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**From DNA to Proteins: Investigating Gene Expression at The Biochemical Level****Dr Sangeeta Bhimrao Dongre<sup>a</sup>, Dr.R.R.Kumar<sup>b</sup>, Dr. Gaviraj.E.N<sup>c</sup>,  
Dr. Khushal N. Pathade<sup>d</sup>****Article History: Received: 10.05.2023****Revised: 29.05.2023****Accepted: 09.06.2023****Abstract**

In all living things, gene expression converts DNA into functional proteins. Understanding gene expression pathways is essential to understanding life and has major consequences for medicine, biotechnology, and evolutionary biology. This review discusses gene expression biochemical processes and important molecular actors. We study RNA transcription, post-transcriptional changes, and protein translation. We also address gene expression regulation and biological methods. Understanding gene expression pathways will progress molecular biology and enable targeted medicines and biotechnological uses.

**Keywords:** *Gene expression, Transcription, Post-transcriptional Modifications, Translation, Protein Modifications,*

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## Introduction

All living things depend on gene expression, which converts DNA into functional proteins. Gene expression converts genetic instructions into proteins for metabolism, growth, differentiation, and environmental response. Molecular biology, genetics, medicine, biotechnology, and evolutionary biology all need to understand gene expression. Researchers can understand life's basic processes, the molecular foundation of illnesses, innovative therapies, and organism engineering by deciphering gene expression.

The field of gene expression research has a rich historical context, marked by several key discoveries that have shaped our understanding of this intricate process. One of the ground-breaking achievements was the elucidation of the structure of DNA by James Watson and Francis Crick in 1953, which laid the foundation for understanding how genetic information is stored and transmitted. This discovery paved the way for subsequent investigations into gene expression.

The genetic code revealed the relationship between DNA nucleotides and amino acids in the 1960s. This discovery, together with Sydney Brenner, Francois Jacob, and Matthew Meselson's discovery of transfer RNA (tRNA), proved RNA's importance in protein synthesis and illuminated translation. In the 1960s, Severo Ochoa and Marianne Grunberg-Manago discovered RNA polymerase, which was crucial to understanding transcription. This created the groundwork for transcriptional regulation and gene expression transcription factor research.

RNA capping, splicing, and polyadenylation, which affect RNA stability and protein production, have been better understood in recent decades. PCR and DNA sequencing quantified and profiled RNA molecules, revolutionising gene expression studies. High-throughput methods like microarrays and RNA sequencing have enabled genome-wide

gene expression pattern research, revealing gene expression regulation and dynamics in many biological situations. Single-cell genomics and transcriptomics have shown cellular heterogeneity and gene expression changes throughout development, disease progression, and therapy response.

As we continue to unravel the complexities of gene expression, there is a growing realization of its clinical significance. Gene expression profiling has become a valuable tool in diagnosing diseases, predicting patient outcomes, and guiding personalized medicine approaches. Additionally, gene expression studies have contributed to the development of biotechnological applications, such as the production of recombinant proteins, genetic engineering of crops, and the development of gene therapies. In this review article, we will delve into the biochemical processes underlying gene expression, including transcription, post-transcriptional modifications, translation, and the regulatory mechanisms that control gene expression. We will also explore the techniques and technologies used to investigate gene expression at the biochemical level. By comprehensively understanding the intricacies of gene expression, we can unlock new avenues for research, innovation, and the improvement of human health [1].

### Transcription: DNA TO RNA

#### *Transcription overview*

Gene expression requires transcription of DNA into RNA molecules. Eukaryotic and prokaryotic cells undergo this procedure. Transcription produces mRNA, tRNA, and rRNA, which serve diverse cell roles.

#### *RNA polymerase/transcription factors*

RNA polymerase synthesises RNA from DNA. In prokaryotes, a single RNA polymerase complex synthesises all RNA, whereas in eukaryotes, various RNA polymerases (I, II, and III) transcribe distinct classes of genes. Transcription factors influence RNA polymerase to regulate gene expression. Promoter regions upstream of a gene bind transcription factors. RNA polymerase and the

transcription start complex may promote or suppress transcription.

