ISSN 2063-5346



EFFECTS OF PESTICIDES AND SAMPLE DRYING ON ENZYME ACTIVITY OF β-GLUCOSIDASE AND CELLULASE ENZYME IN SOIL OF FARRUKHABAD REGION (UP) INDIA

	¹ Adarsh Kumar and Ranvir Singh					
Article History: Received: 10.05.2023	Revised: 29.05.2023	Accepted: 09.06.2023				

Abstract

In the present study, pesticides are widely used in crop production and are known to cause major environmental problems. Effect of pesticides on enzyme activity of β -glucosidase and cellulase enzymes in soil of Farrukhabad region. Soil were collected in winter season (January 2022) from Farrukhabad region viz., Kaimganj, Amritpur and Farrukhabad tahsil. Soil microbial diversification is indispensible to maintain functional diversity and enzyme mediated critical soil process that detoxify soil from environmental pollutants like pesticides, due to excessive use of pesticides viz., Cypermethrin, Endosulfan and Mancozeb controlling the insects. In India the present study was carried out to assess the effect of different concentration of the pesticides. Results shown that the effect of pesticides on soil enzyme activity slightly decreases while with out pesticides enzyme activity of β -glucosidase and cellulase increases in selected soil.

Keyword : Pesticides, Soil, β -glucosidase and Cellulase enzyme.

Department of Chemistry, S.V. College (Dr. B.R.Ambedkar University Agra) Aligarh ¹Email ID: kumaradarsh65@gmail.com

DOI:10.31838/ecb/2023.12.s1-B.484

INTRODUCTION

Pesticides are widely used in crop production and are known to cause major environmental problems. With increasing pesticides use. questions are rising on potential effect regarding public health and environment. Pesticides pollute air, soil, water resources and contaminate food chain The interaction the between soil components and pesticides influences the biochemical processes driven by microorganisms. The effect of pesticides on soil microorganism could be determined by the study of functional parameters such as carbon and nitrogen mineralization that are governed by enzymatic activities. Those activities, play an important role biochemical because all transformations in soil depend on or are related to the presence of enzymes.¹ Soil enzymes play key biochemical functions in the overall organic process of matter decomposition in the soil system.² They are important in catalysing several important reactions necessary for the life processes of microorganisms in soils and the stabilization of soil structure, the decomposition of organic organic wastes, matter formation and nutrient cycling.³ These enzymes are constantly being synthesized, accumulated, inactivated and/or decomposed in the soil, hence playing an important role in agriculture and particularly in nutrients cycling.⁴ The activities of these enzymes in soils undergo complex biochemical processes consisting of integrated and ecologically-connected synthetic processes, and in the immobilisation and enzyme stability.⁵ In this regard, all soils contain a group of enzymes that determine soil metabolic processes⁶ which, in turn, depend on its physical, chemical, microbiological and biochemical properties.

β-glucosidase

β-glucosidase is characteristically useful as a soil quality indicator, and may give a reflection of past biological activity. The capacity of soil to stabilise the soil organic matter, and can be used to detect management effect on soils. It is named according to the type of bond that it hydrolyses. This enzyme plays an important role in soils because it is involved in catalysing the hydrolysis and biodegradation of various Bglucosides present in plant debris decomposing in the ecosystem. Its final product is glucose, an important carbon energy source of life to microbes in the soil.⁷

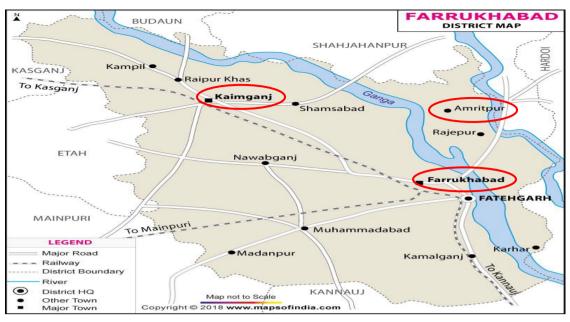
Cellulase

Cellulase is the most abundant organic compound in the biosphere. comprising almost 50% of the biomass synthesized by photosynthetic fixation of CO₂.⁸ Growth and survival of micro-organisms important in most agricultural soils depends on the carbon source contained in the cellulase occurring in the soils.⁹ However, for carbon to be released as an energy source for use by the microorganisms, cellulase in plant debris has to be degraded into glucose, cellobiose molecular and high weight oligosaccharides cellulases by enzymes.¹⁰ Cellulases are a group of enzymes that catalyse the degradation of cellulase, polysaccharides build up of 1, 4 linked glucose units.¹¹

