

TRANSFERSOMES BASED DERMAL GEL OF BACLOFEN FOR MANAGEMENT OF CHRONIC PAIN

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Abstract

Baclofen's ability to relax skeletal muscles has been linked to its anti-inflammatory and stressrelieving effects. The two current baclofen treatments, oral tablets and intrathecal injection, both come with a number of undesirable systemic side effects. Developing a topical baclofen formulation to reduce the drug's systemic adverse effects was the primary goal of this research. This is accomplished by increasing the medication's penetration rate into the epidermis by increasing the time it spends in contact with the skin. The fast degradation of baclofen during first-pass metabolism accounts for its short half-life (0.4-0.8 h) and limited bioavailability (20%). This study aimed to determine the efficacy of elastic transfersome containing formulations in enhancing the skin's bioavailability of baclofen and decreasing its hepatic metabolism. It may dissolve in mild acids and bases, such as 1% acetic acid and 0.1 M NaOH. As expected, the normal melting point of 208 degrees Celsius was found to be within the range of 202 to 205 degrees Celsius. Using varied quantities of Soya-phosphatidylcholine, tween 80, span 20, and sodium deoxycholate, nine alternative formulations were developed and evaluated for particle size. The concentration of baclofen in each phase was estimated using a specific formula, and the result was used to calculate the partition coefficient. Partition coefficient data suggest that baclofen is likely to be a lipophilic compound. In order to evaluate the NPs' size and stability, we measured their particle size and zeta potential and compared the results to those of freshly made samples. Although the NPs grew in size significantly, the values of their zeta potential remained positive despite a significant decrease caused by the generation of counter charges in solution. Aggregation of NPs causes particles to grow in size, and this decrease in zeta potential may help explain why. The extrudability of gel compositions was also evaluated, with findings favouring carbopol gels.

Keywords: Baclofen, degradation, gels, Chronic pain, Partition coefficient and Transfersomes

1.0 Introduction

A pharmaceutical product must be carried in the body safely, employing secure practises, formulations, technologies, and systems to achieve the intended therapeutic effect. Quantity and duration of medication presence are common areas of focus in pharmacokinetics, including facilitating systemic pharmacokinetics or scientific site-targeting within the body. Improved therapeutic effectiveness and patient tolerance may be achieved by the use of drug delivery systems that alter the release profile, absorption, distribution, and elimination of the medication. Diffusion, degradation, swelling, and affinity all play roles in drug release from containers. Inhalation, topical application to the skin, and transmucosal routes (which include the nasal, buccal/sublingual, vaginal, ophthalmic, and rectal routes) are the most often used non-invasive modes of administration. Traditional dose forms, including extended-release ones, fall short of these requirements. Several novel drug delivery mechanisms have made significant efforts to improve the subpar performance of the already accepted methods of medication administration. Recent studies have revealed lipid vesicles' use in genetic engineering, diagnosis, membrane biology, and immunology.

The administration of drugs through vesicles is the current standard. The vesicular delivery approach reduces pharmaceutical toxicity without adding new adverse effects by transporting medications directly to the site of infection. Vesicular drug delivery improves bioavailability, decreasing treatment expenses, especially for drugs that aren't readily soluble. Substances that are hydrophilic and lipophilic may work together. The vesicular system employs cutting-edge drug delivery mechanisms, including liposomes, niosomes, sphinosomes, transferosomes, and pharmacosomes. A vesicular system is a very organised assembly of one or more concentric lipid bilayers that is formed when certain amphiphilic building materials are combined with water.

Multiple hydrophilic materials may be used to produce vesicles. In 1965, Bingham was the first to determine that these containers had a biological basis; he named them Bingham bodies. Site-specific, stimuli-responsive, and slowly degrading drug carriers are feasible to design. The end result should be less medication waste and deterioration, fewer adverse drug reactions, and more drugs available at the location of the illness. Enclosing a drug in vesicular structures is thought to extend its systemic half-life and lower its potential toxicity if selective absorption is achievable. Controlled delivery platforms such as lipid vesicles have been established by experimental modeling of biomembranes. Because of the limited ability of most drugs to reach cells, standard chemotherapy cannot treat infections that have already spread within the body. This problem may be solved by using vesicular medicine administration techniques. Vesicular medication administration offers the potential to decrease toxicity and increase treatment efficacy by delivering the drug directly to the site of infection for selective absorption. Improves drug bioavailability, particularly for poorly soluble medicines. Lipophilic and hydrophilic molecules might coexist. Functions are a slow-release mechanism, preventing treatments from metabolizing too quickly. These vesicular systems may include drug carriers and passively loading, externally triggered (e.g., magnetically sensitive) carriers; nonetheless, they may suffer from poor drug loading efficiency and leakage during in vivo preparation, preservation, and delivery.

