

Prevalence and Antifungal Susceptibility of Candida species at a Tertiary Care Hospital in North India

SaherKhan¹, Mastan Singh², Vineeta Khare³, M. M. Abid Ali Khan⁴, Prashant Gupta⁵ S. Tasleem Raza⁶

^{1,2,3} Department of Microbiology, Era's Lucknow Medical College & Hospital, Lucknow, U.P.

India

⁴Department of Botany, Shia P.G, College, Lucknow, U.P. India

⁵Department of Microbiology, King George's Medical University, Lucknow, U.P., India

⁶Department of Biochemistry, Era's Lucknow Medical College & Hospital, Lucknow, U.P. India

Article History:	Received: 22.04.2023	Revised: 12.05.2023	Publication: 30/05/2023
	DOI: 10.4804	7/ecb/2023.12.5.501	

Abstract:

Background: The introduction of Candida species other than *Candida albicans* as the primary agents has caused a recent shift in the epidemiology of candidiasis, which is extremely concerning on a global scale. Some species of Candida are naturally resistant to azoles. For directing therapy, in vitro susceptibility testing is crucial.

Aim: This study was conducted to determine the prevalence of Candida spp. and evaluate its susceptibility profile against antifungal agents at a tertiary care center in North India.

Methods: A total of 140 confirmed cases of Candida infection were enrolled. Demographic and epidemiological details like gender and specimen type were noted. Species identification was done by phenotypic methods and Antifungal susceptibility testing was done by E test.

Results: Majority of patients were females (n=87; 62.1%). Urine (n=65; 46.4%), was the most common specimen followed by Sputum (n=30; 21.4%) %), Blood (n=12; 8.5%) Body fluids (n=10; 7.14%) oral swab (n=10; 7.14%), High vaginal swab (n=8; 5.7%) and Nails (n=5; 3.5%) respectively. *C. tropicalis* (n=48, 34.3%) was the most common species followed by *C.albicans* (n=35; 25%) *C. glabrata* (n=21; 15%), *C krusei*. (n=14; 10%), *C. parapsilosis* (n=12; 8.5%), *C. guilliermondii* (n=6; 4.2%) and *C. rugosa* (n=4; 2.8%) respectively. Regarding antifungal sensitivity Caspofungin and Amphotericin B were found to be most sensitive (100%) against all the species. Fluconazole was the most resistant antifungal agent followed by Itraconazole. Urine was the major source for Candida infection in women suggesting their high susceptibility to candida infection. **Conclusion**: Our study highlights the predominance of NAC species. Caspofungin and Amphotericin B were the most sensitive antifungal agents against all candida species whereas, Fluconazole was least effective. Species Identification and antifungal susceptibility should be done to help the treatment process of Candidal infection.

Keywords: Candida, Antifungal susceptibility, E-test, Caspofungin, Voriconazole.

Introduction:

Evidence suggests that more than one billion people are affected by fungal infections annually, and nearly 1.5 million are expired due to infection severity (1). This suggests that fungal infections have a high mortality rate. Among various fungal species, Candida is the most common type. It is an endogenous species and its spread is marked as an opportunistic infection (2). Candida spp. is normally distributed in the flora of skin, mucous membrane and gastrointestinal tracts of healthy individuals. However, with the increasing trends of immunocompromised persons being at high risk, there is an upward trajectory in the rate of fungal infection, specifically, due to Candida (3). Although, there are over 200 species of Candida. *C.albicans* is the most common. However, a number of other species including *C.glabrata*, *C.parapsilosis*, *C.tropicalis* and *C. krusei*

are also commonly isolated from hospital specimen. Out of 200 species, nearly 20 species cause serious infection in humans(4). When these fungi grow, the result in Candiasis/Candidosis/Candidemia causing superficial, subcutaneous and deep mycosis(5). Candidiasis is a life threatening with a mortality rate as high as 38%. It also responsible for prolonging duration of hospital stay (6). In order to offer effective treatment, it is essential that fungal species affecting the patient should be identified correctly because; sensitivity profile of a fungal species against various antifungal agents varies. Moreover, in view of the emergence of antifungal resistant species, it is essential that the antifungal susceptibility profile of different *candida* species should be studied. With this objective, the present study was carried out to describe distribution of various *Candida* spp. among clinical isolate and antifungal susceptibility profile of various *candida* species at a tertiary care centre in north India.

