

EVALUATION OF DRUG DELIVERY SYSTEM – ION GATE CHANNELS

Anil kumar Boda, Dr.Shweta Shriwas, Dr. Rakesh Patel

research scholar department of pharmaceutical sciences, Dr.A.P.J.Abdul kalam university indore Dewas Bypass road, Arandia, indore, MP, 452010, India.

Associate professor, school of pharmacy, Dr. A.P.J.Abdul kalam University Indore Dewas Bypass road, Arandia, indore, MP,452010, India.

Professor & Principal, School of Pharmacy, Dr. A. P. J. Abdul Kalam University Indore Dewas Bypass Road, Arandia, Indore MP- 452010, India.

anil.pharma1988@gmail.com

ABSTRACT

Ion-channels are proteins which sit in the membrane of every cell in the body and control the flow of positively charged ions such as sodium and potassium into and out of the cell. Ion channels are thought to be in one of two stochastic states, i.e. open or closed, according to the classical model. However, the discovery of sub conductance' states in a variety of ion channels, including numerous potassium (K+) channels, casts doubt on this theory. Using the fact that heteromeric Kir4.1/Kir5.1 potassium channels exhibit long-lived sub-conductance states and an ortholog of Kir5.1 from Xenopus tropicalis causes a major change in the frequency and duration of these sub-states, we want to analyse these sub-conductance states in this study. A mathematical model will be used to retrace the path of single-channel currents. We will use the threshold-crossing approach, amplitude histograms, and HMM (Hidden Markov Model) analysis to assess sub-conductance states in all types of channels. As a first step, recordings of single-channel events are idealised into closed and open dwells, then dwell times are fitted to the data using Clampfit 9.2 and HJCFIT software, respectively. QuB analysis software will be used to ensure the unambiguous detection of brief sublevel occurrences and the comparison of sublevel durations.

Keywords: Ion-channels, Xenopus, Heteromeric, stochastic states.

1.INTRODUCTION

Ion-channels have been always related with drug discovery process. Their types, primarily recognized as Na⁺, K⁺, Ca²⁺, Cl⁻, have been basically associated with neuronal processes. Therefore, drugs targeted to them influence all organs or systems related with neuronal activity: the central nervous system (CNS), the peripheral nervous system, and the cardiovascular system. Within the CNS, basic indications of drugs are: sleep disorders, anxiety, epilepsy, pain, etc. This is because a variety of CNS diseases are linked to different etiologies, and in vivo screening has been used to discover many new medications since it mimics the state of sickness in the human brain. Anticonvulsants like lamotrigine, a well-known and relatively new medication, tend to act primarily on Na+ and Ca2+ channels.Ion-channel subtypes have recently been discovered, providing the foundation for drug discovery processes that target specific channel subtypes.[1]

ION-CHANNELS AND RELEVANT DRUG SCREENING APPROACHES

An electric potential across the membrane is created as a result of the outward movement of positively charged ions, causing the cell's interior to become negative in relation to the exterior. After a certain amount of time, the cell's negative charge builds up and resists the flow of K+ out of the cell. The resting membrane potential of the cell is set between -30 and -90 mV, depending on the cell type, as a result of the K+ concentration gradient and the presence of open K+ channels. Individual electrochemical gradients control the path taken by permeant ions in every case. Under normal circumstances, Na+ and Ca2+ flow into the cell and exert a depolarizing impact, whereas K+ flows out of the cell and Cl- flows into the cell, both exerting a repolarizing or hyperpolarizing influence. Ion channels, both selective and nonselective, are involved in numerous physiological processes in cells. These include setting up the resting membrane potential and propagating action potentials in excitable cells as well as muscle contraction, release of neurotransmitters, hormone secretion, signal transduction, cell volume regulation, and cell proliferation. Ion channels play an important role in all of these physiological processes. Furthermore, many disorders of the neurological system, cardiovascular system, skeletal muscle, kidney, and endocrine system are caused by abnormalities in Ion-channels (channelopathies). This confirmation of ion-channels is critical in the medical arena, where scientists not only

attempt to comprehend their complicated roles in both healthy and disease states, but also to identify and create medications to influence their function.[2]

2.MATERIALS AND METHODS

EXPERIMENTAL

PREFORMULATION STUDY

Organoleptic Properties and Description

The organoleptic properties of tramadol hydrochloride like colour, odour, and appearance were performed.