### ***Initiation, elongation, and termination of transcription***

Transcription can be divided into three main stages: initiation, elongation, and termination.

#### ***Initiation***

At gene promoters, RNA polymerase and transcription factors assemble during initiation. Transcription initiation complex. The initiation complex in prokaryotes is the RNA polymerase holoenzyme, whereas in eukaryotes it is RNA polymerase II and general transcription factors [2].

#### ***Elongation***

After initiation, RNA polymerase unwinds the DNA helix, revealing the template strand. By adding DNA-complementary nucleotides, it synthesises RNA from 5' to 3'. RNA polymerase continuously elongates the RNA transcript by unwinding and rewinding the DNA template.

#### ***Termination***

Intrinsic and Rho-dependent transcription termination occur in prokaryotes. Intrinsic termination occurs when the freshly synthesised RNA forms a hairpin loop and uracil (U) residues, dissociating RNA polymerase from the DNA template. Rho-dependent transcription termination needs Rho protein interaction to the RNA transcript. Termination in eukaryotes is more complicated and requires termination factors and DNA termination sequences.

### **FACTORS INFLUENCING TRANSCRIPTION EFFICIENCY**

***Several factors influence the efficiency of transcription***

#### ***Promoter strength***

The strength of the promoter sequence determines how often transcription is initiated. Strong promoters result in more frequent transcription, while weak promoters lead to less frequent transcription.

### ***Transcription factors***

The availability and activity of transcription factors play a critical role in regulating transcription. Activator proteins enhance transcription by promoting the assembly of the transcription initiation complex, while repressor proteins inhibit transcription by interfering with the binding of RNA polymerase [3].

#### ***Chromatin structure***

The accessibility of the DNA template is influenced by the organization of chromatin. Tighter packaging of DNA in nucleosomes can hinder transcription, while more open chromatin allows for easier access to the DNA template.

#### ***Epigenetic modifications***

Chemical modifications of DNA and histone proteins, such as DNA methylation and histone acetylation, can impact transcriptional activity. Methylation of DNA at specific sites is often

#### ***Post-transcriptional Modifications***

#### ***RNA processing: capping, splicing, and polyadenylation***

Following transcription, newly synthesized RNA undergoes several post-transcriptional modifications, collectively known as RNA processing. These modifications are essential for maturation, stability, and functionality of the RNA molecules.

#### ***Capping***

7-methylguanosine caps mRNA's 5' end in eukaryotes. This cap structure helps transfer mRNA from the nucleus to the cytoplasm and prevents degradation. The cap helps ribosomes recognise mRNA during translation.

#### ***Splicing***

Most eukaryotic protein-coding genes have introns. Splicing removes introns from pre-mRNA and joins coding areas. The spliceosome—a combination of snRNPs and other proteins—performs this activity. Alternative splicing generates several mRNA isoforms from a gene, boosting protein diversity.

#### ***Polyadenylation***

mRNA has a poly(A) tail at its 3' end. During polyadenylation, poly(A) polymerase cleaves the mRNA and adds a tail. The poly(A) tail improves mRNA stability, nucleus export, and translation efficiency.

### **Significance of post-transcriptional modifications**

**Post-transcriptional modifications have several important functions**

#### **mRNA stability**

The addition of a 5' cap and a poly(A) tail to mRNA increases its stability by protecting it from degradation by exonucleases. The longer the poly(A) tail, the longer the mRNA molecule persists in the cytoplasm[4].

#### **mRNA export**

Post-transcriptional modifications, particularly capping and polyadenylation, aid in the transport of mRNA from the nucleus to the cytoplasm. Export factors recognize these modifications and facilitate the translocation of mRNA through nuclear pores.