MATERIALS AND METHODS

Collection of Soil Sample

The soil pertaining to the experimental setup were collected from the region of Farrukhabad U.P., India at different sample drying.





Pesticides

Selected pesticides are cypermethrin (insecticides), endosulfan (insecticide) and mancozeb (fungicide). It were used techniques Flamphotometer, UV Vis spectrophotometer and X-ray differaction spectroscopy in proposed research work.

Soil Analysis

Physicochemical characteristics of soil (control) and selected pesticides determined with were soil of Farrukhabad Region. Parameters were determined such as pH, EC (Electric Conductivity), TOC (Total Organic Carbon). TKN (Total Kieldahl Nitrogen), TP (Total Phosphorous), TK (Total Potassium) and TKN (Total Kjeldahl Nitrogen) from selected soil and include pesticides soil. It were used analytical procedures by total kjeldahl nitrogen (TKN) and total organic carbon (TOC) of the soil analysis were measured with the micro kjeldahl methods¹² and Walkely and Black's Rapid titration method (1934)¹³ respectively, total phosphorous (TP) determined was spectrophotometrically¹⁴ While total

potassium (TK) was detected by flame photometer.¹⁵

Enzyme Analysis

- β-glucosidase activity was assessed by determination of the released p-nitro phenol after incubation of samples (1 gm fresh weight) with pnitrophenolglucoside (0.025 m) for 1h at 37°C in a microplate reader at 400nm.
- Cellulase activity was estimated by determination of released reducing sugars after incubation of samples (5 g fresh weight) with carboxymethylcellulase (CMC), sodium salt (0.7%) for 24 h at 50°C in a microplate reader 690 nm.¹⁶

RESULT AND DISCUSSION

β-glucosidase is characteristically useful as a soil quality indicator, and may give a reflection of past biological activity. The capacity of soil to stabilise the soil organic matter, and can be used to detect management effect on soils¹⁷. Understanding the dynamics of enzyme activities in these systems is crucial for predicting their interactions as their activities may, in turn, regulate nutrient uptake and plant growth. Cellulase enzymes play an important role in global recycling of the most abundant polymer, cellulose in nature, it would be of critical importance to understand this enzyme better so that it may be used more regularly as a predictive tool in our soil fertility programmes. The effect of pesticides on soil enzyme activity slightly decreases while with out pesticides enzyme activity of β -glucosidase and cellulase increases in selected soil.18 Thus the effects of pesticides on enzyme activity of cellulase and \Box glucosidase enzymes in soil of (Amritpur, Farrukhabad region Kaimganj and Farrukhabad) were observed in the present study, the observed results shown are given below:

TABLE 1

Physico-chemical characteristics of soil of Farrukhabad (Amritpur, Kaimganj and Farrukhabad) region in winter season (January 2022). The various physicochemical properties were obtained from R.G. College of Pharmacy, Hathras.

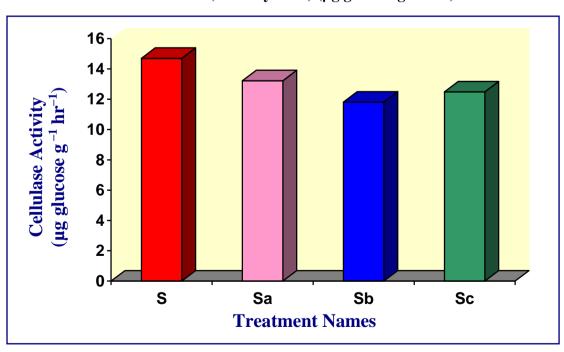
рН (1:2.5)	EC (dS/m) 1:2.5	Organic carbon (%)	Available P2O5 (mg kg ⁻¹)	Available K2O (mg kg ⁻¹)	Available Nitrogen (mg kg ⁻¹)	Sodium (%)		
AMRITPUR REGION								
7.61	7.70	0.51	14.38	403.11	172.85	0.51		
KAIMO	GANJ RE	GION						
7.53	7.85	0.50	14.78	413.02	177.67	0.56		
FARRUKHABAD REGION								
7.53	7.67	0.51	15.12	427.57	166.83	0.67		