Melting Point Determination

Capillary fusion was used to determine the melting point of the medication. In a capillary tube, a little amount of tramadol hydrochloride was injected. A digital melting point equipment was used to record the temperature at which the medication melted in the capillary tube. This was done three times and the average was recorded. Diametric scanning calorimetry also confirmed the melting point.

Ultraviolet Absorption Maxima (λmax)

Using a 100 ml volumetric flask, correctly weighed 100 mg tramadol hydrochloride was put into distilled water and phosphate buffer pH 6.8 for this purpose. Dissolved the medication in distilled water and phosphate buffer pH 6.8 to 100 ml, respectively. To get the correct concentration, this solution was diluted using the appropriate solvent. A UV spectrophotometer was used to record the UV spectrum between 200 and 400 nm. This information was used to establish the solvent's optimum absorption wavelength (max).

FT-IR Spectroscopy

The FT-IR analysis of the tramadol hydrochloride was carried out for qualitative compound identification. The pure drug and drug with resins were mixed with potassium bromide to prepare a pellet. The pellet was mounted in IR compartment then scanned over a wavelength of 4000 cm-1 -400 cm-1 . 6.1.5 Calibration curve for tramadol hydrochloride *Eur. Chem. Bull.* **2023**,12(Special issue 8), 6089-6101 6091

Calibration curve for tramadol hydrochloride in distilled water

An accurate 100 mg dose of tramadol hydrochloride was accurately weighed and put into an empty 100 ml volumetric flask. The volume was increased to 100 ml by dissolving it in distilled water. This is a ready-to-go solution: From a standard stock solution, a series of tramadol hydrochloride solutions ranging from 10 to 100 g/ml were created. The absorbance of all solutions was measured against blank at λ max by UV spectrophotometer at 271 nm.

Calibration curve for tramadol hydrochloride in phosphate buffer of pH 6.8

Accurately measured 27.2 mg of potassium dihydrogen phosphate was dissolved in 250 ml of distilled water and stirred; the volume was then adjusted to 1 L using water and mixed thoroughly. Tramadol hydrochloride was carefully weighed and placed into a 100 ml volumetric flask, which was then shaken. pH 6.8 buffer solution was used to dilute it to 100 ml, and it was kept at that concentration. Tramadol hydrochloride solution varying in concentration from 10 to 100 g/ml was made from a stock solution of the drug. A UV spectrophotometer set to 271nm assessed the absorbance of each solution.

Solubility study

The solubility of tramadol hydrochloride was determined in different solvents like distilled water, 0.1 N HCl, ethanol and at different pH solutions. The solubility study was carried out by adding excess amount of the drug in 10 ml of each solvent in screw capped glass vials and shaken for 12 hours at room temperature. The solution was filtered and filtrate was diluted for suitable concentrations and quantified by UV spectrophotometer.

SELECTION AND CHARACTERIZATION OF RESIN

The resins were chosen based on the nature of the medicine and the formulation needs. Because the medication tramadol hydrochloride used in this study is basic and has an amine functional group, a catIon-channel resin would be the best Ion-channel resin to use in the complexing process. When it comes to a BCS Class I medicine that disintegrates too quickly, a sulphonic

EVALUATION OF DRUG DELIVERY SYSTEM - ION GATE CHANNELS

acid functional moiety-based Ion-channel resin, such as that found in Indion224, Indion244, Indion254 or Indion284, would be ideal.