1.1 Liposomes

For pharmaceutical medications to be effective, they must enter the body unharmed. This method of administering medication is known as "drug delivery". Scientific site-targeting inside the body or systemic pharmacokinetics are two examples, but in any case, the amount and duration of drug presence are often addressed. Medication administration tactics alter the drug's release profile, absorption, distribution, and elimination to improve product efficacy, patient comfort, and compliance. Several parameters may affect drug release, including diffusion, degradation, swelling, and affinity-based processes. Transmucosal (through the nose, buccal/sublingually, vaginally, topically, or rectally), topical (via the skin), peroral (by the mouth), and inhalation are the most frequent modes of delivery. Standard dose forms, including sustainedrelease versions, can meet none of these conditions. Although there is not yet an ideal technique for administering medicines, several innovative drug delivery methods have been used to get close. Multiple fields use lipid vesicles, including those of genetic engineering, immunology, membrane biology, and diagnosis. Drugs are often administered through vesicles. Vesicular delivery method reduces drug toxicity and eliminates unwanted side effects by delivering medication directly to the site of an illness. Especially for insufficiently soluble medications, vesicular drug delivery reduces treatment costs by enhancing drug bioavailability. It is possible for hydrophilic and lipophilic substances to coexist. Liposomes, niosomes, sphingosomes, transferases, and pharmacosomes are all examples of the vesicular system, which is one of the most important drug delivery techniques. As per Jadhav et al. some amphiphilic building blocks,

when put in contact with water, form vesicles composed of one or more concentric lipid bilayers. When phospholipids self-assemble in an aqueous solution, they form closed bilayered structures called liposomes. Liposomes may be tiny (unilamellar or multilamellar) vesicles. In order for closed bilayer structures to develop spontaneously, an energy source is often required, such as physical agitation, sonication, etc. Since a lipid bilayer membrane surrounds an aqueous core, liposomes may encapsulate both lipid- and water-soluble molecules. Liposomes' drawbacks include their high manufacturing costs, susceptibility to drug/molecule leakage and fusion, phospholipid oxidation and hydrolysis, rapid degradation, limited solubility, and poor stability. Small unilamellar vesicles (SUVs), large unilamellar vesicles (LUVs), multilamellar vesicles (MLVs), and multivesicular vesicles (MVVs) are the four varieties of liposomes that their size and number of bilayers may identify. In their unilamellar form, liposomes are composed of a monophospholipid bilayer; in their multilamellar form, they take on an onion-like shape. Several unilamellar vesicles fuse in bigger liposomes to produce a more complex multi-lamellar structure. Liposome size is directly related to the efficiency with which hydrophilic molecules may be encapsulated, however, this efficiency diminishes as the bilayer thickness becomes larger. Vesicle size has a significant impact on the half-life of circulating liposomes. The size and quantity of bilayers determine the total amount of medication stored. When liposomes deliver medications, the vesicles utilised are on the order of 50-150 nm in size. The interaction of liposomes with cellular membranes has been explained in a number of different ways. These include selective (receptor-mediated) and nonspecific endocytosis, local fusion (adhesion), phagocytosis, and absorption. The interactions between liposomes and cells are affected by a number of variables.

1.3 Types of liposomes

There are many different kinds of liposomes, each with its own chemical composition and set of uses. These include the more common "traditional" liposomes, as well as charged liposomes, stealth-stable liposomes, actively targeted liposomes, stimulusresponsive liposomes, and bubble liposomes.

Table 1: Examples of new vesicular drug delivery systems					
Vesicular system	Description	Application			
Aquasomes	Nanocrystalline carbon	Molecular shielding and			
	ceramic core and	targeted targeting.			
	cellobiose covering make				
	up this three-layered self-				
	assembling composite.				
Cryptosomes	Lipid vesicles with a	Ligand mediated drug			
	suitable polyoxyethylene	Targeting			
	phosphotidyl				
	ethanolamine derivative				
	and a PC surface layer.				
Discosomes	Niosomes dispersed in	Ligand mediated drug			
	polyoxyethylene cetyl ether (a non-ionic	Targeting			
	surfactant) solutions.				
Emulsomes	Microscopic lipid assemblies with a polar	Parenteral delivery of poorly			
	nucleus were used to water soluble drugs				
	create nanoscale lipid particles (bioadhesives				
	nanoemulsion).				
Enzymosomes	A tiny bioenvironment is	Targeted delivery to tumor			
	created by covalently	Cells			
	immobilizing or coupling				
	enzymes to the surface of				
	liposomal structures.				
Ethosomes	Ethosomes are lipid "soft Targeted delivery to d				
	malleable vesicles" skin layer cells				
	composed of ethanol,				

2.0 Materials and Methods

2.1 Chemical Requirements

Various materials were obtained from standard suppliers. The analytical grade chemical reagents were used in all experiments.