Materials and Methods

The approval for the study was obtained from the Institutional Ethics Committee and permission to analyze the data was obtained from respective authorities. As the study did not directly involve inclusion of patients and/or their clinical profile, hence there was no need to obtain informed consent from the patients.

Collection of Specimens

A total of 140 isolates of *Candida* obtained from different clinical specimen such as Blood, Urine, Sputum, Body fluids, High vaginal swab, Oral swab and Nails were collected from patients attending various departments of Era's Lucknow Medical College Lucknow. Samples were collected in sterile collection devices and containers with appropriate label. Institutional Ethics Committee of Era's Lucknow Medical College and Hospital (ELMC & H) approved the study. Since this study did not involve direct inclusion of patients and/or their clinical profile, Informed consent from the patients was not required.

Processing of Specimens

Primary identification was performed by direct smear examination by Gram's staining and KOH mount. Samples were collected using aseptic precautions and inoculated on Sabouraud's Dextrose Agar SDA (Hi media) screw caped bottles and incubated at 37°C for48-72hours.After growth, species identification was done by Germ tube test,HiCromeAgar media(Himedia),carbohydrate fermentation test and sugar assimilation test.

Antifungal susceptibility testing done by E-TEST-The E-test gradient strips of Amphotericin

B,Fluconazole,Voriconazole,Itraconazole and Caspofungin was obtained from HIMEDIA. The concentration gradient for Fluconazole ranged from (0.016 -256 µg/mL) while for other drugs was0.002-32µg/mL,The E-testwasperformedbyfollowingthemanufacturer's instructions. E strips were applied and the plates were incubated at 35°C and read after 48hours. The MIC was determined from the inhibition ellipse that intersected the scale on the strip (7) C. parapsilosis ATCC 22019 and C. krusei ATCC 6258 were included as quality controls.The interpretative criteria for Fluconazole(FCZ),Itraconazole(ICZ),and Voriconazole (VCZ) susceptibility of Candida species in vitro were derived from the CLSI M27-A3 and M60-ED 1 documents(8,9) while the interpretative criteria for Amphotericin B (AMB) were taken from the literature (10,11). For each group, the lowest concentrations at which 50% (MIC 50) and 90% (MIC 90) of the isolates were inhibited were determined.

DataAnalysis

Data was tabulated and presented as frequency and proportions. The antifungal efficacy was evaluated in terms of Sensitive, Susceptible Dose dependent and Resistant outcomes. MIC 50 and MIC 90 are also calculated. **Results**

In this study, we analyzed 140 specimens including 62.1% (n=87) females and 37.9% (n=53) males. The most common specimen was Urine (n=65; 46.4%) followed by Sputum (n=30; 21.4%), Blood (n=12; 8.5%) Body fluids (n=10; 7.14%) Oral swab (n=10; 7.14%), High vaginal swab(n=8;5.7%)and Nail(n=5; 3.5%)respectively (Table 2).

C.tropicalis (n=48, 34.3%) was the most common species followed by C.albicans (n=35;25%)

5938

C.glabrata(n=21;15%), C.krusei (n=14;10%), C.parapsilosis(n=12;8.5%), C.guilliermondii (n=7;5%) and C. rugosa (n=4; 2.8%) respectively (Table 3).

Table 4 summarises the MIC ranges and MIC50 and MIC90 values obtained in susceptibility testing using the CLSI method. According to the MIC values obtained for Amphotericin B,and Caspofungin against all the isolates of candida species were classified as susceptible according to the described breakpoints. The MIC values for Itraconazole were 0.006to1µg/mL, indicating that 103 of the candida isolates were susceptible (73.5%), 29 were dose-dependently susceptible (20.7%), and 8 isolates were resistant (5.7%). Regarding Fluconazole79 of the candida isolates were susceptible (56.42%), 7 were intermediate (5%) and 56 were resistant (40%) Voriconazole showed sensitivity against 127 isolates of Candida (90.7%) and resistant against 13 isolates of Candida (9.2%) respectively. The highest MIC was showed by Fluconazole while lowest Caspofungin and Amphotericin B. Caspofungin and Amphotericin B were found to be most sensitive antifungal agent against all the Candida species (100%). Overall, Fluconazole was the most resistant antifungal agent followed by Itraconazole.