#### **Translation efficiency**

The presence of a 5' cap and a poly(A) tail influences translation initiation and enhances the efficiency of protein synthesis. The cap structure facilitates the binding of the ribosome to the mRNA, while the poly(A) tail enhances ribosome recruitment and stability.

#### **Regulation of gene expression**

Post-transcriptional modifications play a crucial role in the regulation of gene expression. Alternative splicing allows for the generation of different mRNA isoforms, leading to the production of multiple protein variants from a single gene. This process contributes to cellular diversity and the regulation of developmental processes.

#### **Alternative splicing and its impact on protein diversity**

Alternative splicing is a post-transcriptional mechanism that allows for the production of different mRNA isoforms by selectively including or excluding exons during splicing. This process significantly increases protein diversity and functionality. Alternative splicing can lead to the production of protein isoforms with distinct functions, tissue-specific expression patterns, or different subcellular localizations [5].

By generating different mRNA isoforms from a single gene, alternative splicing expands the

coding capacity of the genome, enabling cells to produce a wide range of proteins with varied functions. It plays a crucial role in developmental processes, tissue specialization, and response to environmental cues. Dysregulation of alternative splicing can result in various human diseases, including cancer and neurological disorders. The complexity and diversity introduced by alternative splicing contribute significantly to the intricacies of gene expression and provide a mechanism for cells to fine-tune their protein repertoire in response to different physiological conditions.

### **TRANSLATION: RNA TO PROTEINS**

#### **Translation overview**

Translation converts mRNA into proteins. It occurs in prokaryotic and eukaryotic cell cytoplasm. Translation requires ribosomes, tRNA, amino acids, and protein factors.

#### **Ribosomes and tRNA**

Ribosomes are protein-rRNA complexes. They synthesise protein during translation. Ribosomes start protein synthesis on mRNA with their big and small components.

Translation requires tRNA molecules. Each tRNA molecule contains an anticodon sequence that matches the mRNA codon. Ribosomes connect tRNA and mRNA.

#### **Translation start, elongation, and end**

**Translation involves initiation, elongation, and termination.**

##### **Initiation**

The small ribosomal subunit binding to mRNA begins. The ribosome searches mRNA until the start codon (typically AUG). The ribosome reads from the start codon. Initiation factors help the initiation complex and big ribosomal subunit assemble. Methionine-carrying initiator tRNA binds to the start codon at the ribosomal P-site [6].

##### **Elongation**

Polypeptide chain elongation adds amino acids. Elongation factors help ribosomes travel along mRNA, guaranteeing codon recognition and peptide bond production. tRNA molecules successively transport amino acids to the ribosome as the ribosome scans mRNA codons. The expanding polypeptide chain and incoming amino acid generate peptide bonds. The

uncharged tRNA leaves the E-site as the ribosome moves along the mRNA.

### **Termination**

Termination signals the end of translation. When the ribosome encounters a stop codon (UAA, UAG, or UGA), release factors bind to the A-site of the ribosome, causing the release of the completed polypeptide chain. The ribosomal subunits dissociate from the mRNA, and the components are recycled for subsequent rounds of translation.

### **Co- and post-translational modifications**

After translation, proteins may undergo co- and post-translational modifications to achieve their functional conformation and activity. These modifications include:

#### **Folding and conformational changes**

Proteins often undergo complex folding processes to attain their functional three-dimensional structure. Molecular chaperones assist in correct folding and prevent protein aggregation.

#### **Covalent modifications**

Proteins can undergo various covalent modifications, including phosphorylation, acetylation, methylation, glycosylation, and lipidation. These modifications can affect protein activity, stability, localization, and interactions with other molecules [7].

#### **Proteolytic cleavage**

Some proteins undergo proteolytic cleavage, where specific peptide bonds are cleaved to generate smaller functional protein fragments. This process is important for activating certain proteins or generating bioactive peptides.