Sample code	Treatment Name	Replicates	Activity	Mean	Mean ± Standard Deviation	
	G	1	14.85			
1	S	2	14.67	14.70	14.70 ± 0.11	
		3	14.58			
	q	1	13.10	13.22		
2	Sa	2	13.25		13.22 ± 0.09	
		3	13.33			
		1	11.93			
3	Sb	2	11.85	11.81	11.81 ± 0.10	
		3	11.67			
		1	12.37			
4	Sc	2	12.45	12.49	12.49 ± 0.12	
		3	12.67			
S =	soil;		Sa	= so	oil + cypermethrin;	
Sb =	soil + ende	endosulfan; Sc = soil + mencozeb.				

Cellulase Activity of Amritpur soil in winter season (January 2022) (μ g glucose g⁻¹ hr⁻¹)

TABLE 2

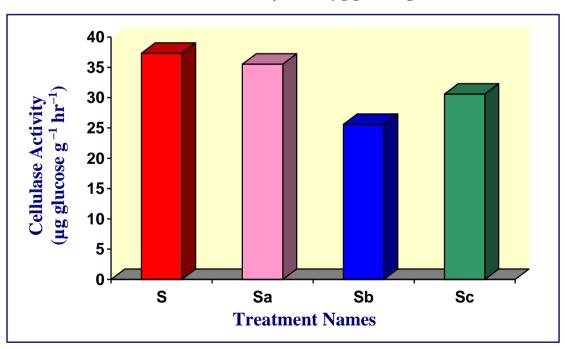
Figure 1 : Cellulase Activity of Amritpur soil in winter season (January 2022) (μ g glucose g⁻¹ hr⁻¹)



		(µg giucos	cg m)		
Sample code	Treatment Name	Replicates	Activity	Mean	Mean ± Standard Deviation
	C	1	37.15		
1	S	2	37.33	37.32	37.32 ± 0.13
		3	37.48		
	G	1	35.85	35.52	35.52 ± 0.25
2	2 Sa	2	35.23		
		3	35.48		
		1	25.48		
3	Sb	2	25.57	25.59	25.59 ± 0.10
		3	25.74		
		1	30.93		
4	4 Sc	2	30.68	30.59	30.59 ± 0.31
		3	30.18]	
S =	soil;		Sa =	soil + c	ypermethrin;
Sb =	soil + endos	ulfan;	Sc =	soil + m	encozeb.

Cellulase Activity of Kaimganj soil in winter season (January 2022) $(\mu g \ glucose \ g^{-1} \ hr^{-1})$

Figure 2 : Cellulase Activity of Kaimganj soil in winter season (January 2022) (μ g glucose g⁻¹ hr⁻¹)

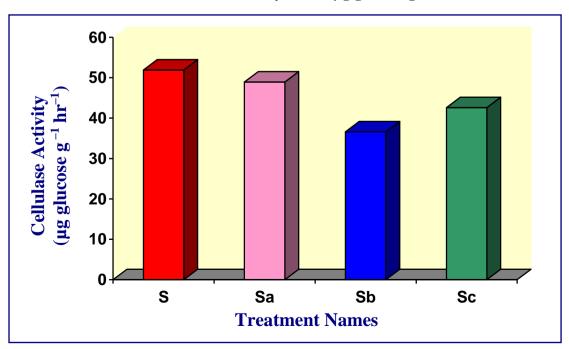


Cellulase Activity of Farrukhabad soil in winter season (January 2022)
$(\mu g \text{ glucose } g^{-1} hr^{-1})$

