tamimi RM, Subtype-Dependent Relationship Between Young Age at Diagnosis and Breast Cancer Survival,** J. Clin. Oncol 2016, 34 :3308–3314.

28. Eriksson L, Bergh J, Humphreys K, Wärnberg F, Törnberg S, Czene K: **Time from breast cancer diagnosis to therapeutic surgery and breast cancer prognosis**: A population-based cohort study, Int. J. Cancer 2018,143: 1093–1104.

29. Mikhaylov G, Klimpel D, Schaschke N, Mikac U, Vizovisek M, Fonovic M, Turk V, Turk B, Vasiljeva O: Selective Targeting of Tumor and Stromal Cells by a Nanocarrier System Displaying Lipidated Cathepsin B Inhibitor, Angew. Chemie Int. Ed 2014, 53: 10077–10081.

30. Pu Y, Leng Y, Wang D, Wang J, Foster NR, Chen J: **Recent progress in the green synthesis of rare-earth doped up conversion nanophosphors for optical bioimaging from cells to animals,** Chinese J. Chem. Eng. 2018, 26: 2206–2218.

31. Guo S, Fan J, Wang B, Xiao M, Li Y, Du J, Peng X: Highly Selective Red Emitting Fluorescent Probe for Imaging Cancer Cells in Situ by Targeting Pim1 Kinase, ACS Appl. Mater. Interfaces.

32. Gao M, Yu F, Ly C, Choo J, Chen L: Fluorescent chemical probes for accurate tumor diagnosis and targeting therapy, Chem. Soc. Rev. 2017,46: 2237–2271, (2018) ,[10]: 1499–1507.

33. Chen X, Lee KA, Ren X, Ryu JC, Kim G, Ryu JH, Lee WJ, Yoon J: Synthesis of a highly HOCl-selective fluorescent probe and its use for imaging HOCl in cells and organisms, Nat. Protoc 2016, 11 :1219–1228.

34. Dean KM, Palmer AE: Advances in fluorescence labeling strategies for dynamic cellular imaging, Nat. Chem. Biol 2014,10: 512–523.

35. Chen X, Wang F, Hyun JY, Wei T, Qiang J, Ren X, Shin I, Yoon J: Recent progress in the development of fluorescent, luminescent and colorimetric probes for detection of reactive oxygen and nitrogen species, Chem. Soc. Rev 2016, 45:2976–3016.

36. Wu L, Zeng W, Feng L, Hu Y, Sun Y, Yan Y, Chen HY, Ye D: An activatable ratio metric near-infrared fluorescent probe for hydrogen sulfide imaging in vivo, Sci. China Chem 2020, 63:741–750.

37. Xia W, Zhang S, Fan J, Li Y, Peng X: **Imaging and inhibiting cyclooxygenase-2 using aspirin-based fluorescent reporter for the treatment of breast cancer.** Sensors and Actuators B: Chemical. 2021 Feb 15,329:129217.

38. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A: Global cancer statistics
2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in
185 countries. CA A Cancer J Clin 2018 Nov,68(6):394e424.

39. Chen W, Zheng R, Baade PD, Zhang S, Zeng H: Cancer statistics in China, 2015. CA A Cancer J Clin 2006 Mar-Apr,66(2):115e32.

40. Feng RM, Zong YN, Cao SM, Xu RH. Current cancer situation in China: good or bad news from the 2018 Global Cancer Statistics? Canc Commun 2019 Apr 29,39(1):22

41. Ren C, Xu RH: **The drug treatment research of gastrointestinal cancer in China**. European Journal of Surgical Oncology. 2020 Oct 1,46(10): 3-6.

42. Siegel RL, Miller KD, Jemal A: Cancer statistics, CA Cancer J Clin 2019,69:7–34.

43. Gatta G, Capocaccia R, Botta L, et al: Burden and centralized treatment in Europe of rare tumour: results of RARECAREnet-a population-based study. Lancet Oncol 2017,18:1022–39.

44. Fletcher CDM BJ, Hogendoorn PCW, Mertens F: Who classification of tumors of soft tissue and bone. 4th ed. Lyon: IARC Press 2013.

45. Schaefer IM, Cote GM, Hornick JL, et al: Genetics, and genomics. J Clin Oncol 2018,36:101–10.

46. Pollack SM, Ingham M, Spraker MB, et al: **Emerging targeted and immune-based therapies in sarcoma.** J Clin Oncol 2018,36:125–35.