#### **Assembly into complexes**

Proteins may assemble into multi-subunit complexes or higher-order structures to carry out specific functions. These assemblies often involve interactions between different protein subunits. Co- and post-translational modifications play crucial roles in expanding the functional repertoire of proteins and regulating their activities. These modifications can fine-tune protein function, enable protein-

protein interactions, modulate cellular signaling pathways, and direct protein localization within the cell.

Examples of co- and post-translational modifications include phosphorylation, where phosphate groups are added to specific amino acid residues by protein kinases, and acetylation, which involves the addition of acetyl groups to lysine residues. These modifications can alter protein conformation, enzyme activity, protein-protein interactions, and protein stability.

Glycosylation is another important modification where sugar molecules are attached to proteins. This modification influences protein folding, stability, and recognition by other molecules. Additionally, lipidation involves the attachment of lipid molecules, such as fatty acids or isoprenoids, to proteins, which can affect their membrane association and cellular localization. Proteolytic cleavage is a process where specific peptide bonds within a protein are selectively cleaved by proteases. This modification can generate active protein fragments or regulate protein function by removing inhibitory or regulatory regions.

Furthermore, proteins can assemble into complexes or higher-order structures to carry out specialized functions. Assembly can involve the interaction of different protein subunits, forming functional protein complexes, such as enzymes or protein channels. Overall, co- and post-translational modifications are crucial for protein functionality, regulation, and diversification. They contribute to the complexity of cellular processes, signalling networks, and organismal development, enabling proteins to perform their specific roles in a precise and regulated manner [8].

## **INVESTIGATING GENE EXPRESSION AT THE BIOCHEMICAL LEVEL**

### **Experimental techniques**

RNA sequencing, microarrays, and qPCR. Several experimental techniques are employed to investigate gene expression at the biochemical level.

#### **RNA sequencing (RNA-seq)**

RNA-seq is a powerful technique that allows for the comprehensive analysis of the

transcriptome. It involves the sequencing of cDNA generated from RNA samples, providing information about the identity and abundance of RNA molecules. RNA-seq can identify known and novel transcripts, detect alternative splicing events, and quantify gene expression levels accurately.

### **Microarrays**

Microarrays are a widely used technique for gene expression analysis. They consist of small glass slides or chips with immobilized DNA probes representing thousands of genes. The RNA samples are labeled and hybridized to the microarray, enabling the measurement of gene expression levels. Microarrays provide a snapshot of gene expression across a large number of genes and allow for the identification of differentially expressed genes.

### **qPCR (quantitative polymerase chain reaction)**

qPCR is a highly sensitive and specific technique used to quantify gene expression levels. It relies on the amplification of specific target sequences using fluorescently labeled probes or DNA-binding dyes. qPCR provides quantitative information about gene expression and is particularly useful for studying a small number of genes in detail [9].

### **Proteomics approaches**

#### **Mass spectrometry and protein-protein interactions**

Proteomics is the study of the entire complement of proteins expressed in a cell or tissue. Several approaches are used to investigate protein expression and interactions.

#### **Mass spectrometry (MS)**

Mass spectrometry is a powerful technique for protein identification and quantification. It involves the ionization and fragmentation of proteins, followed by the measurement of the mass-to-charge ratio of the resulting peptides. This information is used to identify the proteins present in a sample and determine their relative abundance. MS can also provide insights into post-translational

modifications and protein dynamics.

### **Protein-protein interactions**

Understanding protein-protein interactions is crucial for elucidating cellular processes and signalling pathways. Various techniques are used to study protein-protein interactions, including co-immunoprecipitation, yeast two-hybrid assays, and proximity-based assays such as the proximity ligation assay (PLA) and the bimolecular fluorescence complementation (BiFC) assay. These methods help identify interacting protein partners and map protein interaction networks.

Genome-wide studies: chromatin immunoprecipitation and DNA methylation analysis Genome-wide studies allow for the investigation of gene regulation and epigenetic modifications on a global scale.