S. No.	Chemical used	Company
1	Methanol	S.D. fine chemicals Ltd. (India)
2	Tween 80	S.D. fine chemicals Ltd. (India)
3	Sodium deoaxycholate	S.D. fine chemicals Ltd. (India)
4	Baclofen	Sun Pharma
5	Span 80	S.D. fine chemicals Ltd. (India)
6	Sephadex G-50	SIGMA-ALDRICH
7	n-Octanol	CDH
8	Chloroform	S.D. fine chemicals Ltd. (India)
9	Triethanolamine	S.D. fine chemicals Ltd. (India)
10	Carbopol 934	Lubrizol (India)
11	NaCl	HiMedia
12	Soya lecithin	Agro Solvent Products Pvt. Ltd

2.2 Bioactive and Excipients Profile

Physicochemical Properties of baclofen

Table 4.2 Physicochemica	al Properties	
IUPAC name	4-amino-3-(4-chlorophenyl)butanoic acid Computed by Lexichem TK 2.7.0 (PubChem release 2021.05.07)	
Chemical Formula	C10H12CINO2	
Chemical structure		
Molecular weight	213.66 g/mol	
Appearance	white powder	
Solubility	It is soluble in dilute acids and alkalis (1% acetic acid and 0.1 M NaOH)	

Tween 80

The two main ingredients in polysorbate 80 are polyethoxylated sorbitan and oleic acid. These hydrophilic groups are ethylene oxide polymers called polyethers or polyoxyethylene groups. In polysorbate nomenclature, the numerical designation after polysorbate denotes the lipophilic group, which in this instance is oleic acid. The well-known antiarrhythmic amiodarone is produced using polysorbate 80 as an emulsifier to stabilize aqueous formulations of pharmaceuticals for parenteral administration. Some influenza vaccinations in Europe and Canada use it as an excipient. There are 25 grams of polysorbate 80 in each vial of flu vaccination. Many vaccinations sold in the United States include polysorbate 80. Middlebrook 7H9 broth is used to cultivate Mycobacterium tuberculosis. It is also utilized in the Middlebrook 7H9 broth culture of Mycobacterium tuberculosis.

Soyalecithin

Lecithins are a kind of surfactant having lubricating and emulsifying abilities. They are harmless and well tolerated since humans can metabolize them (see inositol), while the kidneys must eliminate other emulsifiers. It is a dispersion agent, stabiliser, carrier for choline enrichment, and moisturizing agent in the pharmaceutical sector. It may be infused intravenously as a therapeutic agent to make lipid infusions. Studies have shown soy-derived lecithin to increase HDL ("good cholesterol") levels and decrease animal blood cholesterol and triglyceride levels. However, there is mounting evidence that the microbes in your gut convert lecithin into trimethylamine-Noxide (TMAO), which is absorbed in the intestine and may have a role in developing atherosclerosis and the onset of heart attacks.

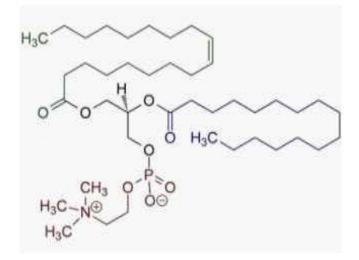


Figure 1: Chemical Structure of Soya Lecithin

Instrument	Model	Manufacturer
Rotary Evaporator	GJJ-2	Heidolph, Germany
Digital weighing balance	AB265-S/ FACT	Mettler Toledo, Switzerland
Vortex Shaker	Spinix	Tarsons products Pvt. Ltd,
		India
Zeta Size Analyzer	Delsa Nano C	Beckman Coulter Pvt. Ltd.
Magnetic stirrer	PT 400	Perfit, India
Centrifuge	R-24C	Remi Instruments division
		Maharashtra, India
Franz Diffusion cell	4G- 01- 00-09-05	Prefit, India
Melting Point Apparatus	DB- 31354	Decibels Instruments,
		India

Drug Procurement (Baclofen)

The bioactive sample was obtained from Sun Pharma Ltd, India as a Gift sample.

