IN VITRO ANTIMICROBIAL ACTIVITY OF THE STEMS AND LEAF EXTRACTS OF Paederia foetida Linnaeus (Kantutai), family

Rubiaceae

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Abstract

Reportedly, *Poederia foetida* locally known as 'kantutai' has long been used in traditional medicine in the Philippines in the treatment of rheumatism, ulcers, diarrhea, dysentery, inflammation of the spleen, indigestion and flatulence. However, these reports were anecdotal, thus the need to provide scientific basis to traditional healing practices of marginalized and geopgraphically isolated communities. This study aimed to investigate the antimicrobial properties of *P. foetida* extracts from stems and leaves against *S. aureus*, *E. coli*, and *C. albicans*. The extracts exhibited antimicrobial activity against bacteria but not against the fungus. Ethyl alcohol extracts showed larger inhibition zones compared to ethyl acetate extracts. Phytochemical screening revealed the presence of alkaloids, saponins, and tannins. Further research is suggested to optimize extraction protocols and explore the full potential of *P. foetida* as an antibacterial agent.

Keywords: antimicrobial, extract, Candida albicans, Escherichia coli, Paederia foetida, Staphylococcus aureus

1. Introduction

Paederia foetida, a plant species known for its unpleasant odor resembling sulfur or fetid aroma, possesses various common names such as *kantutai or otot-bagtik* in Northern Samar. This unique smell is attributed to the presence of sulfur compounds, particularly dimethyl disulfide, in the oils of its leaves and stems. Belonging to the Rubiaceae family, this herbaceous plant features slender vines that can be hairy or smooth, allowing it to climb over shrubs and trees and reach the canopy. Notably, the leaves of *Paederia foetida* are rich in carotene, vitamin C, and proteins with various essential amino acids.

The versatile leaves of this plant have multiple medicinal uses. They are effective in treating gastrointestinal issues like diarrhea, dysentery, and flatulence-related distension. Additionally, they can alleviate intestinal problems such as cramps and colic dysentery, and external application can aid in healing bruises and reducing swelling. *Paederia foetida* leaves are also beneficial for nose ulcerations, earaches, swollen eyes, and abdominal swelling. Bathing with crushed leaves can provide anti-rheumatic treatment, and leaf decoctions are helpful in relieving urinary retention and urinary bladder stones. Moreover, the decocted leaves can be used internally or externally for fever relief.

The roots of this plant have emetic properties and are utilized in the treatment of piles, spleen inflammation, chest pain, and liver discomfort. The fruits are used for toothaches and restoring the color of darkened teeth, while the bark is known for its effects on constipation, asthma, urethral calculi, and aiding in placenta removal after a miscarriage.

Paederia foetida has been extensively studied for its therapeutic properties. It has shown antioxidant, antinociceptive, antiviral, cough suppressant, anti-cancer, antidiarrheal, anti-arthritic, analgesic, anti-helminthic, anti-inflammatory, hepatoprotective, and testosterone-boosting effects. The plant has also demonstrated positive results in the treatment of colitis, shigellosis, and chemolithotripsy. However, further research is needed to explore its antibacterial activity.

With this, *Paederia foetida* proves to be a remarkable plant with diverse applications in traditional medicine, highlighting its potential as a valuable natural resource for various health conditions. Hence, this study will try to evaluate its antimicrobial properties.

2. METHODOLOGY

Sample preparation

Fresh leaves of *P. foetida* from Palapag, Northern Samar, underwent taxonomic authentication at the Natural History Museum, Philippines. Around 150g of leaves were washed, dried, and manually juiced. The dried material was mixed with a 1:2 ratio of ethyl alcohol and ethyl acetate and left to sit for 24 hours at 32°C in an oven.

The mixture of extract and solvent underwent filtration using cheesecloth and Whattman no 41mm filter paper. The filtered extract was then distilled to remove the solvent. After distillation, the resulting extract was transferred to a sterilized bottle and labeled for identification.

The detection of secondary metabolites was carried out using the protocols of Guevara (2005), and some from the study Dagalea *et al.* (2022).

