

Phytochemical Investigation of Methanolic Extracts of Bryophyllum Pinnatum by Gas Chromatography-Mass Spectrometry (GC-MS) Method

Kalpana Verma¹, Diksha Pandey², Lavkush Dwivedi^{1*}

| | Article History: | Received: 25.06.2023 | Revised:15.07.2023 | Accepted: 25.07.2023 |
|--|------------------|-----------------------------|--------------------|----------------------|
|--|------------------|-----------------------------|--------------------|----------------------|

Abstract

India is a gold mine of historical medical knowledge. The central Indian region of Bundelkhand has a unique environment, distinguished by sandy sand and rocky soil, yet it is also home to a wide variety of plants. This area is home to several medicinal plants that are prized for their ability to treat a wide range of disorders and dangerous diseases. Over a year, the thorough examinations of this area uncovered about 66 therapeutic herbs and shrubs as well as 38 tree species. In the current study, the phytochemical components of the *Bryophyllum pinnatum* leaf were evaluated. GCMS is used to identify the bioactive components in the methanol-based leaf extract. A preliminary phytochemical screening showed the presence of various Phytocompounds such as glycosides, cardiac glycosides, phenol, tannins, Flavonoids, Phytosterols, Saponins, etc. In a gas chromatography-mass spectrometer examination of the extracts mentioned, a maximum of 42 compounds were identified by mass spectrum matching with the database of the National Institute Standard and Technology (NIST). *Bryophyllum pinnatum* leaves are reported for their biological activities such as Antimicrobial, antioxidant, antitumor, anti-inflammatory, antifungal, Antiviral, Diabetes, inflammation, cancer, Antidote, coronary-dilator, and diuretic activity.

Keywords: *Bryophyllum pinnatum*, GC-MS analysis, Phytochemical Screening, Methanol, Soxhlet extraction.

¹Department of Biotechnology Bundelkhand University, Jhansi-284128, U.P, India

^{2*1}Department of Biomedical Sciences Bundelkhand University, Jhansi- 284128 U.P, India

*Corresponding Author: Dr. Lavkush Dwivedi, Email: <u>lavkush@bujhansi.ac.in</u>

DOI: 10.48047/ecb/2023.12.Si12.181

1. INTRODUCTION

Bryophyllum pinnatum (Lam.) Oken (family: Crassulaceae) (synonym: Kalanchoe pinnata, Lam.; common names: Love plant, life plant, Mexican air plant, cathedral bells, Canterbury bells, etc is a perennial herb that is widely grown and utilized in folk medicine in tropical Asia, India, China, Africa, and Australia [1]. In medicinal and aromatic plants, there is a vast potential for the discovery of novel bioactive chemicals [2]. Many of the medications that are today prescribed by doctors have a lengthy history of use as herbal cures or were created as plant-based equivalents [3]. The value of traditional medical systems and particular traditional medical practices is widely acknowledged on a global scale. Due to its ease of accessibility, availability, cultural acceptability, therapeutic potential, and affordability, some authors have recently argued that traditional medicinal systems should be incorporated into the mainstream of healthcare services^[4]. The growing interest in using medicinal plants, particularly in primary healthcare, has prompted a rise in interest in plants that may be useful as medicines [5]. In traditional medicine, the leaves and bark of *B. pinnatum* are used to cure lithiasis, earaches, burns, abscesses, diarrhea, and other conditions. They are also bitter tonic, astringent, analgesic, and carminative. Leaf juice is applied topically to treat conditions such as smallpox, earaches, otitis, coughs, asthma, palpitations, headaches, convulsions, and general weakness [6]. The treatment is also given to young children who are ill. It is mostly used in traditional medicine to treat hypertension and kidney stones [7]. Recent studies have emphasized positive qualities such as free radical scavenging activity [8].B. Pinnatum has high levels of lipids, alkaloids, triterpenes, glycosides, flavonoids, cardenolides, steroids, and bufadienolides. The leaves include a class of substances known as Bufadienolides, which are highly active and have antibacterial, antitumor, cancer-preventive, and insecticidal properties [9]. There hasn't been any research done on the phytochemistry of the *B. Pinnatum* that is grown in the Bundelkhand region. Based on information from the literature, we are describing the phytochemical analysis of with association of these phytochemicals in the management of various illnesses in this research. This study will clarify the active principles in charge of its multiple reported therapeutic effects and offer a template for identifying, isolating, and purifying these molecules for additional research and medication development.