Physical Appearance

The physical appearance of bioactive like color and the odor was analyzed by visual observation.

Purity and Identification

Various studies have been performed to check the purity and identification of the drug.

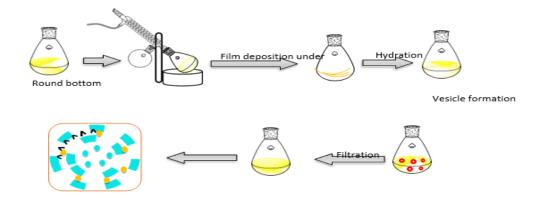
Solubility Analysis

The drug's solubility was tested using purified water. The experiment used three conical flasks, introducing 10mg of bioactive and 5 mL of distilled water. After that, you put the flasks in a shaking incubator for awhole day. The drug concentration may be calculated by filtering the fluid with Whatman filter paper. The solubility of a drug, or its ability todissolve in a given solvent, is an important aspect in attaining the concentration of the drug in the systemic circulation essential for the intended pharmacological action. The issue with bioavailability may stem from poor solubility or permeability. As a result, most solutions of varying concentrations are impacted by the solubility issue. Thus, improved therapeutic advancements are in requirement as syntheticscience advances. The mechanism of solute-solvent interaction describes the behaviour of homogeneous liquid phases with a fixed ratio of solute to solvent. The selection of which technique to utilize to boost solubility is based on the qualities of the medication, the retention site, and the property of the required measurement structure.

Formulation and Optimization of Baclofen Loaded Transferosome Gel

Formulation by thin-film hydration method

The suspension was made using a thin-film dispersion-hydration technique. In a 20 mL roundbottomed vial, the medicine, soya phosphatidylcholine, and surfactants were dissolved in dehydrated alcohol. Alcohol was evaporated in a rotary evaporator to create a consistent, ultrathin lipid layer. The film is going to be hydrated with 20 ml of pH 6.5 phosphate buffer at 20 °C for an hour to create coarse dispersions. The dispersions were sonicated in a bath sonicator for 9 minutes at 400 watts. After that, we'll squeeze them down further by passing them through 0.45 mm-pore- diameter Nylon-66 membranes.



Preparation of Transfersomes

The process of preparing transfersomes occurs in two phases. Sonicated vesicles are extruded via a membrane filter once the thin film has been prepared to ensure consistency. A phospholipid and surfactant solution was heated above the lipid transition temperature in a rotary evaporator to remove the volatile organic solvent (chloroform-methanol). The leftover solvent was swept up after an overnight soak. The deposited lipid films were treated with buffer (6.4) for one hour at the propertemperature while spinning at 60 revolutions per minute. The size of the resulting vesicles increased after two hours at room temperature. Tiny vesicles were made by probe sonicating LMVs for 30 minutes at ambient temperature. Homogenization was accomplished by manually injecting a membrane filter into the sonicated vesicles.

Preparation of Topical Transfersome Gel Formulation:

The prepared transfersomes were mixed in with the carbopol-934 (1%) with gel base at a 1:1 ratio. After soaking the powder in water for 30 minutes, the 1 percent carbopol-934 gel basis was prepared by continually spinning the solution. At a concentration of 1%, Carbopol-934 gel base gels nicely and has a passable consistency.

Zeta Potential Determination

Electrophoresis and measuring the electrical conductivity of emulsion samples were used to calculate the vesicle zeta potential. Software based on the Helmholtz- smoluchowski equation was used to determine the zeta potential of the vesicles from their electrophoretic mobility.

Compatibility Study

The stability of a formulation depends on the compatibility of the activecomponent with the excipients. The bioavailability and stability of a drug maybe affected by a number of physical (or chemical) interactions, making their identification crucial. Differential scanning is a fast and accurate way to test the compatibility and interaction between a medicine and an excipient was performed by the calorimetry method.

Characterization of Transfersomes

Particle Size Determination

Scanning electron microscopy (SEM) was used to take pictures of and analyze the size of Acarbose Transfersomes. The Hitachi vacuum evaporator was used to plate the acarbose transfersomes with gold. SEM analysis and imaging of coated samples was performed using a Hitachi s-3000H instrument.

Size Distribution of Transfersomes

To determine the size distribution of transfersomes, we used a laser particle counter (Spectrex brand).

Entrapment Efficiency

The efficiency of the traps was easily determined. Detergents may be used to damage the transferosome membranes. After being incubated at 37 degrees Celsius for 1.5 hours with 1 millilitre of 0.1% Triton X- 100 (Triton X- 100 dissolved in phosphate buffer), the transferosome membrane was disrupted and the confined material was released. After filtering the sample over a 0.25 m Millipore membrane, the acarbose content in the filtratewas measured at 425 nm. Acarbose concentration was calculated using the standard curve.