IN VITRO DRUG RELEASE FROM ION-CHANNEL RESONATES

In vitro dissolution study was carried out in triplicate for resinates equivalent to 100 mg of tramadol hydrochloride, by using the USP paddle apparatus (Electrolab TDT 06L). At predetermined intervals 5 ml aliquots were withdrawn and replaced with the same volume of fresh dissolution medium. The collected aliquots were filtered through whatman filter paper no.41 and amount of drug released was analyzed by UV-vis spectrophotometer at 271 nm following suitable dilutions. [3-10]

3.RESULTS AND DISCUSSION

PREFORMULATION STUDY:

Organoleptic Properties

The sample of tramadol hydrochloride was observed as per the method described in experimental section and was found. These attributes were found to be in line with those described in EP.

Melting point of tramadol hydrochloride

A capillary technique was used to determine the melting point of tramadol hydrochloride. The temperature at which the medication melted in the capillary tube was recorded in a digital melting point instrument. The procedure was repeated three times and the mean temperature was 182.0±0.60C. Conformity with the standard text was achieved. The DSC investigation revealed the melting point to be 184.790C. Figure 4.1 depicts the thermogram results.

Ultraviolet Absorption Maxima scan (λmax)

The UV absorption spectra were observed as presented in Figure 7.2-3. The λ max of the tramadol hydrochloride was observed and recorded to be 271.00nm in distilled water as well as in the phosphate buffer pH 6.8.andit comply with the requirement of the standard texts.

FT-IR Spectroscopy

The FT-IR spectrum of tramadol hydrochloride was recorded and the major functional groups are observed as in table 4.3.

Calibration curve for tramadol hydrochloride distilled water

Tramadol hydrochloride standard calibration curves were created by graphing absorbance vs. concentration in distilled water and phosphate buffer of pH 6.8 medium. Results were averaged and analysed using a basic linear regression model.

Solubility study

The solubility of tramadol hydrochloride was determined in different solvents. The drug solution mixtures were filtered diluted with distilled water for a suitable concentrations and the solubility was determined spectrophotometrically. The results obtained are presented in table 4.4.

Dissolution medium volume necessary for the complete dissolution of the drug can be expressed as a Dose/Solubility Ratio (D/S). These findings show that tramadol hydrochloride is available in the dissolution media at 100mg in a D/S ratio of 0.00146 at pH6.8, 0.1 N HCl is found to be 0.00155, and water is found to be 0.00154. However, if the medication is supplied using a controlled release formulation, especially by means of complexing and encapsulating the molecule utilising Ion-channel resinates, it would deliver the drug at zero order release and boost bioavailability.

Selection and characterization of resins

Visual inspection and microscopy were used to determine the appearance and particle size of the Ion-channel resins. The swelling time and the moisture content were also determined... Indion 244, Indion254 resin particles were discovered to have a particle size of 0.15mm, which was in agreement with the literature. Both Indion 224 and 284 had particle sizes between 0.21 and 1.2 millimetres, as stated in the literature. The resin's moisture level was measured and found to be between 2% and 9%, which was within the acceptable range.

Preparation of drug resin complexes

Effect of resin activation on drug complexation

The resinates were pretreated with 1M HCl and 1M NaOH and the percent drug loading of the drug onto resin.

The percent drug complexation in table 7.7 shows that the inactivated resins have a lower drug loading. However, the percentage of drug complexation was shown to rise in the following sequence of pretreatment HCl, NaOH, and Acid+Base as indicated.. It is clear from the data that the activation of resin was required to produce the maximal drug complexation with resins. Because the Ion-surface channel's charge may be responsible for loading the resins with drugs. Ionic form changes are sometimes required when resin does not contain the desired counterions to be converted from one form to another. Na+ catIon-channel resins are the most common variety of strongly acidic catIon-channel resins. In most cases, the H+ ion is used to transform them. Acids and alkalis can be used to soak the resins and then washed to neutralise the elute until the resins are neutral. So the resins needed to be purified and activated before they could be used in further research.