2. MATERIALS & METHODS

2.1 Collection of the plant material

The leaves of *B.pinnatum* were collected in October and got authenticated (Accession No. 28571) by The Central Council for Research in Ayurvedic Science

(CCRAS) - Regional Ayurveda Research Institute Gwalior Road, Jhansi Uttar Pradesh, during November 2019.

2.2. Preparation of plant extract:

The plant material was first washed with tap water and distilled water, followed by a 2-4 week period of drying in the shade. Using an electric mixer, the dried leaves (20gm) were converted into a fine powder. *B. pinnatum* leaf powder was put in a Soxhlet apparatus and extracted for 9(W/V) or 16–18 hours at 64.6°C using 200ml of Methanol AR grade solvent. Whatman filter paper No. 41 (110 mM.) and cotton wool were used to filter the pure extract of B. pinnatum leaves after it had been collected. The extract was concentrated using a Rotary Evaporator after filtering. Specifications for the model include EYELA N-12008 and EYELA-UNI TRAP UT-1000, with the water bath set to 45–50°C. The resultant solution was kept chilled at 4°C for further testing.

2.3. Preparation Phytochemical Screening:

The extract was subjected to preliminary phytochemical screening and qualitative analyses using the established techniques for determining the presence of carbohydrates, alkaloids, amino acids, phenol, tannins, flavonoids, saponins, terpenoids, quinines, cardiac glycosides, and steroids by distinctive color change [10,11].

2.4. GC-MS (Gas Chromatography-Mass spectroscopy) analysis:

On the GC-Claus 680 MS-SQ-8C PerkinElmer system, which includes an AOC-20i autosampler and a gas chromatograph attached to a mass spectrophotometer (GC-MS), GC-MS analysis was carried out. The following detection parameters were used to examine the sample: Furnace beginning temperature 40°C for 5 minutes, ramp 12°C/min to 260°C, hold 10 minutes, Inj B auto 250°C, volume 0 μ L, division 50:1, carrier gas He (99.999%), solvent delay 2.00 minutes, transfer temperature 1800 °C, source temperature 200°C, scan: 50 to 500 Da[12].

3. RESULTS AND DISCUSSION

3.1 Preliminary Phytochemical screening:

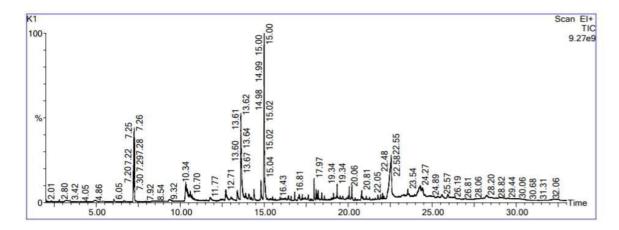
Phytochemical screening for primary metabolites (carbohydrates, Starch, proteins, amino acids, oils, and fat) and secondary metabolites (Anthraquinones, Quinines, Glycosides, cardiac, glycosides, Phenol, Tannins, Flavonoids, Phytosterols, Saponins, and Steroids) were carried out.

Table1: Preliminary phytochemical evaluation of methanolic extracts of *B*. pinnatumleaves. (+) = Present; (-) = Absent

| Phytochemical Compounds Name | Tests Name | <i>Bryophyllum pinnatum</i> leaf |
|------------------------------------|---------------------------------|--------------------------------------|
| Carbohydrates | Benedict's test | + |
| Starch | Iodine test | - |
| Proteins | Nitric acid test | + |
| Amino acids | Ninhydrin reagent test | + |
| Quinines | KOH test | - |
| Glycosides | Glacial acetic acid | - |
| Cardiac glycosides | Keller-Killani test | + |
| Phenol | Lead acetate test | + |
| Tannins | Ferric chloride test | + |
| Flavonoids | H2SO4 test | + |
| Phytosterols | KOH+ acetic acid+H2SO4 | + |
| Saponins | Vigorously shaken water Test | + |
| Steroids | chloroform +H2SO4 test | - |

3.2 Components are identified:

The National Institute of Standard and Technology (NIST) database, a private collection of more than 7,900 chemicals, was used by MS to interpret the GC chromatogram. Figure 1 provides information about the compound, such as its name, molecular weight, and the composition of its constituent parts as they are present in the test materials.





3.3 GC-MS analysis of methanolic extract from *Bryophyllum pinnatum*:

One of the extensively used methods for identifying the components of volatile matter, including long-chain and branched-chain hydrocarbons, alcohols, acids, and esters, is gas chromatography-mass spectrometry (GC-MS) with the NIST library.