Transfersomes Gel Evaluations Determination of viscosity

The viscosities of the gels were determined using a Brookfield Viscometer (RVTP type).Spindle speed of 20 RPM, RV-7, after spinning the spindle for 5 minutes with 100 g of the gel in a beaker, and reading was taken.

Extrudability

In this case, an empirical test that measures the force required to remove the substance from the tube might be useful. We measured how much gel squeezedout of the tube's tip under pressure to determine how well each formulation extruded through the tube's nasal tip, which had an aperture of 5mm. It was determined how well the formulation extruded, and the results were compiled.

4.8 Stability Study

The transfersome gel formulations were randomly assigned to one of three categories. The transfersome gel composition was divided into three groups andkept in aluminium collapsible tubes.

a. Room temperature
b. 37± 5°C
c. 4-5°C

It took three months to store the transfersome gel formulation. Over the courseof three months, the drug concentration in monthly samples was calculated. They were evaluated for quality and structure after three months.

2.8.1Physical evaluation

Physical criteria included things like product appearance, product kind, pH, viscosity, leak, phase separation, and extrudability.

Chemical evaluation

Drug content was assessed by sampling the formulation from the four corners of the tube. One gramme was taken from the combined samples and put throughanalysis. The dosage concentration was determined per protocol.

Drug content

500 milligrammes of gel were added to 100 millilitres of methanol, given a good stir for 30 minutes, and then left alone for an hour. The drug concentration in the solution was then determined using spectrophotometry at 239 nm after it had been filtered.

Stiffness

The gel formulation of AF-TG that is now on the market was used to apply around 100 mg to the skin using a pointed finger. We saw evidence of resistance to dissemination inour qualitative analysis. Based on how it felt, it was assigned a rating of,, or (14 most stiff, 14 medium, and 14 least stiff).

2.0 Results & Discussions

Preformulation studies

Various formulation parameters were evaluated, such as solubility, melting point, and partition coefficient.

Purity and Identification studies of Baclofen

The Baclofen was visually observed and was found to be white powder, practically odorless as mention in **Fig. 1**.

Melting Point

The purity of the bioactive was determined by measuring its melting point. Researchers determined that the melting point of Baclofen is between 202 and 205 degrees Celsius. It can be shown in Table 5.1 that a meets the required temperature of 208° C. The crystallinestructure of the medication is shown by the full melting of the bioactive across a small temperature range.

Melting Point	Standard	Test
	208° C	202° C- 205° C

Table 5.1 Determination of Melting Point

Baclofen solubility study

Baclofen has a low solubility in water and is fully insoluble in the common organic solvents ethanol, methanol, and acetone. Table 01 shows that it dissolves in dilute solutions of acetic acid (1%) and sodium hydroxide (0.1 M).

S.no	Solvent	Solubility mg/ml
1	0.1M NAOH	46.2±0.12
2	1% acetic acid	34.2±0.47
3	Acetone	0.9±1.02
4	Ethanol	4.2±0.29
5	Phosphate buffer 7.4	16.2±0.82
6.	Methanol	3.4±0.64

Table-4.2. Baclofen Solubility data

Measurements of polydispersity index, particle size, and zeta potential as part of a physicochemical characterization

The consistency and precision with which the formula is dispensed depend heavily on the particle size and size distribution. The ultimate dimension was impacted by both polymer concentration and organic phase composition. All of the different preparations had particle sizes that ranged from 123 nm to 340 nm. It decreased proportionally as the quantity went up. The organic phase's composition also impacted particle size, as medication solubility varied with organic solvent type and concentration. Particle size was found to be lowest in formulas comprising methanol and acetone in a ratio of (3:1), and biggest in formulae containing methanol and acetone in a ratio of

(1:3), both at the same ERL concentration. This might be because, as demonstrated in Table 2, Baclofen ismore soluble in methanol than in acetone. The PDI values for all of the samples were within a respectable range (0.271–0.532), indicating that the NPs were uniform and of a reasonable size distribution and that no NP aggregates were present. If the PDI is more than 0.7, then the distribution is not homogeneous. As the zeta potential value increases, the electrostatic repulsion between particles grows, which inhibits aggregation and allowsgreat separation between particles and from surfaces. The zeta potentials of the samples were quite high, spanning +28.9 mV to +47.6 mV. Since it is known that a zeta potential of 30 mV indicates perfect electrostatic stability, this finding reflected the validity of the formulations. The positive environmental charges are due to the free amino group in ERL.Particle size, particle distribution index, and zeta potential are all included.