S.No	Gender	Number	Percentage
1	Female	87	62.1
2	Male	53	37.9
3	Total	140	100

1 1. 4. 11 4. CD /

abitation of concercu numbers of samples							
S.No	Clinical Specimens	Number	Percentage (%)				
1	Urine	65	46.4				
2	Sputum	30	21.4				
3	Blood	12	8.5				
4	Fluids	10	7.14				
5	Oral Swab	10	7.14				
6	High Vaginal Swab	8	5.7				
7	Nail	5	3.5				

Table? Distribution of collected numbers of samples

Table3.S	necies	of	Candida	Isolated	from	clinical	samples
1 anico.o	JULIUS	UI	Cununuu	isolattu	nom	unnuar	sampics

S.No.	Candida species	Number	Percentage (%)
1	C.tropicalis	48	34.3%
2	C.albicans	35	25%
3	C. glabrata	21	15%
4	C.krusei	14	10%
5	C.parapsilosis	12	8.5%
6	C.guilliermondii	6	4.2%
7	C.rugosa	4	2.8%



Fig1. Candida species growth on HiCromeMedia



Fig2.Caspofungin showing sensitivity against candida albicans

Table4.Susceptibility patterns of *Candida* species isolated from patients

Candida species(No.of strain)	Antifungal agent	Range	MI C5 0	MIC9 0	Susceptibili ty break points	SDD/I	R
C.albicans (35)	Amphotericin B	0.004–.25	.032	.19	≤1	-	≥1
	Fluconazole	0.125-12	0.25	1	≤2	4	≥ 8
	Itraconazole	0.125-1	.25	1	≤0.125	0.25-	≥1
						0.5	
	Voriconazole	0.006-1	.008	.032	≤0.12	0.25-	≥1
						0.5	
	Caspofungin	0.002016	0.006.	.008	≤0.25	0.5	≥1

C.tropicalis (48)	AmphotericinB	0.002-0.38	.064	0.94	≤1	-	≥1
	Fluconazole	0.25-12	0.5	0.75	≤2	4	≥ 8
	Itraconazole	0.012-0.5	0.125	0.5	≤0.125	0.25-	≥1
						0.5	
	Voriconazole	<0.12-2	.012	1	≤0.12	0.25-	≥1
						0.5	
	Caspofungin	.004-0.23	.004	.012	≤0.25	0.5	≥1
C.glabrata (21)	AmphotericinB	0.01225	0.125	0.75	≤1	-	≥1
	Fluconazole	0.023–64	6	8	-	≤32	≥64
	Itraconazole	0.125–0.5	0.125	0.5	≤0.125	0.25-	≥1
						0.5	
	Voriconazole	0.006-2	0.125	0.38	<u>≤0.5</u>	-	<u>≥1</u>
	Caspofungin	.002012	.004	.012	≤0.12	0.25	<u>≥0.5</u>
C.krusei (14)	AmphotericinB	0.06475	.25	.38	<u>≤1</u>		<u>≥1</u>
	Itraconazole	0.094-1	.25	0.5	≤0.125	0.25-	≥1
		0.064.4	0.64	0.105	.0. 7	0.5	
	Voriconazole	0.064-4	.064	0.125	<u>≤0.5</u>	1	<u>≥2</u>
	Caspotungin	0.006016	0.006	0.008	≤0.25	0.5	≥I
C	A	0.002	022	004	<1	-	> 1
C.parapsilosis	AmphotericinB	0.002-	.032	.094	≤ 1	-	≥1
(12)		0.125					
	Fluconazole	0.032–8	0.25	0.38	≤2	4	≥ 8
	Itraconazole	0.006-0.5	.094	.25	≤0.125	0.25-0.5	≥ 1
	Voriconazole	0.004–	0.006	0.023	≤0.12	0.25-0.5	≥1
		0.032					
	Caspofungin	0.004032	0.004	.016	≤2	4	≥8
c.rugosa(4)	AmphotericinB	0.19–0.5	0.25	0.5	≤1	-	≥1
	Itraconazole	0.125–.25	.125	.25	≤0.125	0.25-0.5	≥1
	Voriconazole	.012094	.012	.064	≤0.12	0.25-0.5	≥1
	Fluconazole	8-16	8	8	≤2	4	≥8
	Caspofungin	0.002016	0.006	.012	<2	4	>8
	ousporting	0.002.010	0.000				_0
C avillianm an dii	AmphotoriainD	0.002	012	022	<1		>1
C.guillermonall	Amphotericing	0.002 - 0.125	.012	.032	<u>></u> 1	-	≥1
(0)		0.125		<u> </u>			
	T. 1	0.04-0.5	.012	0.5	≤0.125	0.25-0.5	≥ 1
	Itraconazole						
	Voriconazole	.032094	.032	.064	≤0.12	0.25-0.5	≥1
	Fluconazole	8-16	8	8	≤2	4	≥ 8
	Caspofungin	0.004032	.006	.012	<2	4	>8
	1				_		