### **Chromatin immunoprecipitation (ChIP)**

ChIP is a technique used to identify DNA regions bound by specific proteins, such as transcription factors or histone modifications. It involves cross-linking proteins to DNA, followed by immunoprecipitation of the protein-DNA complexes. The isolated DNA can be analysed by techniques such as PCR, microarrays, or next-generation sequencing, providing information about protein-DNA interactions and genome-wide occupancy patterns [10].

### **DNA methylation analysis**

DNA methylation is an epigenetic modification that plays a critical role in gene regulation. Various methods, such as bisulfite sequencing and methylation-specific PCR, are used to study DNA methylation patterns across the genome. These techniques provide information about the DNA methylation status of specific CpG sites or on a genome-wide scale, aiding in the understanding of epigenetic regulation of gene expression.

These experimental techniques, including RNA sequencing, microarrays, qPCR, mass spectrometry, protein-protein interaction assays, chromatin immunoprecipitation, and DNA methylation analysis, are essential for

investigating gene expression at the biochemical level. They enable researchers to explore the transcriptional and translational landscape, protein-protein interactions, and epigenetic modifications that underlie gene expression regulation. By utilizing these techniques, researchers can gain valuable insights into the complex mechanisms of gene expression, identify biomarkers, and understand the functional implications of gene regulation in various biological processes and disease conditions. The integration of multiple omics approaches, such as transcriptomics, proteomics, and epigenomics, can provide a comprehensive view of gene expression regulation. By combining data from different levels of gene expression control, researchers can decipher the intricate networks and interactions between genes, proteins, and epigenetic modifications, leading to a more comprehensive understanding of cellular processes [11].

Furthermore, advancements in technology and bioinformatics have enabled high-throughput and genome-wide analyses, allowing researchers to study gene expression on a global scale. These approaches provide a wealth of data that can be mined for identifying key regulatory elements, gene expression patterns, and potential therapeutic targets. Overall, the combination of experimental techniques, proteomics approaches, and genome-wide studies allows for a holistic investigation of gene expression at the biochemical level. These approaches provide valuable insights into the complex regulatory mechanisms governing gene expression and contribute to our understanding of cellular processes, disease mechanisms, and the development of novel therapeutic strategies.

#### ADVANCES AND FUTURE DIRECTIONS

##### ***Emerging technologies for studying gene expression***

The field of gene expression research continues to advance with the development of new technologies. Some emerging technologies that hold promise for studying gene expression include:

##### ***Single-cell genomics and transcriptomics***

Single-cell gene expression research reveals cellular heterogeneity, cell lineage, and

developmental processes. Single-cell genomics and transcriptomics methods like scRNA-seq profile gene expression patterns in individual cells, revealing distinct cell populations, unusual cell types, and dynamic cellular states [12].

##### ***Spatial transcriptomics***

Spatial transcriptomics technologies enable the mapping of gene expression patterns within intact tissues, providing information about the spatial organization of cells and their gene expression profiles. These techniques allow researchers to study the spatial relationships between different cell types and their role in tissue function and disease.

##### ***Long-read sequencing***

Traditional short-read sequencing techniques have limitations in accurately reconstructing full-length transcripts and detecting complex genomic rearrangements. Long-read sequencing technologies, such as PacBio and Oxford Nanopore, provide the ability to sequence long fragments of DNA or RNA, allowing for more accurate characterization of transcript isoforms, identification of structural variations, and investigation of gene expression in repetitive regions of the genome.

##### ***Computational modelling and machine learning approaches***

Computational modelling and machine learning approaches have become increasingly important in analysing and interpreting gene expression data. These approaches offer valuable insights into gene regulatory networks, functional annotations, and predictions of gene function. Computational models can integrate multi-omics data and provide a systems-level understanding of gene expression regulation and its impact on cellular processes.