S.no	Formula code	Particle size	Zeta potential (mV)	PDI
1	Baclo F1	206±8.4	+33.3±1.4	0.248±0.01
2	Baclo F2	340±5.3	+47.6±5.4	0.532±0.15
3	Baclo F3	181±1.9	+32.6±2.78	0.416±0.04
4	Baclo F4	148±2.29	+28.9±1.3	0.453±0.03
5	Baclo F5	289±11.82	+36.8±0.44	0.420±0.05
6.	Baclo F6	133±1.64	+33.0±1.17	0.229±0.002
7	Baclo F7	123±0.69	+42.3±0.73	0.271±0.005
8	Baclo F8	129±1.18	+37.8±0.41	0.418±0.01
9	Baclo F9	136±1.64	+39.5±1.43	0.278±0.006

Table-4.3. zeta potential, Particle size, and PDI

Viscosity measurement

Viscosity measurement is essential in gel production and application since it determines how the product will behave in terms of form, texture, and fluidity. Viscosities of 261.811,3554.412.34, and 4209316.47 were recorded for Formulas F3, F6, and F9, and for baclofen gel, respectively. The NPs gel was much thicker and stickier than the baclofen gel after being mixed with the ERL polymer. The presence of ERL raises viscosity because the intermolecular attractions of its long molecular chains increases flow resistance. As the percentage of ERL grew, so did the viscosity values.

S.no	Formula code	Viscosity c.P
1	Baclo F1	18156±12.6
2	Baclo F2	20175±8.6
3	Baclo F3	26188±11
4	Baclo F4	19123±10.32
5	Baclo F5	25244±13.89
6.	Baclo F6	35544±12.34
7	Baclo F7	23473±15.65
8	Baclo F8	36524±14.23
9	Baclo F9	42093±16.47

Table	4.4:	Viscosity	of gel
		, 1960916	01 <u>5</u> 01

Partition Coefficient

The partition coefficient of a chemical is defined as the equilibrium concentration ratio of the compound between two immiscible phases. The solubility difference between n- octanol and water, as well as the solubility difference between the component and the two solvents, are both reflected in these coefficients. Based on the assumed baclofen concentration at both time points, the partition coefficient was calculated. Baclofen has alipophilic polarity, as seen by the partition coefficient data.

Reported Log P	Observed Log P
1.27	1.21

Table 4.5.: Partition coefficient of Baclofen

Stability study

Once a month, the selected formulas were visually inspected for colour and consistency. There were no blatant changes to the appearance. The size and stability of the NPs were determined by comparing the measured particle size and zeta potential to those of freshly synthesized samples. The zeta potential values remained positive but decreased dramatically (p < 0.001) when counter charges were generated in solution, while the particle size of NPs increased significantly (p < 0.001). Aggregation of NPs produced the bigger particles, and the decrease in zeta potential accounted for the resulting rise in particle size. Table 04, as well as Figures 05 and 06, present the findings.

S.no	Formula code	Particle size (nm)	Zeta potential (mV)
1	Baclo F1	206±8.4	+33.3±1.4
2	Baclo F2	340±5.3	+47.6±5.4
3	Baclo F3	181±1.9	+32.6±2.78
4	Baclo F4	148±2.29	+28.9±1.3
5	Baclo F5	289±11.82	+36.8±0.44

Table-4.6. Stability studies (0 day)

6.	Baclo F6	133±1.64	+33.0±1.17
7	Baclo F7	123±0.69	+42.3±0.73
8	Baclo F8	129±1.18	+37.8±0.41
9	Baclo F9	136±1.64	+39.5±1.43

Table-4.7.	Stability studies (30 th day)
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S.no	Formula code	Particle size (nm)	Zeta potential (mV)	
1	Baclo F1	207.5±5.4	+32.8±1.2	
2	Baclo F2	343.5±4.6	+44.2±3.7	
3	Baclo F3	182.5±1.5	+31.2±1.8	
4	Baclo F4	152±2.1	+26.8±1.1	
5	Baclo F5	291.3±10.2	+34.9±0.74	
6.	Baclo F6	136±1.4	+31.7±1.70	
7	Baclo F7	124.2±1.2	+41.6±1.8	
8	Baclo F8	130.5±1.8	+36.2±1.21	
9	Baclo F9	137.6±1.4	+38.2±1.43	