The test for the presence of alkaloids was done with the reaction of dragendorff's reagent and mayer's reagent. The test for the presence of saponins was determined using capillary tube. The test for the presence of flavonoids was performed using a test tube screening technique. Lastly, the test for the presence of tannins was performed using the plant sample and was centrifuged.

Preparation of culture media and sensitivity discs

To prepare bacteria culture media, 19g of Nutrient Agar (NA) was dissolved in 500mL distilled water. For fungi culture media, 9.5g of Potato Dextrose Agar (PDA) was dissolved in 250mL water. The mixtures were agitated and sterilized at 121°C and 15 psi. After cooling to 50°C, the media was dispensed into petri dishes and solidified at room temperature.

Round discs for sensitivity testing were created by using filter paper (Whatman No.41) and a puncher. Each disc was then positioned in a petri dish and subjected to pressure sterilization at 121°C and 15 psi pressure. Subsequently, the discs were soaked in the crude extracts of *P. foetida*.

Antimicrobial Screening

The Kirby Bauer antibacterial sensitivity test was used to evaluate the antimicrobial activity of *P. foetida* extract against *S. aureus* and *E. coli*. Positive controls, chloramphenicol and tetracycline, were used. Bacterial inoculation was done on sterile culture media plates, and *P. foetida* extract-soaked filter paper discs were placed in the center. Plates were incubated at

37°C for 24 hours. Measurement of the zone of inhibition using a vernier caliper indicated the inhibition of microorganism growth. The absence of a zone indicated resistance to *P. foetida* extract.

3. RESULTS

Phytochemical screening

Table 1 reveals the presence of tannins, alkaloids, and saponins in the leaf extracts of P. foetida. However, regardless of the solvent used, flavonoids were not detected in the extracts. Notably, saponins and alkaloids were exclusively present in the extracts obtained using ethyl alcohol as the solvent.

Table 1. Chemical Presence of the Extract from the Leaf of *Paederia foetida*

Chemical	Leaf Extract	
properties	Ethyl	Ethyl
	alcohol	acetate
Alkaloids	+	-
Flavonoids	-	-
Saponins	+	-
Tannins	+	+

Legend: (+) = present; (-) = not detected

Antimicrobial Property

In Table 2, ethyl alcohol-based extracts showed larger zones of inhibition for *S. aureus* and *E. coli*. The extracts exhibited inhibitory activity only against bacteria, with no significant difference in antimicrobial activity between leaf extracts against *E. coli* and *S. aureus*.

The study found no significant difference in antimicrobial resistance or activity between the solvents used. Both solvents exhibited significant antimicrobial activity, indicating their suitability as sources for inhibiting or killing microorganisms.

Table 2. Zone of inhibition (mm) formed in Nutrient Agar with crude extracts from leaves of *P. foederia*.

*chlorampenicol / **tetracyline			
Test microorganism			
S. aureus	E. coli		
22.70 mm	21.00		
	mm		
13.95 mm	14.20		
	mm		
21.00*			
mm	19.70**		
	mm		
	Test microo S. aureus 22.70 mm 13.95 mm 21.00*		

4. DISCUSSION

Positive results were obtained for the presence of alkaloids, tannins, and saponins in the ethyl alcohol leaf extracts of *Paederia foetida*. The considerable structural diversity of these natural compounds discovered in *P. foetida* offers promising opportunities for the exploration of new drugs. Regarding the antibacterial activity of the extract, no significant relationship was found between the extract's effect and the microorganisms utilized in the study. However, the leaf extracts of *P. foetida* exhibited notable antimicrobial resistance and activity, suitability as sources for inhibiting or killing microorganisms.

P. foetida leaf extracts demonstrated the ability to inhibit the growth of *E. coli* and *S. aureus*, suggesting its potential as an organic antibacterial source. Further research is recommended to test the extracts against other bacteria and fungi to assess their susceptibility.

5. ACKNOWLEDGMENT

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6. CONFLICT OF INTEREST

None.