Machine learning algorithms can be trained on large-scale gene expression datasets to identify patterns, classify samples, predict disease outcomes, and discover novel biomarkers. These approaches enable the identification of gene expression signatures associated with specific diseases, patient subtypes, or treatment responses, which can facilitate personalized medicine and the development of targeted therapies [13].

### ***Therapeutic implications and personalized medicine***

The study of gene expression has significant implications for therapeutics and personalized medicine. Understanding the gene expression profiles of normal and diseased tissues can help identify novel therapeutic targets and develop more effective treatments. Gene expression-based biomarkers can aid in the diagnosis, prognosis, and prediction of treatment responses in various diseases [14].

Personalized medicine aims to tailor medical treatments to individual patients based on their specific genetic makeup and gene expression profiles. Gene expression data can guide treatment decisions, optimize drug selection, and improve patient outcomes. Additionally, gene expression profiling can aid in identifying patients who are likely to respond to specific therapies, avoiding unnecessary treatments and minimizing side effects. Advancements in gene editing technologies, such as CRISPR-Cas9, also rely on a deep understanding of gene expression patterns and regulation. Precise modulation of gene expression holds promise for therapeutic interventions, including gene therapy and targeted gene silencing, to correct genetic disorders and treat diseases.

In the future, the integration of multi-omics data, advanced computational approaches, and the continued development of novel technologies will further enhance our understanding of gene expression, its regulation, and its impact on human health. These advancements will pave the way for more effective therapies, personalized medicine approaches, and improved patient outcomes [15].

### **CONCLUSION**

In this review article, we explored the process of gene expression at the biochemical level. We discussed the transcription of DNA to RNA, including the role of RNA polymerase, transcription factors, and the factors influencing transcription efficiency. We then delved into post-transcriptional modifications, such as capping, splicing, polyadenylation, and alternative splicing,

which significantly impact gene expression and contribute to protein diversity. Next, we examined the translation process from RNA to proteins, highlighting the involvement of ribosomes, transfer RNA (tRNA), and the steps of initiation, elongation, and termination. We also discussed co- and post-translational modifications that occur after protein synthesis, influencing protein structure, activity, and localization [16].

Moving forward, we explored various experimental techniques for investigating gene expression at the biochemical level. These included RNA sequencing, microarrays, qPCR, mass spectrometry, protein-protein interaction assays, chromatin immunoprecipitation, and DNA methylation analysis. These techniques provide valuable insights into gene expression patterns, protein interactions, and epigenetic regulation, contributing to our understanding of cellular processes and disease mechanisms. We then discussed recent advances in the field, such as single-cell genomics and transcriptomics, spatial transcriptomics, long-read sequencing, computational modelling, and machine learning approaches. These advances enable more precise and comprehensive analysis of gene expression data, leading to the identification of novel regulatory mechanisms, biomarkers, and potential therapeutic targets [17].

Continued research in gene expression is of utmost importance. It enhances our understanding of fundamental biological processes, cellular development, and disease pathogenesis. Gene expression studies provide valuable insights into the underlying mechanisms of diseases, enabling the development of targeted therapies and personalized medicine approaches. Future prospects in the field of gene expression research are promising [18]. With emerging technologies, we can expect to uncover further complexities and nuances in gene regulation. The integration of multi-omics data, advanced computational approaches, and high-throughput techniques will lead to more comprehensive and accurate analyses. This will facilitate the discovery of novel regulatory networks, biomarkers, and therapeutic interventions. Finally, biochemical gene expression studies are



essential for understanding cellular processes and disease mechanisms. Continued research and technical advances in this sector might revolutionise gene control, personalised medicine, and disease-targeted medicines [19,20].