The equilibrium profiles of drug loading onto the Ion-channel resins

In order to determine how long it takes for the Ion-channel resins to reach equilibrium and begin loading tramadol hydrochloride, the equilibrium profiles of the resins were investigated. Particle size and degree of crosslinking of the drug determine the binding surface area necessary for the drug to bind to Ion-channel resins, as shown in equilibrium profiles of the drug loading on to different resins. Because of their higher particle sizes, indion 224 and indion 284 have less surface area available for binding. There were two quick equilibrium profiles in the range of 10-15 minutes for indion 244 and indion 254. For the single batch process, the equilibrium investigated profiles showed that the equilibrium was almost reached in the first 30 minutes for all of the Ion-channel resins. Double and triple batch loading techniques were also tested, and from the figures 4.7 and 4.8 it is obvious that an additional 13-19 percent of the drug was loaded when employing double batch loading. Due to the saturation of resin materials with drug molecules, the loading of the medication was ineffective in the triple batch process. In order to conduct additional research, equilibrium profiles of the single batch and double batch processes were used.

It is important to maximise loading efficiency while minimising the size of dosage forms and the properties of drug delivery systems when loading drugs onto Ion-channel resins. A single batch of drugs isn't enough to produce complexes with large levels of drug loading. Double-batch loading of medicines into the resins could be advised based on the results in tables 4.8 and 4.9. The catIon-channel resin employed in this experiment has a functional group of SO3 - H +. More acidic by-products can be formed as complexes grow in size and complexity. To avoid this, it is important to remove them from the system as they may impact a variety of factors, including pH and drug loading.

Effect of pH on drug loading

To further understand the influence of pH on drug loading onto resins, tramadol hydrochloride was investigated in various pH mediums, and the results are shown in table 7.9. Tramadol hydrochloride has a pKa of 9.41, which means that it is totally ionised at all pH values, according to the pH partition theory (1-8). Sulphonic (strong acid) moiety catIon channels in Indion resins will be ionicized regardless of pH variations. Figure 4.9 shows that pH has a negligible influence. At lower pH values, it was shown that pH had no effect on drug loading, but at higher pH, less H + channelable ions in the loading media resulted in an increased complexation of the drug, resulting in an increase in drug loading. Tramadol hydrochloride complexation with the Ion-channel resins was shown to be unaffected by the pH of the loading media in these tests. Furthermore, as the complex formation neared completion, the pH of the eluent returned to the starting pH of the eluent due to the restricted supply of H+ ions available for channel. Another method to assess drug loading onto Ion-channel resins is to monitor changes in pH of the complexation. The endpoint of the complex development can be determined by measuring.

Table 1. Effect of pH on drug loading onto resins

EVALUATION OF DRUG DELIVERY SYSTEM - ION GATE CHANNELS

рН	Indion 224	Indion 244	Indion 254	Indion 284
2	51.84±0.22	72.82±1.02	58.24±0.44	50.48±0.16
4	51.25±0.68	71.22±0.34	58.04±0.63	50.27±0.47
5	50.88±0.29	70.92 ± 0.28	58.29±0.38	49.81±0.52
6	51.28±0.54	71.08±0.26	57.98±0.29	50.09±0.18
7	51.24±0.28	71.12±0.51	58.08±0.25	49.98±0.53
8	48.04±0.37	69.35±0.37	56.28±0.84	46.09±0.38
10	46.32±0.19	68.83±0.12	55.21±0.39	44.94±0.62

Mean \pm S.D., n = 3

In vitro drug release profiles of tramadol hydrochloride-resinates

The in vitro drug release of the tramadol hydrochloride was performed to study the effect of pH, ionic strength of dissolution medium and valency of ions as:

Effect of pH media on drug release from the resinates

It was found that varied pH media affected the release of drugs from resinates. For this purpose the investigations were conducted in distilled water, 0.1N HCl, and pH 6.8 buffer. According to table 4.10-11, distilled water, pH 6.8 buffer and 0.1 N HCL were investigated for the amount of drug released (i.e. percentage of drug released).