Using the GCMS-NIST library, we were able to isolate 43 main components from the methanolic extract of *B. pinnatum* (Table 2). For each, the Pubchem search was conducted. These important chemicals and details on their alleged pharmacological activities were acquired from previously published literature. tert-Hexadecanethiol, Nonahexacontanoic acid,17-Pentatriacontene, Disulfide, di-tert-dodecyl, cis-1-Chloro-9-octadecene, Tetradecanein our study through GC-MS analysis has revealed the added medicinal value such asAntibacterial and antioxidant activity, used in cosmetics, Antiinflammatory, Anticancer, Antiarthritic, Antidote, coronary-dilator, diuretic, increase superoxide dismutase activity, Skin protection and respiratory protection, and Cytotoxicity, Antipyretic, Anthelmentic, Tumour, Bronchitis, Asthma, Tuberculosis, Dyspepsia, Constipation, Anemia, Throat diseases, Elephantiasis, Antidiabetic, Antidiarrhoeal, etc. properties of the identified compounds have also been reported in the literature (table 2). The current report on the bioactive components of *B. pinnatum* supports the plant's application in the treatment of numerous illnesses [6]. These substances change according to the climate, soil quality, and geographic location of the plant [13]. B. pinnatum contains a variety of bioactive chemicals, which explains why traditional healers employ the entire plant to treat a variety of diseases. Additionally, a thorough examination of the numerous substances found in *B. pinnatum* and their pharmacological assessment may aid in the development of medications with multiple effects for the treatment of cancer and other microbial disorders

Table 2: Detail of compounds identification from GC-MS analysis of an acetone extract of *B. pinnatum*.

| Sr. No | RT | Peak % | Phytochemical Compound | Formula | M.W | Activity |
|-----------|--------|-----------|--|--|--------|--|
| 1. | 11.289 | 1.505 | tert- Hexadecanethiol | C ₄₈ H ₉₉ Au S ₃ | 969.5 | Antibacterial and antioxidant activity [14] |
| 2. | 11.289 | 1.505 | Nonahexacontano ic acid | C ₆₉ H ₁₃₈ O ₂ | 999.8 | used in cosmetics [15] |
| 3. | 11.289 | 1.505 | Tetrapentacontan e, 1,54-dibromo- | $C_{54}H_{108}Br_2$ | 917.2 | Antioxidant [16] |
| 4. | 11.289 | 1.505 | 2-Nonadecanone 2,4- dinitrophenylhydr azine | C ₂₅ H ₄₂ N ₄ O ₄ | 462.6 | Antimicrobial [17] |
| 5. | 11.289 | 1.505 | 17- Pentatriacontene | C ₃₅ H ₇₀ | 490.9 | Antiinflammatory Anticancer Antibacterial Antiarthritic [18] |
| 6. | 11.688 | 2.673 | Tetradecane | $C_{14}H_{30}$ | 198.39 | Antimicrobial [19] |
| 7. | 11.688 | 2.673 | Nonane, 2- methyl-5-propyl- | C ₁₃ H ₂₈ | 184.36 | Anticancer [20] |
| 8. | 11.688 | 2.673 | Pentadecane | C ₁₅ H ₃₂ | 212.41 | Antibacterial activity [21] |
| 9. | 11.870 | 1.093 | Disulfide, di-tert- dodecyl | $C_{24}H_{50}S_2$ | 402.8 | Antidotes, coronary-dilator, and diuretics increase superoxide dismutase activity [22] |
| 10. | 11.870 | 1.093 | Methoxyacetic acid, 4-tetradecyl ester | C ₁₇ H ₃₄ O ₃ | 286.4 | Anti-microbial [23] |
| 11. | 12.011 | 1.826 | 4- Methyldocosane | C ₂₃ H ₄₈ | 324.6 | Anti- inflammatory, antimicrobial, antioxidant, diuretic activity, anticancer [24] |