47.Chen HW, Chen TW: Genomic-guided precision therapy for soft tissue sarcoma. ESMO open. 2020 Jan 1,5(2):000626.

48.Nanni C, Fanti S: **FDG-PET and PET/CT for evaluating soft tissue sarcomas.** PET clinics. 2010 Jul 1,5(3):341-7.

49. Vassal G, Schrappe M, Ladenstein R, Pritchard-Jones K, Arnold F, Basset L,

et al: The SIOPE strategic plan: European cancer plan for children and

adolescents. J. Cancer Policy 2016,8: 17–32, http://dx.doi.org/10.1016/j.jcpo.2016.03.007.

50.Andritsch E, Beishon M, Bielack S, Bonvalot S, Casali P, Crul M, Delgado-Bolton R, Donati DM, Douis H, Haas R, Hogendoorn P: **ECCO essential requirements for quality cancer care: soft tissue sarcoma in adults and bone sarcoma.** A critical review. Critical reviews in oncology/hematology. 2017 Feb 1:110:94-105.

51.Ray-Coquard I, Montesco MC, Coindre JM, Dei Tos AP, Lurkin A, Ranchère-Vince D, et al: Sarcoma concordance between initial diagnosis and centralized expert review in a population-based study within three European regions. Ann. Oncol 2012, 23 (9) :2442–2449, http://dx.doi.org/10.1093/annonc/mdr610

52.Beishon M: When in doubt, ask an expert. Cancer World 2013 (May/June) http://www.cancerworld.org/pdf/2385 pagina 30 34 Systems & Services.pdf.

53.Andritsch E, Beishon M, Bielack S, Bonvalot S, Casali P, Crul M, Delgado-Bolton R, Donati DM, Douis H, Haas R, Hogendoorn P: **ECCO essential requirements for quality cancer care: soft tissue sarcoma in adults and bone sarcoma.** Critical reviews in oncology/hematology. 2017 Feb 1; 110:94-105.

54. Seddon B, Strauss SJ, Whelan J, et al: Gemcitabine and docetaxel versus doxorubicin as first-line treatment in previously untreated advanced unresectable or metastatic softtissue sarcomas (Giddies): a randomized controlled phase 3 trial 11Lancet Oncol. 18 (10) (2017) 1397–1410, <u>https://doi.org/10.1016/S1470-2045</u> (17)30622-8. Epub 2017 Sep 4. PMID: 28882536; PMCID: PMC5622179 55. Maillard M, Chevreau C, Le Louedec F, et al: **Pharmacogenetic study of trabectedininduced severe hepatotoxicity in patients with advanced soft tissue sarcoma,** Cancers (Basel) 2020, 12 (12) 3647, https://doi.org/10.3390/ cancers12123647. PMID: 33291741; PMCID: PMC7761985

56. Laurenty AP, Thomas F, Chatelut E, et al: **Irreversible hepatotoxicity after** administration of trabectedin to a pleiomorphic sarcoma patient with a rare ABCC2 polymorphism: a case report, Pharmacogenomics 14 (12) (2013) 1389–1396, https://doi.org/10.2217/pgs.13.124. PMID: 24024892

57. Bao Q, Hu Y, Wen J, et al: VEGFR2 and ITGA polymorphisms as novel predictors of therapeutic response and toxicities for pediatric and young adult sarcoma

undergoing anti-angiogenic therapy, Ann. Oncol2019, 30 (suppl_5).

58.Elisabetta G, Anna B, Adriano P, Andrea CD, Guido S, Ilaria P, Andrea B, Lorenzo A, Serena P. Pharmacogenomics of soft tissue sarcomas: **new horizons to understand efficacy and toxicity**. Cancer Treatment and Research Communications. 2022 Feb 1:100528.

59. Harrison M, Lang J, Pazopanib for the treatment of patients with advanced renal cell carcinoma, Clin. Med. Insights Oncol 2010, 4: 95–105.

60.GSK, Prescribing Information for Votrient® (Pazopanib), Medication Guide, 2015.

61. Kapadia S, Hapani S, Choueiri TK, et al: **Risk of liver toxicity with the angiogenesis** inhibitor pazopanib in cancer patients, Acta Oncol. (Stockh.) 2013,52 :1202–1212.

62. Shah RR, Morgan Roth J, Shah DR, **Hepatotoxicity of tyrosine kinase inhibitors**: clinical and regulatory perspectives, Drug Saf 2013, 7: 491–503.