*Fluconazole,Itraconazole and Voriconazole range values according to Clinical and Laboratory Standards Institute(CLSI) literature

*Ranges:FCZ(0.016-256µg/mL),ICZ(0.002-32µg/mL),VCZ(0.002-32µg/mL),andAMB (0.002-32µg/mL)andCAS(0.002-32µg/mL)

Caudidaanaaiaa	Antifungol		Decodemondente	Desistant
Canalaaspecies	Antifungal	Susceptible	Susceptible Dosedependents	
	agents		lintormodiato	
C all i_{2} and (25)	•	25(1000/)	/intermediate	
C.albicans(33)	A	35(100%)	-	-
	F	27(77.14%)	-	8(22.85)
		29(82.85%)	4(11.42%)	2(5.71%)
	V	32(100%)	-	3
	С	35(100%)	-	-
C.tropicalis(48)	A	48(100%)	-	-
	F	38(87.5%)	-	10(12.5%)
	Ι	44(91.6%)	4(8.33%)	-
	V	44(91.6%)	-	4(8.33%)
	С	48(100%)	-	-
		· · ·		
C. glabrata(21)	А	21(85.71%)	-	-
6	F	4(19.04%)	7(52.38%)	10(47.61%)
	Ι	15(85.71%)	6(14.28%)	
	V	19(90.47%)	-	2(9.5%)
	С	21(100%)	-	-
C krusei(14)	А	14(100%)		
	F	-	-	14(100%)
	I	3(71.42%)	5(35.7%)	6(28 57%)
	V	10(71.42%)	-	4(28.5%)
	C	10(71.4270) 14(100%)	_	-
C		14(100%)		_
C.	Λ	12(10070)	-	-
purupsilosis		10(02.20/)		2(1((0/)
(10)	F	10(83.3%)	-	2(16.6%)
(12)				
	I	7(58.3%)	5(41.6%)	-
	V	12(100%)	-	-
	С	12(100%)	-	-
C.rugosa(4)	А	4(100%)	-	-
	F	0(0)	-	4(100%)
	Ι	2(100%)	2(50%)	-
	V	4(100%)	-	-
	С	4(100%)	-	-
С.	А	6(100%)	-	-
guilliermondii				
	F	1(16.6%)	-	5(83.3%)
(6)				
	T	3(100%)	3(50%)	
	V	6(100%)	-	_
	V C	0(100%) 6(100%)	-	-
L		0(100%)		-

 Table5.Susceptibilitypattern of Candidaspecies based on MIC values

Discussion

In the present study, gender-specific differences in the prevalence of Candida infection were observed. In all specimen collected, percentage of women (62.1%) was higher than man(37.9%) suggesting that women have high risk for developing candida infection. In an earlier study, Guru and Raveendra also obtained majority of their candida specimen from women(54.5%)(2). Similarly, Pawar et al. and Prabhakaran et al. also reported higher proportion offemales compared to males in various specimen they studied (4,12). The reason for this could be high prevalence of Candida in vulvovaginal infections. Vulva and vagina provide the most conducive environment for fungal growth particularly Candida. More over, the affected women often complain of recurrent