According to Figure 4.10, the release profiles reveal that pH had no effect on the medication release. The degree of ionisation of the resin particles in the dissolution medium has a direct impact on the drug release mechanism. According to the findings, the drug release is primarily due to the substitution of drug ions in the dissolving media by H+ ions. There was no evidence that the pH of the dissolving medium had any effect on the amount of medication released. In the dissolution media used in the study, resin was ionised in the same way. Consequently, all additional dissolution studies were carried out in distilled water.[11-20]

Effect of ionic strength of dissolution medium on drug release

The effect of the ionic strength of the dissolution medium was studied by adjusting the ionic strength of the dissolution medium by 0.01 M, 0.05M and 0.1 M NaCl.

Table 2. Effect of ionic strength of dissolution medium on % drug release from the Indion224 resinate

Time (hrs)	% Drug released			
	0.01M NaCl	0.05M NaCl	0.1M NaCl	
0.5	12.84±0.18	14.36±1.24	18.29±1.02	
1	19.38±0.24	23.46±0.97	27.68±0.39	
2	38.34±0.89	42.68±1.12	49.26±0.28	
3	53.83±1.28	63.24±0.62	66.78±0.69	
4	69.24±0.58	73.35±0.81	80.58±1.08	
5	82.49±0.37	86.32±1.62	93.29±1.48	

Mean \pm S.D., n = 3

Table 6. Effect of ionic strength of dissolution medium on % drug release from the Indion244 resinate

Time (hrs)	% Drug released				
	0.01M NaCl	0.05M NaCl	0.1M NaCl		
0.5	10.12±1.27	12.89±0.69	15.26±0.48		
1	14.28±0.58	17.24±1.42	20.46±1.18		
2	30.25±0.72	36.61±0.93	44.18±0.62		
3	47.86±1.09	56.47±0.68	61.24±1.03		
4	58.29±0.42	65.27±0.89	72.86±0.57		
5	62.98±2.62	78.53±1.68	84.19±2.48		

Mean \pm S.D., n = 3

Eur. Chem. Bull. 2023, 12(Special issue 8), 6089-6101

4.CONCLUSION

Weak acids and weak bases make up the majority of the pharmaceuticals. In order to use a prolonged release mechanism, drugs with limited water solubility will be challenging to incorporate To slow down the dissolving rate of a medicine with high solubility and quick dissolution, it is typically challenging. High water solubility drugs dissolve easily in water or gastrointestinal fluid and tend to release their dosage form in a burst, which results in an increase in blood drug concentration more quickly than with less soluble drugs. Some commercial resinbased formulations for controlled drug administration have been developed after extensive research into the use of ion-channel resins in medical and pharmaceutical applications. Resins with ion channels have been explored for at least oral, transdermal, nasal and ocular drug administration as a controlled release formulation at this time. To achieve prolonged, site-specific, or pulsed effects, many different pharmacological therapies benefit from a regulated release of active components. Constant or sustained release solutions utilising Ion channels have been used to improve therapeutic safety and efficacy; boost patient compliance; shorten dosing times; and increase drug stability, among other benefits.

5.REFERENCES

1. Anger T, Madge DJ, Mulla M, Riddall D. Medicinal chemistry of neuronal voltage-gated sodium channel blockers. *J. Med. Chem.* 2001;44(2):115–137.

2. Mantegazza M, Curia G, Biagini G, Ragsdale DS, Avoli M. Voltage-gated sodium channels as therapeutic targets in epilepsy and other neurological disorders. *Lancet Neurol.* 2010;9(4):413–424.

3. Chahine M, Chatelier A, Babich O, Krupp JJ. Voltage-gated sodium channels in neurological disorders. *CNS Neurol. Disord. Drug Targets.* 2008;7(2):144–158.

4. Tarnawa I, Bolcskei H, Kocsis P. Blockers of voltage-gated sodium channels for the treatment of central nervous system diseases. *Recent Pat. CNS Drug Discov.* 2007;2(1):57–78.