63. Shah C, Saiyed MM: Hematological Toxicities Associated with Pazopanib Use in Cancer Patients: A Meta-Analysis, Value Health, 2015.

64. Gupta S, Spies PE: The prospects of pazopanib in advanced renal cell carcinoma, Ther Adv Urol 2013, 5: 223–232.

65. Guidelines for Bioanalytical Method Validation, FDA: Preclinical and early-phase clinical thresholds for response, 2013.

66.Toh YL, Pang YY, Shwe M, Kanesvaran R, Toh CK, Chan A: **Ho HK. HPLC-MS/MS** coupled with equilibrium dialysis method for quantification of free drug concentration of pazopanib in plasma. Heliyon. 2020 Apr 1;6(4): e03813.

67. Kumar R, Knick VB, Rudolph SK, et al: **Pharmacokinetic pharmacodynamic** correlation from mouse to human with pazopanib, a multikinase angiogenesis inhibitor with potent antitumor and antiangiogenic activity. Mol Cancer Ther. 2007;6(7):2012-2021. <u>https://doi.org/10.1158/1535-7163.MCT-07-0193</u>.

68. Podar K, Tonon G, Sattler M, et al: **The small-molecule VEGF receptor inhibitor pazopanib (GW786034B) targets both tumor and endothelial cells in multiple myeloma.** Proc Natl Acad Sci U S A. 2006;103 (51):19478-19483. https://doi.org/10.1073/pnas.0609329103 Epub 2006 Dec 12.

69. Hurwitz HI, Dowlati A, Saini S, et al: Phase I trial of pazopanib in patients with advanced cancer. Clin Cancer Res. 2009;15(12):4220- 4227. https://doi.org/10.1158/1078-0432.CCR-08-2740 Epub 009 Jun 9

70. US Food and Drug Administration: **Center for Drug Evaluation and Research. Pazopanib clinical pharmacology and biopharmaceutics review(s).** 2008, December 19 [Available from: https://www. accessdata.fda.gov/drugsatfda_docs/nda/2009/022465s000_ ClinPharmR.pdf

71. Suttle AB, Ball HA, Molimard M, et al: **Relationships between pazopanib exposure** and clinical safety and efficacy in patients with advanced renal cell carcinoma. Br J Cancer. 2014;111(10):1909-1916. https://doi.org/10.1038/bjc.2014.503 Epub Oct 28

72. Verheijen RB, Swart LE, Beijnen JH, Schellens JHM, Huitema ADR, Steeghs N: **Exposure-survival analyses of pazopanib in renal cell carcinoma and soft tissue sarcoma patients**: opportunities for dose optimization. Cancer Chemother Pharmacal. 2017;80(6):1171-1178. https://doi.org/10.1007/s00280-017-3463-x Epub 2017 Oct 19.

73. Lin Y, Ball HA, Suttle B, et al: **Relationship between plasma pazopanib concentration and incidence of adverse events in renal cell carcinoma.** J Clin Oncol. 2011;29(7_suppl):345–345.

74. Noda S, Yoshida T, Hira D, et al: Exploratory investigation of target Pazopanib concentration range for patients with renal cell carcinoma. Clin Genitourin Cancer. 2018;7(18):30734-30731.

75. Westerdijk K, Desar IM, Steeghs N, van der Graaf WT, van Erp NP: Dutch Pharmacology and Oncology Group (DPOG). **Imatinib, sunitinib and pazopanib**: from flat- fixed dosing towards a pharmacokinetically guided personalized dose. British journal of clinical pharmacology. 2020 Feb;86(2):258-73.

76. Motzer RJ, Hutson TE, Tomczak P, et al: Sunitinib versus interferon alfa in

metastatic renal-cell carcinoma. N Engl J Med. 2007;356(2):115–124. doi:10.1056/NEJMoa065044.

77. Sternberg CN, Davis ID, Mardiak J, et al: **Pazopanib in locally advanced or metastatic renal cell carcinoma: results of a randomized phase III trial.** J Clin Oncol.2010;28(6):1061–1068. doi:10.1200/jco.2009.23.9764. 78. Choueiri TK, Halabi S, Sanford BL, et al: **Cabozantinib versus sunitinib as initial targeted therapy for patients with metastatic renal cell carcinoma of poor or intermediate risk**: the alliance A031203 CABOSUN trial. J Clin Oncol. 2017;35(6):591–597. doi:10.1200/jco.2016.70.7398

79. Motzer RJ, Nosov D, Eisen T, et al: **Tivozanib versus sorafenib as initial targeted therapy for patients with metastatic renal cell carcinoma**: results from a phase III trial. J Clin Oncol. 2013;31(30):3791–3799. doi:10.1200/jco.2012.47.4940.