Eur. Chem. Bull. 2023, 12(Special issue 12), 2075-2084

| 12. | 12.011 | 1.826 | Heneicosane, 11- (1-ethylpropyl)- | C ₂₆ H ₅₄ | 366.7 | Major component of safflower flower essential oil [25] |
|-----|--------|-------|--------------------------------------|--|--------|---|
| 13. | 12.389 | 1.483 | Tetradecane | C ₁₄ H ₃₀ | 198.39 | Antifungal and Antibacterial [26] |
| 14. | 12.389 | 1.483 | Hexadecane | C ₁₆ H ₃₄ | 226.44 | Antifungal, Antibacterial, and antioxidant activity [26] |
| 15. | 12.460 | 1.462 | Heptadecane, 9- hexyl- | C ₂₃ H ₄₈ | 324.6 | Antifungal [27] |
| 16. | 12.460 | 1.462 | Heptadecane, 9- octyl- | C25H52 | 352.7 | Medical ultrasound diagnostic system [28] |
| 17. | 12.788 | 1.233 | Butabarbital | $\begin{array}{c} C_{10}H_{16}N_2\\ O_3 \end{array}$ | 212.25 | Analgesic agent [29] |
| 18. | 13.048 | 1.824 | Octadecane, 1- chloro- | C ₁₈ H ₃₇ Cl | 288.9 | Antioxidant, antimicrobial activity [30] |
| 19. | 13.217 | 1.197 | tert- Hexadecanethiol | C ₄₈ H ₉₉ Au S ₃ | 969.5 | Diabetes, inflammation, cancer [31] |
| 20. | 13.217 | 1.197 | Geranylisovalerat e | C ₁₅ H ₂₆ O ₂ | 238.37 | Anti- inflammatory, antioxidant, and anti-viral activities [32] |
| 21. | 13.217 | 1.197 | 17- Pentatriacontene | C35H70 | 490.9 | Antimicrobial activity [33] |
| 22. | 13.217 | 1.197 | Tetrapentacontan e, 1,54-dibromo- | C ₅₄ H ₁₀₈ Br ₂ | 917.2 | antibacterial activity [34] |
| 23. | 13.217 | 1.197 | 1-Hexadecanol, 2-methyl- | C ₁₇ H ₃₆ O | 256.5 | antimicrobial, antioxidant activity [35] |

| 24. | 13.217 | 1.197 | E-8-Methyl-9- tetradecen-1-ol acetate | C ₁₇ H ₃₂ O ₂ | 268.4 | Insect pheromone [36] |
|-----|--------|-------|---|--|--------|---|
| 25. | 13.217 | 1.197 | Ethanol, 2- (octadecyloxy)- | C ₂₀ H ₄₂ O ₂ | 314.5 | antibacterial activity [37] |
| 26. | 13.217 | 1.197 | 5-Octadecenal | C ₁₈ H ₃₄ O | 266.5 | Flavoring agent, antimicrobial [38] |
| 27. | 13.355 | 2.562 | Nonane, 2- methyl-5-propyl- | C ₁₃ H ₂₈ | 184.36 | Anti-cancer [39] |
| 28. | 13.436 | 1.403 | tert- Hexadecanethiol | C ₁₆ H ₃₄ S | 258.5 | Antitumoral, antioxidant, insecticidal, antifungal [40] |
| 29. | 13.629 | 1.450 | cis-1-Chloro-9- octadecene | C ₁₈ H ₃₅ Cl | 286.9 | Skin protection and respiratory protection [41] |
| 30. | 13.629 | 1.450 | Bacteriochloroph yll-c-stearyl | C ₅₂ H ₇₂ Mg N4O4 ⁻² | 841.5 | lipoxygenaseinhibi tor, pesticide [42] |
| 31. | 13.629 | 1.450 | Acetic acid, chloro-, octadecyl ester | C ₂₀ H ₃₉ ClO 2 | 347 | (odor-forming compound) [43] |
| 32. | 13.629 | 1.450 | 17- Pentatriacontene | C ₃₅ H ₇₀ | 490.9 | Antiinflammatory Anticancer Antibacterial Antiarthritic [44] |
| 33. | 13.767 | 1.338 | 8-Octadecenal | C ₁₈ H ₃₄ O | 266.5 | anti-bacterial and anti-fungal activities [45] |
| 34. | 13.767 | 1.338 | 4-Octadecenal | C ₁₈ H ₃₄ O | 266.5 | Antimicrobial, Anti-inflammatory [46] |
| 35. | 13.922 | 1.659 | 1-Hexadecanol, 2-methyl- | C ₁₇ H ₃₆ O | 256.5 | antimicrobial, antioxidant [47] |
| 36. | 14.309 | 1.158 | Tetradecane | C ₁₄ H ₃₀ | 198.39 | Antimicrobial, Cytotoxicity, Antipyretic, Anthelmintic, Tumour, |