References

- [1] Afroz S, Alamgir M, Khan MTH, Jabbar S, Nahar N and MSK Choudhuri. 2006. Antidiarrhoel activity of the ethanol extract of *Paederia foetida* Linn. (Rubiaceae). Journal of Ethnopharmacol, 105:125–30.
- [2] Barua U, Hore DK and R. Sarma. 2007. Wild edible plants of Ajuli Island and Darrang districts of Assam. Indian Journal of Traditional Knowledge, 6:191–194.
- [3] Bauer AW, Kirby MM, Sherris JC and M Turck. 1996. Antibiotic susceptibility testing by a standardized single disk method. The American Journal of Clinical Pathology 45(4).
- [4] Borah PK, Gogoi P, Phukan AC, Mahanta J. 2006. Traditional medicine in the treatment of gastrointestinal diseases in upper Assam. Indian Journal of Traditional Knowledge, 5: 510–512.
- [5] Chanda S, Sarethy IP, De B and K Singh. 2013. *Paederia foetida-* a promishing ethnomedicinal tribal plant of Northeast India. Journal of Forestry Research, 24:801–8.
- [6] Chanda S, De B, Tiwari RK. 2011. Traditional and ethnobotanical investiga- tion of some edible plants among the tribes of Tripura, India. In: M.D. Choudhury, G.D Sharma, S. Choudhury, A.D. Talukdar (eds), Status and conservation of biodiversity in North East India. Delhi: Swastik publication, pp. 118–124.
- [7] Chopra, R.N., Nayar, L.C., Chopra, L.C. 1956. Glossary of Indian Medicinal Plants. Council of Scientific and Industrial Research. New Delhi India.
- [8] Chopra, R.N., Chopra, I.C. Verma, B.S., Supplement to Glossary of Indian Medicinal Plants. Council of Scientific and Industrial Research. New Delhi India.
- [9] Dagalea, F. M. S., Jr., A. A. U. F., Cortez, F. E., & Lim, K. M. C. (2022). Secondary Metabolite Screening of Extracted Oil from Nypa fruticans Wurmb. (Nipa Palm). *Asian Plant Research Journal*, *9*(3), 17–24. https://doi.org/10.9734/aprj/2022/v9i330208

- [10] Das P. 1997. Wild tribal plants of Tripura tribes. Agartala: Tripura Tribal Cultural Research Institute & Museum Government of Tripura, p. 57–58
- [11] Das S, Kanodia L, Mukherjee A, Hakim A., 2013. Effect of ethanolic extract of leaves of Paederia foetida Linn. on acetic acid induced colitis in albino rats. Indian J Pharmacol.
- [12] Dasgupta N, De B. 2007. Antioxidant activity of some leafy vegetables of India: A comparative study. Food Chemistry, 101: 471-474.
- [13] De S, Shukla VJ, Ravishankar B, Bhavsar GC. 1996. A preliminary study on the hepatoprotective activity of methanol extract of Paederia foetida leaf. Fitoterapia, 67: 106–109.
- [14] De S, Ravishankar B and GC Bhavsar. 1994. Investigation of the anti–inflammatory effects of *Paederia foetida*. Journal of Ethnopharmacology, 43(1):31–38.
- [15] De S, Ravishankar B and GC Bhavsar. 1993. Evaluation of *Paederia foetida* for hepatoprotective and anti–inflammatory activities. Indian Journal of Natural Product, 9:7–11.
- [16] Dey YN and MK Pal. 2011. Evaluation of anti–helmintic activity of leaves of *Paederia foetida*. International Journal of Pharma Bio Sci, 2(1):227–231.
- [17] Duke, J. A. 2002. Handbook of medicinal herbs. CRC Press: Boca Raton, Florida; 2nd edition, pp 896.
- [18] Ghani A. 2003. Medicinal plants of Bangladesh with chemical constituents and uses. 2nd edition. Asiatic Society of Bangladesh; 331–33.
- [19] Guevara BQ. 2005. A Guidebook to Plant Screening: Phytochemical and Biological. Research Center for the Natural Sciences and University of Santo Tomas Publishing House, Beato Angelico Bldg. Espana, Manila.
- [20] Haider R, Khan AK and KM Aziz. 1991. Evaluation of indigenous plants in the treatment of acute shigellosis. Trop Geogr Med. 43(3):266–270.
- [21] Hossain M, Mohammad SA, Saha A, Alimuzzaman M. 2006. Antinociceptive activity of whole plant extracts of Paederia foetida. Journal of Pharmaceutical Sciences, 5: 67–69.
- [22] Kalita D and B Phukan. 2010. Some Ethnomedicine used by the Tai Ahom of Dibrugarh district, Assam, India. Indian Journal of Natural Product, 1:507–11.
- [23] Kar A, Borthakur SK. 2008a. Wild vegetables of Karbi-Analog district, Assam. Natural Product Radiance, 7: 448–460.
- [24] Kar A, Borthakur SK. 2008b. Medicinal plants used against dysentery, diarrhea and cholera by the tribes of erstwhile Kameng district of Arunachal Pradesh. Natural Product Radiance, 7: 176–181.
- [25] Langeland, K. A. and K. Craddock Burks (eds.) 1998. Identification and Biology of Non-native Plants in Florida's Native Areas. University of Florida, Gainesville, Florida, USA
- [26] Nandkoni KM (Ed) 2002. Indian Materia Medica. Mumbai: Bombay Popular Prakashan: 892.
- [27] Osman H, Rahim AA and NM Isa. 2009. Antioxidant activity and phenolic content of *Paederia foetida* and *Syzygium aqueum*. Molecules, 14(3):970–978. 17.
- [28] Prakash V, Mishra PK, Mishra M. 2009. Screening of medicinal plant extracts for