Candidiasis suggesting their susceptibility of infection(13). The high prevalence of urine specimen(46.4%) also support the dominance of vulvovaginal source of infection. In the present study, sputum (21.4%) was the second most common specimen found positive for Candida, which suggests that oral Candidiasis is also common. Guru and Raveendran also reported urine specimen as the most common source for Candida (53%), with high prevalence in women (54.5%) (2). In recent years, oral candidiasis has been recognized as one of the most common mycoses in human beings (14). The present findings also suggest that oral candidiasis is amongst the most common mycoses in humans.

A number of previous studies have reported *C.albicans* as the most common Candida species. However, in the recent years several studies have reported a high prevalence of other species too. In present study, *C. tropicalis* (n=48,34.3%) was the most common Candida species followed by *C.albicans* (n=35,25%) followed by *C.glabrata* (n=21,15%), *C.krusei* (n=14;10%), *C.parapsilosis* (n=12; 8.5%), *C.guilliermondii* (n=6;4.2%) and *C.rugosa* (n=4;2.8%).Our findings are in agreement with the observations made by Chaudhary et al. who found *C.tropicalis* as the most prevalent (43%) species followed by *C. albicans* (41%), *C. krusei*(9%) and *C.parapsilosis* (7%) (15). In another study, Mathur et al. also did not find *C.albicans* to be the most common isolate. They observed that *C.tropicalis* (39.4%) was the most common species, followed by *C.auris* (17.5%), *C.albicans*(14%) and *C.parapsilosis*(11.4%). These findings show that there is shifting the trend of Candida species prevalence (16).

Regarding the susceptibility of various Candida species to antifungal drugs, we found that all species of candida were highly susceptible (100%) to new generation antifungal drugs like Caspofungin and Amphotericin B whereas, however, maximum resistance was observed for Fluconazole(40%) followed by Itraconazole (5.7%) which also had a dose-dependent susceptibility to 29(20.7%) specimen. In their study,Guru and Raveendran also found Amphotericin B to be most sensitive (98%) however, they did not evaluate the antifungal susceptibility pattern against the newer generation antifungals like Caspofungin(2). However,Gandhi and Patel in their study reported the sensitivity of Amphotericin B to be maximum (97%) followed by Voriconazole(77%), Fluconazole(75%) and Itraconazole(64%) respectively(17).

In this study a hundred percent resistance was reported for *C. krusei* against Fluconazole because of its intrinsic resistance toward azoles. Also, the resistance of *C.glabrata* to Fluconazole was consistently higher (47.61%) resistant was observed however in their study, GeetaS.H. found *C. glabrata* was100%sensitivity against Fluconazole (18).

Conclusion

The findings of present study thus showed changing trends of *Candida* species and their antifungal susceptibility pattern at a tertiary care centre in North India. Keeping in view of the changing profile of *Candida* species and their antifungal susceptibility, there should be continuous monitoring of the prevalence of various fungal species in order to design effective treatment strategies Antifungal MIC determination is also essential in this period, as *Candida* species with higher MICs values are on the rise.

Conflict of Interests Statement: The authors declare that there is no conflict of interests regarding the publication of this article.

Funding-None

Ethical-Ethical committee of ERA's Medical College, approved the study protocol

Authors' contribution: SK, drafted the manuscript, gathered information from the literature, performed the experiments , designed the figures & tables and wrote the paper. MS, PK, MA and VK supervised and reviewed the manuscript.

Acknowledgements

Facilities provided by the Department of Microbiology, Era's Lucknow Medical College are gratefully acknowledged.