5. Cestèle S, Catterall WA. Molecular mechanisms of neurotoxin action on voltage-gated sodium channels. *Biochimie*. 2000;82(9-10):883–892.

EVALUATION OF DRUG DELIVERY SYSTEM - ION GATE CHANNELS

6. Kyle DJ, Ilyin VI. Sodium channel blockers. J. Med. Chem. 2007;50(11):2583-2588.

7. Patino G, Isom LL. Electrophysiology and beyond: multiple roles of Na+ channel β subunits in development and disease. *Neurosci. Lett.* 2010;486(2):53–59.

8. Sun GC, Werkman TR, Battefeld A, Clare JJ, Wadman WJ. Carbamazepine and topiramate modulation of transient and persistent sodium currents studied in HEK293 cells expressing the Na(v)1.3 alpha-subunit. *Epilepsia*. 2007;48(4):774–782.

9. Hammarström AKM, Gage PW. Hypoxia and persistent sodium current. *Eur. Biophys.* J. 2002;31(5):323–330.

10. Patino GA, Brackenbury WJ, Bao Y, Lopez-Santiago LF, O'Malley HA, Chen C, Calhoun JD, Lafrenière RG, Cossette P, Rouleau GA, Isom LL. Voltage-gated Na+ channel β1B: a secreted cell adhesion molecule involved in human epilepsy. *J. Neurosci.* 2011;31(41):14577–14591.

11. Martin MS, Tang B, Papale LA, Yu FH, Catterall WA, Escayg A. The voltage-gated sodium channel Scn8a is a genetic modifier of severe myoclonic epilepsy of infancy. *Hum. Mol. Genet.* 2007;16(23):2892–2899.

12. Large CH, Kalinichev M, Lucas A, Carignani C, Bradford A, Garbati N, Sartori I, Austin NE, Ruffo A, Jones DNC, Alvaro G, Read KD. The relationship between sodium channel inhibition and anticonvulsant activity in a model of generalised seizure in the rat. *Epilepsy Res.* 2009;85(1):96–106.

13. Wimmer VC, Reid CA, So EY, Berkovic SF, Petrou S. Axon initial segment dysfunction in epilepsy. *J. Physiol.* 2010;588(Pt 11):1829–1840.

14. Whitaker WR, Faull RL, Dragunow M, Mee EW, Emson PC, Clare JJ. Changes in the mRNAs encoding voltage-gated sodium channel types II and III in human epileptic hippocampus. *Neuroscience*. 2001;106(2):275–285.

15. Sheets PL, Heers C, Stoehr T, Cummins TR. Differential block of sensory neuronal voltage-gated sodium channels by lacosamide [(2*R*)-2-(acetylamino)-N-benzyl-3-3-methoxypropana mide], lidocaine, and carbamazepine. *J. Pharmacol. Exp. Ther.* 2008;326(1):89–99. *Eur. Chem. Bull.* 2023,12(Special issue 8), 6089-6101 6100

16. Nicholson E, Randall AD. Na(v)1.5 sodium channels in a human microglial cell line. *J. Neuroimmunol.* 2009;215(1-2):25–30.

17. Rush AM, Dib-Hajj SD, Waxman SG. Electrophysiological properties of two axonal sodium channels, Nav1.2 and Nav1.6, expressed in mouse spinal sensory neurones. *J. Physiol.* 2005;564(Pt 3):803–815.

18. Waxman SG, Cummins TR, Dib-Hajj S, Fjell J, Black JA. Sodium channels, excitability of primary sensory neurons, and the molecular basis of pain. *Muscle Nerve*. 1999;22(9):1177–1187.

19. Sander JW. The epidemiology of epilepsy revisited. *Curr. Opin. Neurol.* 2003;16(2):165–170.

20. Khan HN, Kulsoom S, Rashid H. Ligand based pharmacophore model development for the identification of novel antiepileptic compound. *Epilepsy Res.* 2012;98(1):62–71.