Table-4.8. Stability studies (60 th d	lay)
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S.no	Formula code	Particle size (nm)	Zeta potential (mV)
1	Baclo F1	211.1±2.4	+30.5±2.2
2	Baclo F2	346.5±3.8	+41.8±2.4

3	Baclo F3	184.5±4.2	+30.2±1.4
4	Baclo F4	156±3.8	+24.3±1.6
5	Baclo F5	294.3±7.2	+33.2±0.24
6.	Baclo F6	138.5±1.05	+29.7±1.10
7	Baclo F7	126.9±1.6	+39.8±1.08
8	Baclo F8	133.7±1.4	+34.5±1.35
9	Baclo F9	139.8±1.8	+36.5±1.21

Extrudability

When tested, carbopol gels showed exceptional extrudability when compared to other gel compositions. It is shown in **Table-4.9**.

S.no	Formula code	Particle size (nm)
1	Baclo F1	+++
2	Baclo F2	+++
3	Baclo F3	+++
4	Baclo F4	+++
5	Baclo F5	+++
6.	Baclo F6	+++
7	Baclo F7	+++
8	Baclo F8	+++

9	Baclo F9	+++

+++Excellent

Characterization of Transferosome Gel

The results of analyses of Baclofen's acidity, viscosity, separability, brittleness, and active ingredient concentration are provided. These concentrations are well under the permissible limits for topical use. The average viscosity of baclofen is around 271.27, according to a number of different tests. Spreadability values of 11.23 g.cm/s for F2 indicate that they may be readily spared on the skin's surface under mild stress.

From these data, it may be concluded that the baclofen transferosomes exhibited more usual behaviours. Formulation-4 is the best formulation in terms of drug entrapment. Reduced dosage, fewer administrations per treatment period, and more patient compliance all resulted from the baclofen transfersome diffusion research. Therefore, Formulation 4 of baclofen transfersomes was an improved and more effective version previous formulations.

Conclusion

Initial investigations indicate that baclofen is a white crystalline material. Weak acids and bases (0.1 M NaOH and 1% acetic acid) can dissolve it. The melting point was between 202 and 205 degrees Celsius, which is close enough to the nominal 208 degrees Celsius to be considered acceptable. Different amounts of Soya- phosphatidylcholine, span 20, tween 80, and sodium deoxycholate were used to create a total of nine different formulations, which were then tested for particle size. Formula F4 was chosen as the best formulation for calculating viscosity. After determining the optimal batch of transferosomes, they were combined into a gel basis and tested for pH, viscosity measurement, and drug content. The partition coefficient was computed after estimating the baclofen concentration in each phase. Based on the results of the partition coefficient calculation, it seems that baclofen is lipophilic.

The stability and size of the NPs were evaluated in comparison to freshly created samples by measuring their particle size and zeta potential. The zeta potential values of the NPs remained positive but declined as a result of the formation of counter charges in solution, leading to a

substantial increase in the particle size of the NPs. This decrease in zeta potential may explain the increased particle size due to NP aggregation. The method was also used to evaluate the extrudability of gel formulations, where it was discovered that carbopol gels exhibited maximum extrudability.

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7.0 Author's Contribution

The manuscript was carried out, written, and approved in collaboration with all authors.

8.0 Conflict of Interest

The authors declared no conflicts of interest.

References

- 1. Nanotechnology in Sustainable Agriculture. CRC Press, 2021.
- Wen, Hong, Huijeong Jung, and Xuhong Li. "Drug delivery approaches in addressing clinical pharmacology-related issues: opportunities andchallenges." The AAPS journal 17 (2015): 1327-1340.
- Liu, D., M-A. Shahbazi, L. M. Bimbo, J. Hirvonen, and H. A. Santos. "Biocompatibility of porous silicon for biomedical applications." In Porous silicon for biomedical applications, pp. 129-181. Woodhead Publishing, 2014.
- 4. Jadhav SM, Morey P, Karpe MM, Kadam V. Novel vesicular system: an overview. Journal of applied pharmaceutical science. 2012 Jan 30(Issue):193- 202.
- Kumar, G. P., Gangarao, B., Kotha, N. S., & Lova, R. (2011). Isolation and evaluation of tamarind seed polysaccharide being used as a polymer in pharmaceutical dosage forms. Research Journal of Pharmaceutical, Biological and Chemical Sciences, 2(2), 274-290.
- Yingchoncharoen, Phatsapong, Danuta S. Kalinowski, and Des R. Richardson. "Lipidbased drug delivery systems in cancer therapy: what is available and what is yet to come." Pharmacological reviews 68, no. 3 (2016): 701-787.