- antioxidant activity. Journal of Medicinal Plants Research, 3: 608–612.
- [29] Raghunathan K and R Mitra. 1982. Pharmacognosy of indigenous drugs vol 2. Central Council for Research in Ayurveda and Siddha: New Delhi, India.
- [30] Rajashekhara N, Sharma PP and P Vasanth. 2009. Comparative study of Prasarni [Merremia tridentate Hallier.f. (Convulvulaceae) and Paederia foetida (Rubiaceae)] in Amavata (Rheumatoid arthritis). 30:503-507
- [31] Ravi V, Saleem TSM, Patel SS, Raamamurthy J, Gauthaman K. 2009. Anti-inflammatory effect of methanolic extract of Solanum nigrum Linn berries. International Journal of Applied Research in Natural Products, 2: 33–36.
- [32] Srivastava RC, Singh RK. 2010. Apatani community, Mukherjee TK. Indigenous biodiversity of Apatani plateau: Learning on biocultural knowledge of Apatani tribe of Arunachal Pradesh for sustainable live hoods. Indian Journal of Traditional Knowledge, 9: 432–442
- [33] de Tavera, T.H. Pardo, 2000. "(Plantas Medicinales de Filipinas) Medicinal Plants of the Philippines". (Makati City: Ayala Foundation Inc., Filipinas Heritage Library, Makati Avenue, ayala Triangle, 59
- [34] The Ayurvedic Pharmacopeia of India (1999). Ministry of Health and Family Welfare Department and Indian System of medicine and Homeopathy, New Delhi. Part I, Vol II: 137 –140.
- [35] The Wealth of India. 2001. A dictionary of Indian raw materials and industrial product, raw materials, vol. 7. New Delhi: Council for Scientific and Industrial Research (CSIR).
- [36] Thompson KD, Ather A., 2005. Antiviral activity of three Bangladeshi medicine plant extracts against herpes simplex viruses. Antiviral Res, 67:107–115
- [37] Wong, K. C.; Tan, G. L. 1994. Steam volatile constituents of the aerial parts of Paederia foetida L. Flavour Fragr. J; 9 (1): 25–28.
- [38] Yang LL, Yen KY, Kiso Y, Kikino H. 1987. Antihepatotoxic actions of Formosan plant drugs. Journal of Ethnopharmcology, 19: 103–110.