| | | | | | | Bronchitis, Asthma, Tuberculosis, Dyspepsia, Constipation, Anemia, Throat diseases, Elephantiasis, Antidiabetic, Anti- inflammatory, Antidiarrhoeal [48] |
|-----|--------|-------|---|---------------------------------|--------|---|
| 37. | 14.816 | 1.150 | Heptadecane | C ₁₇ H ₃₆ | 240.5 | anti-oxidative [49] |
| 38. | 14.816 | 1.150 | Eicosane, 7- hexyl- | C ₂₆ H ₅₄ | 366.7 | Anti-androgenic, aldose reductase inhibitor [50] |
| 39. | 15.005 | 1.447 | Heptadecane, 2,6,10,15- tetramethyl- | C ₂₁ H ₄₄ | 296.6 | Flavouring agent [51] |
| 40. | 15.005 | 1.447 | Octadecane, 3- ethyl-5-(2- ethylbutyl)- | C ₂₆ H ₅₄ | 366.7 | anti-oxidant and antiinflammatory effect [52] |
| 41. | 15.349 | 1.836 | Pentadecane | C ₁₅ H ₃₂ | 212.41 | Antibacterial activity [53] |
| 42. | 15.349 | 1.836 | Dodecane | C ₁₂ H ₂₆ | 170.33 | Antibacterial activity [54] |
| 43. | 15.349 | 1.836 | Heptadecane | C ₁₇ H ₃₆ | 240.5 | Antibacterial activity [55] |

4. Conclusion

The present study has characterized the bioactive components of *B. pinnatum* leaves Methanol extract to include phytochemicals such as carbohydrates, Starch, proteins, amino acid, Anthraquinones, Quinines, Glycosides, cardiac glycosides, Phenol, Tannins, Flavonoids, Phytosterols, Saponins, andSteroids. tert-Hexadecanethiol, Eicosane, 7-hexyl-, and Octadecane, 3-ethyl-5-(2-ethylbutyl)- were found to be predominant according to the GC-MS study. Due to the existence of these bioactive substances, *B. pinnatum* leaves are now recognized as a valuable source of prospective lead compounds that have both biological and pharmacological effects, making them a promising option for drug discovery.

Acknowledgment

We thankfully acknowledge the assistance provided by the entire staff of the Innovation Centre Bundelkhand University during the experimental work on the GC-MS, Rotary evaporator, Lyophilizer, and other Centre instruments.

References:

- 1. Narinderpal, K., Raman, B., Junaid, N. (2014). A Review on Bryophyllam pinnatum-A Medicinal Herb. Journal of Medical and Pharmaceutical Innovation, Vol. 1, Issue 3, 2014.
- 2. .Mbakwem-Aniebo C, Onianwa O, Okonko IO. Effect of Ocimum gratissimum Leaves on common Dermatophytes and Causative Agent of pityriais versicolor in Rivers state, Nigeria. Journal of Microbiology Research 2012; 2:108-113.
- 3. Swain T. Plants in the development of modern medicine. Harvard University Press; 1968.
- 4. Fokunang CN, Ndikum V, Tabi OY, Jiofack RB, Ngameni B, Guedje NM, et al. Traditional medicine: past, present and future research and development prospects and integration in the National Health System of Cameroon.African journal of traditional, complementary, and alternative medicines. 2011;8(3);284–295. Available:https://doi.org/10.4314/ajtcam.v8i 3.65276.
- 5. John JN, Kasenene M, Bernard T, Kiremire B, Robert Kamatenesi-Mugisha BM, Krief S, Dumontet V, kabasa JD. Traditional Plants used for Medicinal Purpose by Local Communities around the Northern sector of kibale National Park, Uganda. Journal of Ethnopharmacology 2011; 136:236-245.
- 6. Jain VC. Antioxidant and Antimicrobial Activities of Bryophyllum caltanum, salisb Leaf. Pharmacology online 2010; 1:393-405.
- 7. Lans CA. Ethnomedicine Used in Trinidad and Tabago for Urinary Problems and Diabetes mellitus. Journal of Ethnobiology and Ethnomedicine 2006; 2:45.
- 8. Bagul MS, Srinivasa H, Anandjiwala S, Rajani M. Phytochemical evaluation and free radical scavenging activity of Nagakesara (Stamen of Mesua ferrea Linn, Var. Ferrea). Indian drugs. 2006;43:665-670.
- Anjoo, K. and Ajay K. S (2009). Bryophyllum pinnatum (Lam.) Kurz: Phytochemical and pharmacological profile: A review. A publication of phcog. Net Pharmacognosy Review. (3) 6: 364-374.
- 10. Harborne A. Harborne, J.B. (1998) Textbook of Phytochemical Methods. A Guide to Modern Techniques of Plant Analysis. 1998;317.
- 11. Sharma S, Kumar R. Antioxidant Activity , TLC and Phytochemical Analysis of Ginger (Zingiber officinale L.) Rhizome. Plant Arch. 2018;18:210–4.
- 12. Kushwaha, P., Alok, S., & Dwivedi, L. (2021). The GC-MS analysis of methanolic extract of Chlorophytum borivilianum and compounds' activities validation at standard databases. *South Asian Journal of Experimental Biology*, *11*(6), 768-774.
- 13. Lexa GM, Josphnat C, Josphat C, Matasyoh FN, Minam GK, Anne WT, Titus KM. Chemical Composition and Antimicrobial Activity of the Essential Oil of Ocimum gratissimum L.