Sample code	Treatment Name	Replicates	Activity	Mean	Mean ± Standard Deviation
	S	1	51.18		
1	3	2	51.85	51.90	51.90 ± 0.61
		3	52.68		
	S.c.	1	48.25		
2	Sa	2	49.18	48.92	48.92 ± 0.47
		3	49.33		
		1	36.85		
3	Sb	2	36.15	36.61	36.61 ± 0.32
		3	36.83		
		1	42.38		
4	Sc	2	42.83	42.58	42.58 ± 0.18
		3	42.55		

S	=	soil;	Sa	=	soil + cypermethrin;
Sb	=	soil + endosulfan;	Sc	=	soil + mencozeb.

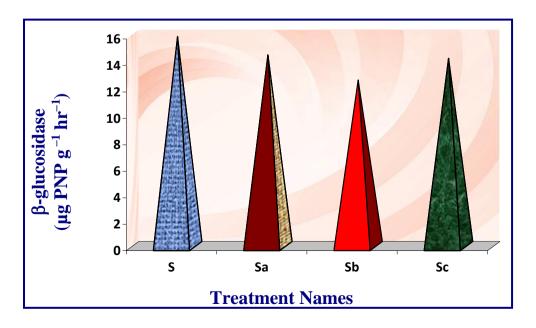
Figure 3 : Cellulase Activity of Farrukhabad soil in winter season (January 2022) (μ g glucose g⁻¹ hr⁻¹)



Sample code	Treatment Name	Replicates	Activity	Mean	Mean ± Standard Deviation		
	C	1	15.85				
1	S	2	15.77	15.81	15.81 ± 0.33		
		3	15.81				
	G	1	14.83				
2	Sa	2	14.33	14.44	14.44 ± 0.29		
		3	14.11	-			
		1	12.14				
3	Sb	2	12.88	12.54	12.54 ± 0.30		
		3	12.61	-			
		1	14.10				
4	Sc	2	14.41	14.18	14.18 ± 0.16		
		3	14.03	1			
S =	soil;	1	Sa =	soil +	cypermethrin;		
Sb =	soil + endosulf	Sc =	soil +	mencozeb.			

β -glucosidase Activity of Amritpur soil in winter season (January 2022) (µg PNP g $^{-1}\,hr^{-1})$

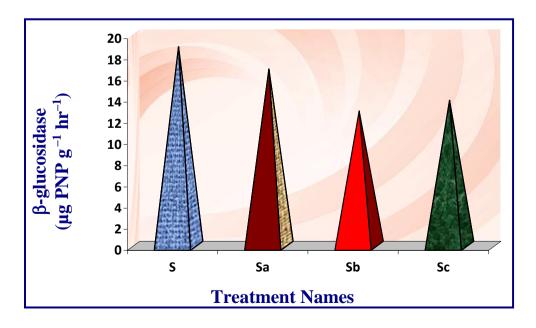
Figure 4 : \Box -glucosidase Activity of Amritpur soil in winter season (January 2022) (µg PNP g⁻¹ hr⁻¹)



Sample code	Treatment Name	Replicates	Activity	Mean	Mean ± Standard Deviation	
		1	18.58			
1	S	2	18.87	18.79	18.79 ± 0.15	
		3	18.93			
		1	16.95		16.68 ± 0.21	
2	2 Sa	2	16.43	16.68		
		3	16.67			
		1	12.67			
3	Sb	2	12.59	12.73	12.73 ± 0.14	
		3	12.93			
		1	13.95			
4 Sc	2	13.89	13.74	13.74 ± 0.25		
		3	13.39	1		
S =	soil;	-	-	Sa =	soil + cypermethrin	
Sb =	soil + e	endosulfan;		Sc =	soil + mencozeb.	