- Sharma Vijay, K., D. Mishra, A. Sharma, and B. Srivastava. "Liposomes: present prospective and future challenges." International journal of current pharmaceutical review & research 1, no. 2 (2010): 6-16.
- Anwekar, Himanshu, Sitasharan Patel, and A. K. Singhai. "Liposome-as drug carriers." International journal of pharmacy & life sciences 2, no. 7 (2011).
- Rai, A., G. Alam, A. P. Singh, and N. K. Verma. "Niosomes: An approach to current drug delivery-A Review." International Journal of Advances in Pharmaceutics 6, no. 2 (2017): 41-48.
- 10. Pradhan, Bhupendra, Narendra Kumar, Suman Saha, and Amit Roy. "Liposome: method of preparation, advantages, evaluation and its application." Journal of applied pharmaceutical research 3, no. 3 (2015): 01-08.
- 11. Goyal, Tanu, Ashwani Singh Rawat, and Meenakshi Chauhan. "Pharmacosomes: Opening new doors for drug delivery." Int. J. Pharm. Pharm. Sci 4 (2012): 25-29.
- 12. Kurhe, Snehal Arjun, Kedar Bavaskar, and Ashish Jain. "Transferosome: A Vesicular Transdermal Delivery System for Enhanced Drug Permeation." Research Journal of Pharmaceutical Dosage Forms and Technology 14, no. 3 (2022): 206-210.
- Ashok, K., A. Rajendra Kumar, S. Nama, B. Brahmaiah, P. K. Desu, and C. B. Rao.
 "Sphingosomes: a novel vesicular drug delivery system." International Journal of Pharmaceutical Research and Bio-Science 2, no. 2 (2013): 305-312.
- 14. Zaid Alkilani, Ahlam, Maelíosa TC McCrudden, and Ryan F. Donnelly. "Transdermal drug delivery: innovative pharmaceutical developments based on disruption of the barrier properties of the stratum corneum." Pharmaceutics 7, no.4 (2015): 438-470.
- Ashok, K., A. Rajendra Kumar, S. Nama, B. Brahmaiah, P. K. Desu, and C. B. Rao.
 "Sphingosomes: a novel vesicular drug delivery system." International Journal of Pharmaceutical Research and Bio-Science 2, no. 2 (2013): 305-312.
- 16. Pastore, Michael N., Yogeshvar N. Kalia, Michael Horstmann, and Michael S. Roberts."Transdermal patches: history, development and pharmacology." British journal of pharmacology 172, no. 9 (2015): 2179-2209.
- Cherukuri, Suneetha, Uma Rajeswari Batchu, Kiranmai Mandava, Vidhyullatha Cherukuri, and Koteswara Rao Ganapuram. "Formulation and evaluation of transdermal drug delivery of topiramate." International journal of pharmaceuticalinvestigation 7, no. 1 (2017): 10.

- Singh, Inderjeet, and Andrew Phillip Morris. "Performance of transdermal therapeutic systems: Effects of biological factors." International journal of pharmaceutical investigation 1, no. 1 (2011): 4.
- 19. Sengar, Vaishali, Kiran Jyoti, Upendra Kumar Jain, Om Prakash Katare, Ramesh Chandra, and Jitender Madan. "Lipid nanoparticles for topical and transdermal delivery of pharmaceuticals and cosmeceuticals: A glorious victory." In Lipid Nanocarriers for Drug Targeting, pp. 413-436. William Andrew Publishing,2018.
- 20. Chandrashekar, N. S., and RH Shobha Rani. "Physicochemical andpharmacokinetic parameters in drug selection and loading for transdermal drug delivery." Indian journal of pharmaceutical sciences 70, no. 1 (2008): 94.
- 21. Jeong, Woo Yeup, Mina Kwon, Hye Eun Choi, and Ki Su Kim. "Recent advances in transdermal drug delivery systems: A review." Biomaterials research 25 (2021): 1-15.
- 22. Kumar R, Sushant PC, Bhatia R. Advancements in Fluorimetric Detection of Bio-Markers.
- 23. Kumar A, Ali R, Kumar R, Chawla PA. Exploring the potential of neuronutraceuticals as alternatives in the prevention of Alzheimer's disease. Pharmaspire. 2022 Jan;14:47-58.
- 24. Billowria K, Ali R, Rangra NK, Kumar R, Chawla PA. Bioactive Flavonoids: A Comprehensive Review on Pharmacokinetics and Analytical Aspects. Critical Reviews in Analytical Chemistry. 2022 Jul 23:1-5.