References

- 1. Bongomin F, Gago S, Oladele RO, Denning DW. Global and multi-national prevalence offungaldiseases—estimateprecision. Journal offungi. 2017 Oct 18;3(4):57.
- Guru P, Raveendran G. Characterisation and antifungal susceptibility profile of Candidaspeciesisolatedfromatertiarycarehospital.JournalofTheAcademyofClinicalMicrobiologists.2016 Jan 1;18(1):32.
- 3. Rizvi MW, Malik A, Shahid M, Singhal S. Candida albicans infections in a north Indiantertiarycarehospital:antifungalresistancepatternandroleofSDS-PAGEforcharacterization.Biol Med. 2011;3:176-81..
- 4. Pawar M,Misra RN, Gandham NR,AngadiK, JadhavS, Vyawahare C, Hatolkar S.Prevalence and Antifungal Susceptibility Profile of Candida Species Isolated from TertiaryCareHospital,India.JournalofPharmaceuticalandBiomedicalSciences.2015Oct16;5(10).
- 5. Wey SB, Mori M, Pfaller MA, Woolson RF, Wenzel RP. Hospital-acquired candidemia:the attributable mortality and excessions of internalmedicine. 1988Dec1;148(12):2642-5.
- 6. GuptaP,PrateekS,ChatterjeeB,KotwalA,SinghAK,MittalG.Prevalenceofcandidemia inICU in a tertiary care hospitalin North India.International JournalofCurrentMicrobiology and Applied Sciences. 2015;4(6):10.
- KuMar D, Bhattacharyya S, Gupta P, Banerjee G, Singh M. Comparative analysis of discdiffusion and Etest with broth micro-dilution for susceptibility testing of clinical Candidaisolates against amphotericin B, fluconazole, voriconazole and caspofungin. Journal ofclinicaland diagnostic research: JCDR. 2015 Nov;9(11):DC01.
- 8. Clinical and Laboratory Standards Institute: M27-A3 Reference Method for Broth DilutionAntifungal Susceptibility Testing of Yeasts; Approved Standard Third Edition. CLSI, Wayne, 2008.
- 9. Clinical and Laboratory Standards Institute: M60-ED 1: 2017 Performance Standards forAntifungalSusceptibilityTesting ofYeasts. CLSI,Wayne, 2017.
- 10. Negri M, Henriques M, Svidzinski TI, Paula CR, Oliveira R. Correlation between Etest®, disk diffusion, and microdilution methods for antifungal susceptibility testing of Candidaspecies from infection and colonization. Journal of clinical laboratory analysis. 2009;23(5):324-30.
- 11. Song YB, Suh MK, Ha GY, Kim H. Antifungal susceptibility testing with etest for Candidaspeciesisolatedfrompatientswithoralcandidiasis. AnnalsofDermatology.2015Dec 1;27(6):715-20..
- 12. Prabhakaran N, Umamageswaria SS, Mohan K. Prevalence of candidial infections with their antifungal susceptibility pattern in a tertiary care hospital. JPure Appl Microbiol. 2016 Dec 1;10:3173-81.
- 13. Denning DW, Kneale M, Sobel JD, Rautemaa-Richardson R. Global burden of recurrentvulvovaginal candidiasis: a systematic review. The Lancet infectious diseases. 2018 Nov1;18(11):e339-47..
- 14. Rosa EA. Oral Candidosis Epidemiology. InOral Candidosis 2015 (pp. 1-6). Springer, Berlin, Heidelberg.
- 15. ChaudharyU,GoelS,MittalS.ChangingtrendsofCandidemiaandantifungalsusceptibility pattern in a tertiary health care centre. Infectious Disorders-Drug Targets(FormerlyCurrent Drug Targets-Infectious Disorders). 2015Oct 1;15(3):171-6.
- Mathur P, Hasan F, Singh PK, Malhotra R, Walia K, Chowdhary A. Five-year profile ofcandidaemiaatanIndiantraumacentre:highratesofCandidaaurisbloodstreaminfections.Mycoses. 2018 Sep;61(9):674-80.
- 17. Gandhi V, Patel M. Prevalence of Candida species and its antifungal susceptibility isolatedfrombloodcultureattertiarycarehospital,Ahmedabad,India.Int.J.Curr.Microbiol.Appl. Sci.2017;6:884-92.
- 18. Geeta SH. Study of characterization & antifungal susceptibility testing of clinically significantCandidaspecies.Journalof EvolutionofMedicaland DentalSciences.2014Jun 2;3(22):5973-9.