β -glucosidase Activity of Kaimganj soil in winter season (January 2022) (μ g PNP g⁻¹ hr⁻¹)

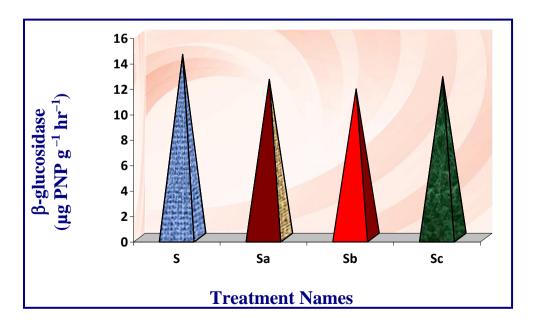
Figure 5 : \Box -glucosidase Activity of Kaimganj soil in winter season (January 2022) (µg PNP g⁻¹ hr⁻¹)



β -glucosidase Activity of Farrukhabad soil in winter season (January 2022) ($\mu g PNP g^{-1} hr^{-1}$)

Sample code	Treatment Name	Replicates	Activity	Mean	Mean ± Standard Deviation
		1	14.05		
1	S	2	14.35	14.39	14.39 ± 0.36
		3	13.97		
	Sa	1	12.11		12.43 ± 0.31
2		2	12.33	12.43	
		3	12.85		
		1	11.89		
3	Sb	2	11.77	11.69	11.69 ± 0.20
		3	11.41		
	Sc	1	12.23	12.64	
4		2	12.81		12.64 ± 0.29
		3	12.89		

Figure 6 : □-glucosidase Activity of Farrukhabad soil in winter season (January 2022) (µg PNP g⁻¹ hr⁻¹)



CONCLUSION

The results obtained in the present study indicate that pesticides were increasingly used in agriculture in order to limit crop diseases and increase food production. Effects of pesticides on enzyme activity of β -glucosidase and cellulase enzymes feable decrease as compared to without pesticides soil of Farrukhabad region (U.P.) India.

ACKNOWLEDGEMENT

We acknowledge the technical support of the R.G. College of Pharmacy, Hathras to carryout this study.

REFERENCES

- 1. Raith W., Laval K., Laroche-Ajzenberg E., Mougin C., Latour X., Trinsoutrot-Eattin I. (2014). Effects of pesticides on soil enzymes : A review. Environ Chem. Lett.
- 2. Burns R.G. (1983). In: Microbes in Their Natural Environment pp. 249-298. Cambridge University Press, London.
- Dick R.P., Sandor J.A., Eash N.S. (1994). Agric. Ecosyst. Environ. 50: 123 – 131.
- 4. **Tabatabai M.A. (1994).** SSSA Book Series No. **5**. Soil Sci. Soc. Am. Madison, Wis., pp. 775-833.
- 5. Khaziyev F.K., Gulke A.Y. (1991). Pochvovedenie, 8: 88-103.
- 6. **McGill W.B., Colle C.V.** (**1981**). *Geoderma*. **26**: 267-286.

- 7. Ndiaye E.L. (2000). Am. J. Alterm Agric., 15: 26-36.
- Deng S.P., Tabatabai M.A. (1995). Soil Biol. Biochem. 27(7): 977-979.
- 9. Eriksson K.E.L., Blancbette R.A., Ander P. (1990). Springer-Verlag, New York. 89-180.
- 10. **Richmond P.A.** (1991). In : Biosynthesis and Biodegradation of Cellulose (Haigler CH and Weimer PJ Eds), Dekker, New York, 5-23.
- 11. White, A.R. (1982). Plenum *Press, New York*, 489-509.
- 12. Shaw J., Beadle, L.C. (1949). J. Exp. Biol. 26: 15-23.
- 13. Walkely, J.A., Black, J.A. (1934). Soil Science. 37: 29-31.
- 14. **Fiske, C.H., Row Subha, Y.** (1925). J. Biol. Chem. 66: 375-400.
- 15. **Person, R.W. (1952).** J. Soil Science, **74(4)**: 301-310.
- Schinner F., Vol Mersi W. (1990). Soil Biol. Biochem. 22: 511-515.
- 17. Ndiaye E.L. (2000). Am. J. Alterm Agric., 15: 26-36.
- 18. **Kumar A and Singh, R.** (2023). Journal of survey in fisheries science, 10(2S): 1310-1317.