Growing in Eastern Kenya. African Journal of Biotechnology 2007; 6(6):760-765

- 14. Qanash, H., Yahya, R., Bakri, M.M. *et al.* Anticancer, antioxidant, antiviral and antimicrobial activities of Kei Apple (*Dovyalis caffra*) fruit. *Sci Rep* **12**, 5914 (2022).
- 15. Adesalu, T. A., Temenu, T. O., & Julius, M. L. (2016). Molecular characterization, lipid analysis and GC-MS determination of bioactive compounds identified in a West African strain of the green alga Oedogonium (Chlorophyta). *Journal of Pharmacognosy and Phytochemistry*, 5(6), 01-06.
- 16. Addai, Z. R., Abood, M. S., & Hlail, S. H. (2022). GC-MS profiling, antioxidants and antimicrobial activity of prickly pear (Opuntiaficus-indica) pulp extract. *Pharmacognosy Journal*, *14*(2).
- Naz, R., Roberts, T. H., Bano, A., Nosheen, A., Yasmin, H., Hassan, M. N., ... & Anwar, Z. (2020). GC-MS analysis, antimicrobial, antioxidant, antilipoxygenase and cytotoxic activities of Jacaranda mimosifolia methanol leaf extracts and fractions. *PLoS One*, *15*(7), e0236319.
- 18. Kumar, D., Karthik, M., & Rajakumar, R. (2018). GC-MS analysis of bioactive compounds from ethanolic leaves extract of Eichhornia crassipes (Mart) Solms. and their pharmacological activities. *Pharma Innov J*, 7(8), 459-462.
- Idris, O. A., Wintola, O. A., & Afolayan, A. J. (2019). Comparison of the proximate composition, vitamins (ascorbic acid, α-tocopherol and retinol), anti-nutrients (phytate and oxalate) and the GC-MS analysis of the essential oil of the root and leaf of Rumex crispus L. *Plants*, 8(3), 51.
- Al-Qurainy, F., Tarroum, M., Khan, S., Nadeem, M., Gaafar, A. R. Z., Alansi, S., & Alfarraj, N. S. (2022). Genome estimation and phytochemical compound identification in the leaves and callus of Abrus precatorius: A locally endangered plant from the flora of Saudi Arabia. *Plants*, *11*(4), 567.
- 21. Konovalova, O., Gergel, E., & Herhel, V. (2013). GC-MS Analysis of bioactive components of Shepherdia argentea (Pursh.) Nutt. from Ukrainian Flora. *The Pharma Innovation*, 2(6, Part A), 7.
- 22. Parijatham, S., Sharmila, D., Prabhu, K., Raghavandra, R., Rao, M. R. K., Dinakar, S., & Sundaram, R. L. (2020). The gas chromatography-mass spectrometry analysis of one Ayurvedic formulation, Srikhandasavam. *Drug Invention Today*, *14*(5).
- 23. Ganesh, M., & Mohankumar, M. (2017). Extraction and identification of bioactive components in Sida cordata (Burm. f.) using gas chromatography-mass spectrometry. *Journal of food science and technology*, *54*, 3082-3091.
- 24. Sanjaya, P., & Lekha, K. (2019). Preliminary phytochemical analysis, GC-MS studies and antioxidant activity of Majidea zangueberica J. Kirk leaf extracts. *J Med Plants Stud*, 7(2), 186-195.
- 25. Khan, S., Kaur, H., & Jhamta, R. (2019). Evaluation of antioxidant potential and phytochemical characterization using GC-MS analysis of bioactive compounds of Achillea filipendulina (L.) leaves. *Journal of Pharmacognosy and Phytochemistry*, 8(3), 258-265.
- 26. Akpuaka, A., Ekwenchi, M. M., Dashak, D. A., & Dildar, A. (2013). Biological activities of characterized isolates of n-hexane extract of Azadirachta indica A. Juss (Neem) leaves. *Nature and science*, *11*(5), 141-147.
- Alqahtani, S. S., Moni, S. S., Sultan, M. H., Bakkari, M. A., Madkhali, O. A., Alshahrani, S., *Eur. Chem. Bull.* 2023,12(Special issue 12), 2075-2084
 2086