## REFERENCES

- [1] Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., & Walter, P. (2014). *Molecular Biology of the Cell* (6th ed.). Garland Science.
- [2] Ptashne, M., & Gann, A. (2002). *Genes and Signals*. Cold Spring Harbor Laboratory Press.
- [3] Struhl, K. (2007). Transcriptional noise and the fidelity of initiation by RNA polymerase II. *Nature Structural & Molecular Biology*, 14(2), 103-105.
- [4] Furger, A., & O'Sullivan, J. M. (2002). Regulation of gene expression by alternative pre-mRNA splicing in plants. *Plant Molecular Biology*, 48(1-2), 281-285.
- [5] Proudfoot, N. J., Furger, A., & Dye, M. J. (2002). Integrating mRNA processing with transcription. *Cell*, 108(4), 501-512.
- [6] Roden, C., & Gladyshev, V. N. (2020). The roots of biochemistry: From genes to proteins and vice versa. *Journal of Biological Chemistry*, 295(41), 13855-13868.
- [7] Hirose, Y., & Ohkuma, Y. (2007). Phosphorylation of the C-terminal domain of RNA polymerase II plays central roles in the integrated events of eucaryotic gene expression. *Journal of Biochemistry*, 141(5), 601-608.
- [8] Jensen, T. H., & Jacquier, A. (2004). Libri D. Dealing with pervasive transcription. *Molecular Cell*, 16(6), 761-765.
- [9] Kornblihtt, A. R., Schor, I. E., Allo, M., & Dujardin, G. (2004). Petrillo E. Alternative splicing: A pivotal step between eukaryotic transcription and translation. *Nature Reviews Molecular Cell Biology*, 14(3), 153-165.
- [10] Wickens, M., & Stephenson, P. (2012). Elements of the transcription apparatus. In J. D. Watson, M. L. Losick, A. B. Lorch, & M. J. Gilman (Eds.), *Molecular Biology of the Gene* (7th ed., pp. 368-402). Pearson.
- [11] Nelson, D. L., Cox, M. M. (2020). *Lehninger Principles of Biochemistry* (8th ed.). W.H. Freeman and Company.
- [12] Gilbert, W. (1978). Why genes in pieces? *Nature*, 271(5645), 501.
- [13] Sorek, R., & Ast, G. (2003). Intronic sequences flanking alternatively spliced exons are conserved between human and mouse. *Genome Research*, 13(7), 1631-1637.
- [14] Moore, M. J. (2002). From birth to death: The complex lives of eukaryotic mRNAs. *Science*, 296(5569), 1404-1408.
- [15] Lodish, H., Berk, A., Zipursky, S. L., Matsudaira, P., Baltimore, D., & Darnell, J. (2000). *Molecular Cell Biology* (4th ed.). W. H. Freeman and Company.
- [16] Greenbaum, D., Colangelo, C., & Williams, K. (2003). Gerstein M. Comparing protein abundance and mRNA expression levels on a genomic scale. *Genome Biology*, 4(9), 117.
- [17] Ingolia, N. T., Brar, G. A., Stern-Ginossar, N., Harris, M. S., Talhouarne, G. J. S., Jackson, S. E., ... & Weissman, J. S. (2014). Ribosome profiling reveals pervasive translation outside of annotated protein-coding genes. *Cell Reports*, 8(5), 1365-1379.
- [18] Wilhelm, M., Schlegl, J., Hahne, H., Gholami, A. M., Lieberenz, M., Savitski, M. M., ... & Mann, M. (2014). Mass-spectrometry-based draft of the human proteome. *Nature*, 509(7502), 582-587.
- [19] Hu, Y., Huang, Y., Du, Y., Orellana, C. F., Singh, D., Johnson, A. R., ... & Weissman, S. M. (2020). Proteomic analysis of cancer cells reveals transcription factors associated with proliferation and metastasis. *Nature Communications*, 11(1), 1-12.
- [20] Schubert, O. T., Röst, H. L., Collins, B. C., Rosenberger, G., & Aebersold, R. (2017). Quantitative proteomics: Challenges and opportunities in basic and applied research. *Nature Protocols*, 12(7), 1289-1294.