80. Motzer RJ, Tannir NM, McDermott DF, et al: **Nivolumab plus ipilimumab versus sunitinib in advanced renal-cell carcinoma**. N Engl J Med. 2018;378(14):1277–1290. doi:10.1056/NEJMoa1712126. 81. Rini BI, Plimack ER, Stus V, et al: Pembrolizumab plus Axitinib versus Sunitinib for advanced renal-cell carcinoma. N Engl J Med. 2019;380(12):1116–1127. doi:10.1056/NEJMoa1816714.

82. Motzer RJ, Penkov K, Haanen J, et al: Avelumab plus Axitinib versus Sunitinib for advanced renal-cell carcinoma. N Engl J Med. 2019;380(12):1103–1115. doi:10.1056/NEJMoa1816047.

83. Motzer R, Alekseev B, Rha SY, et al: Lenvatinib plus Pembrolizumab or Everolimus for advanced renal cell carcinoma. N Engl J Med. 2021;384(14):1289–1300.doi:10.1056/NEJMoa2035716.

84. Choueiri TK, Powles T, Burotto M, et al: Nivolumab plus Cabozantinib versus Sunitinib for advanced renal-cell carcinoma. N Engl J Med. 2021;384(9):829–841.doi:10.1056/NEJMoa2026982.

85. Miyamoto S, Kakutani S, Sato Y, Hanashi A, Kinoshita Y, Ishikawa A: Drug review: **pazopanib.** Jpn J Clin Oncol. 2018,48(6):503–513. doi:10.1093/jjco/hyy053.

86. Escudier B, Porta C, Bono P, et al: Randomized, controlled, double-blind, cross-over trial assessing treatment preference for Pazopanib Versus Sunitinib in patients with metastatic renal cell carcinoma: Pisces study. Journal of Clinical Oncology.2014;32(14):1412–1418. doi:10.1200/jco.2013.50.8267.

87. Gore ME, Szczylik C, Porta C, et al: **Safety and efficacy of sunitinib for metastatic renal-cell carcinoma:** an expanded-access trial. Lancet Oncol. 2009,10(8):757–763. doi:10.1016/s1470-2045(09)70162-7.

88. Motzer RJ, Mazumdar M, Bacik J, Berg W, Amsterdam A, Ferrara J: Survival and prognostic stratification of 670 patients with advanced renal cell carcinoma. J Clin Oncol. 19 99;17(8):2530–2540.

89. Zarkar A, Pirrie S, Stubbs C, Hodgkins AM, Farrugia D, Fife K, MacDonald-Smith C, Vasudev N, Porfiri E, Pazo2 Investigators: A Study of Pazopanib Safety and Efficacy in

Patients with Advanced Clear Cell Renal Cell Carcinoma and ECOG Performance Status 2 (Pazo2): An Open label, Multicentre, Single Arm, Phase II Trial. Clinical Genitourinary Cancer. 2022 Oct 1;20(5):473-81.

90. Chibon F. et al: Validated prediction of clinical outcome in sarcomas and multiple types of cancer on the basis of a gene expression signature related to genome complexity. Nat. Med 2010. 16, 781–787.

91. Guo X. et al: **Clinically relevant molecular subtypes in leiomyosarcoma.** Clin. Cancer Res 2015,21:3501–3511.

92. Abeshouse A. et al: Comprehensive and integrated genomic characterization of adult soft tissue sarcomas. Cell 2017,171:950–965.e28.

93. Lee AT, Jones RL, Huang PH: **Pazopanib in advanced soft tissue sarcomas.** Signal transduction and targeted therapy. 2019 May 17;4(1):1-0.

94. Ranieri G, Mammì M, Di Paola ED, Russo E, Gallelli L, Citraro R, Gadaleta CD, Marech I, Ammendola M, De Sarro G: **Pazopanib a tyrosine kinase inhibitor with strong anti-angiogenetic activity:** a new treatment for metastatic soft tissue sarcoma. Critical reviews in oncology/hematology. 2014 Feb 1;89(2):322-9.

95. Cowey CL, Hutson TE, Figlin R: **Pazopanib in the treatment of renal cell carcinoma. Clinical Investigation.** 2011 Jan;1(1):75-85.