... & Sayed-Ahmed, M. Z. (2022). Potential bioactive secondary metabolites of Actinomycetes sp. isolated from rocky soils of the heritage village Rijal Alma, Saudi Arabia. Arabian Journal of Chemistry, 15(5), 103793.

- 28. Patil, K., & Singh, D. M. (2022). GC-MS Analysis of fresh water Cylindrospermum sp. PCC518, Cylindrospermum sp. PCC 567 ethanol and hexane extracts. Int. J. Herb. Med, 10, 15-25.
- 29. Nischitha, R., Vasanthkumari, M. M., Kumaraswamy, B. E., & Shivanna, M. B. (2020). Antimicrobial and antioxidant activities and chemical profiling of Curvularia tsudae endophytic in Cynodon dactylon (L.) Pers. 3 Biotech, 10, 1-12.
- 30. Kumar, A., Mahanty, B., Goswami, R. C. D., Barooah, P. K., & Choudhury, B. (2021). In vitro antidiabetic, antioxidant activities and GC-MS analysis of Rhynchostylis Retusa and Euphorbia Neriifolia leaf extracts. 3 Biotech, 11, 1-10.
- 31. Sahu, M. K., & Singh, G. Structural identification through GC mass spectrophotometer and determine anti lithiotic activity of hibiscus rosa sinensis by using ethylene glycol induced method.
- 32. Hugar, A. L., Kanjikar, A. P., & Londonkar, R. L. (2017). Bioactive compounds investigation from methanol bark extract of Pterocarpus marsupium using GC-MS analysis. International Journal of Pharmaceutical Quality Assurance, 8(3), 104-110.
- 33. Alghamdi, A. I., & Ababutain, I. M. (2019). Research article phytochemical screening and antibacterial activity of Eucalyptus camaldulensis's leaves and bark extracts. Asian Journal of Scientific Research, 12(2), 202-210.
- 34. Oviya, R., Thiruvudainambi, S., Ramamoorthy, V., & Vellaikumar, S. (2022). Antagonistic potential of Trichoderma hamatum against Alternaria porri causing purple blotch disease of onion through Gas chromatography-mass spectrometry (GCMS) analysis. Journal of Applied and Natural Science, 14(3), 1031-1038.
- 35. Jaddoa, H. H., Hameed, I. H., & Mohammed, G. J. (2016). Analysis of volatile metabolites released by Staphylococcus aureus using gas chromatography-Mass spectrometry and determination of its antifungal activity. Oriental Journal of Chemistry, 32(4), 8-24.
- 36. Ramalakshmi, S., & Muthuchelian, K. (2011). Analysis of bioactive constituents from the of Mallotus tetracoccus (Roxb.) Kurz, by gas chromatography-mass leaves spectrometry. International Journal of Pharmaceutical Sciences and Research, 2(6), 1449.
- 37. Abdullah, R. R. (2019). Insecticidal activity of secondary metabolites of locally isolated fungal strains against some cotton insect pests. Journal of Plant Protection and Pathology, 10(12), 647-653.
- 38. Efiong, E. E., Kwon-Ndung, E. H., Ubhenin, A. E., & Nwaedozie, J. M. Identification of Volatile Compounds in Sweetened Hibiscus Sabdariffa Drink.
- 39. Al-Qurainy, F., Tarroum, M., Khan, S., Nadeem, M., Gaafar, A. R. Z., Alansi, S., & Alfarraj, N. S. (2022). Genome estimation and phytochemical compound identification in the leaves and callus of Abrus precatorius: A locally endangered plant from the flora of Saudi Arabia. Plants, 11(4), 567.
- 40. Chirumamilla, P., & Taduri, S. (2023). GC-MS Fingerprinting and Antibacterial Activity of Solanum khasianum Stem and Fruit Extracts. Proceedings of the National Academy of Sciences, India Section B: Biological Sciences, 1-11. Eur. Chem. Bull. 2023, 12 (Special issue 12), 2075-2084 2087

- 41. Shankari, C., Prabhu, K., & Rao, M. R. K. The GC-MS Study of One Digestive Ayurvedic Powder, Eladi Churnam.
- 42. Madhavan, S. A. (2021). In-vitro pharmacological and Gc-Ms analysis of bioactive compounds presents in Bytteneria herbaceae. *International Journal of Zoological and Entomological Letters*, 1(1), 20-26.
- 43. Rathinamoorthy, R., & Thilagavathi, G. (2016). GC-MS analysis of worn textile for odour formation. *Fibers and polymers*, *17*, 917-924.
- 44. Kumar, D., Karthik, M., & Rajakumar, R. (2018). GC-MS analysis of bioactive compounds from ethanolic leaves extract of Eichhornia crassipes (Mart) Solms. and their pharmacological activities. *Pharma Innov J*, 7(8), 459-462.
- 45. Hussein, H. M., Hameed, I. H., & Ubaid, J. M. (2016). Analysis of the secondary metabolite products of ammi majus and evaluation anti-insect activity.
- 46. Prabakaran, R., Kumar, T. S., & Rao, M. V. (2014). GC-MS Analysis and In vitro Cytotoxicity Studies of Root Bark Exudates of Hardwickia binata Roxb. *methods*, *8*, 12.
- 47. Jaddoa, H. H., Hameed, I. H., & Mohammed, G. J. (2016). Analysis of volatile metabolites released by Staphylococcus aureus using gas chromatography-Mass spectrometry and determination of its antifungal activity. *Oriental Journal of Chemistry*, *32*(4), 8-24.
- 48. Banakar, P., & Jayaraj, M. (2018). GC-MS analysis of bioactive compounds from ethanolic leaf extract of Waltheria indica Linn. and their pharmacological activities. *Int. J. Pharm. Sci. Res*, *9*(5), 2005-10.
- Kim, D. H., Park, M. H., Choi, Y. J., Chung, K. W., Park, C. H., Jang, E. J., ... & Chung, H. Y. (2013). Molecular study of dietary heptadecane for the anti-inflammatory modulation of NF-kB in the aged kidney. *PloS one*, 8(3), e59316.
- 50. Khatua, S., Pandey, A., & Biswas, S. J. (2016). Phytochemical evaluation and antimicrobial properties of Trichosanthes dioica root extract. *Journal of Pharmacognosy and Phytochemistry*, 5(5), 410-413.
- Krishnamoorthy, C., Uma, S. K., Jadhav, P. P., Ghazal, K., & Chidambaram, R. (2018). Gas Chromatography/Mass Spectroscopy Analysis Of Phytochemicals Present In Orange Peel Powder And In Bread Prepared Using It. *Asian Journal Of Chemistry*, 30(7), 1599-1602.
- 52. Al-Marzoqi, A. H., Hameed, I. H., & Idan, S. A. (2015). Analysis of bioactive chemical components of two medicinal plants (Coriandrum sativum and Melia azedarach) leaves using gas chromatography-mass spectrometry (GC-MS). *African Journal of Biotechnology*, *14*(40), 2812-2830.
- 53. Konovalova, O., Gergel, E., & Herhel, V. (2013). GC-MS Analysis of bioactive components of Shepherdia argentea (Pursh.) Nutt. from Ukrainian Flora. *The Pharma Innovation*, 2(6, Part A), 7.
- Padma, M., Ganesan, S., Jayaseelan, T., Azhagumadhavan, S., Sasikala, P., Senthilkumar, S., & Mani, P. (2019). Phytochemical screening and GC–MS analysis of bioactive compounds present in ethanolic leaves extract of Silybum marianum (L). *Journal of drug delivery and therapeutics*, 9(1), 85-89.
- 55. Faridha Begum, I., Mohankumar, R., Jeevan, M., & Ramani, K. (2016). GC–MS analysis of bio-active molecules derived from Paracoccus pantotrophus FMR19 and the antimicrobial activity against bacterial pathogens and MDROs. *Indian journal of microbiology*, 56, 426-*Eur. Chem. Bull.* 2023,12(Special issue 12), 2075-2084

432.