

# Antidiabetic Potential of Herbal DPP-4 Inhibitors- A Molecular Perspective

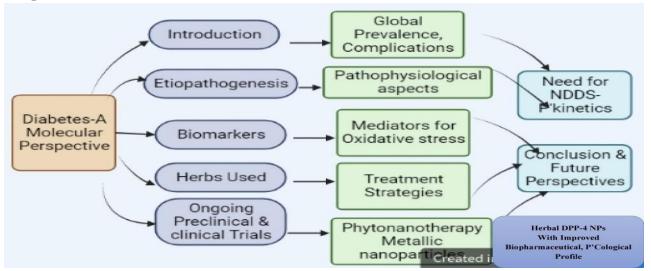
Priyanka Thakur, Vinay Pandit<sup>\*,</sup> Saurabh Gupta

Department of Pharmacology, Shiva Institute of Pharmacy, Chandpur, Bilaspur, HP, India- 174004<sup>1</sup> Department of Pharmaceutics, Laureate Institute of Pharmacy, Kathog, Kangra, HP, India- 176029<sup>2</sup> Department of Pharmacology, Chameli Devi Group of Institutions, Indore, MP, India- 452020<sup>3</sup>

E-mail: priyanka123thakur123@gmail.com<sup>1</sup>, saurabhgupta80@gmail.com<sup>2</sup>, vinay2121@gmail.com<sup>3</sup>

Abstract: Elevated plasma sugar is the foremost metabolic disorder around the globe, with an alarmingly rising mortality rate, and it exacerbates the patient's pathological condition. Following a research review, India had the world's greatest incidence of diabetes in 2000, with 31.7 million cases. Around 2030, the WHO forecasts that the global prevalence of diabetic instances might double. Multiple conventional hypoglycemic medications are adequate, yet they have a multitude of unpleasant side effects. Under the inclusion of numerous constituents in a plant with fewer negative impacts on peripheral organs, herbal novel formulations exhibit excellent synergistic efficacy. This study emphasizes on the biochemical constituents of medicinal herbs that inhibit Dipeptidyl peptidase-4 (DPP-4) enzymes which reduce blood glucose levels. The field of nanotechnology has an affordable price point and a higher safeguarding profile. The biosynthesis of herbal metal nanoparticles is one of the most promising approaches, with an outstanding pharmacokinetic profile, enhanced penetrability, and an environmentally friendly formulation. There are currently a handful of novel herbal formulations accessible on the market for sale. Regarding this, the authors of the present study investigated the molecular pharmacokinetic and pharmacodynamic profile status of medicinal herbs in numerous preclinical and clinical trials in future developments for their utility in treating this serious illness.

**Keywords:** Diabetes mellitus, Herbal drugs, Pharmacokinetic profile, Outstanding Pharmacodynamic status, DPP-4 inhibitors, Nanoparticles Formulations.



#### **Graphical Abstract:**

#### 1. Introduction

Diabetes mellitus is a disorder in which the body is unable to produce or respond to the hormone insulin, resulting in abnormal metabolism of carbohydrates which elevate the levels of glucose in the blood. It is a non-communicable illness and the fourth most cause of mortality as it complicates the micro and macrovascular (cardiovascular) health of patients who need frequent hospitalization due to their associated risk factors [1] [2]. The most significant risk factors for diabetes are overweight and obesity. This is characterized by lots of symptoms like hyperglycemia, hypercholesterolemia, polyphagia, polydipsia, lactic acidosis, polyuria, etc [3]. Plants have always been a rich arsenal for controlling and treating diabetes problems and complication arising due to it. The increase in resistance and populations of patients at some risk, in conjunction with the restricted number of commercially available drugs for diabetes that still have many side effects and problems like unwanted hypoglycemic effects, are the cause to shift the research towards traditionally available medicine which have low side effect and wide range of bioactivity [4] [5]. These traditional treatments do not require laborious pharmaceutical synthesis and seem highly attractive. Dipeptidyl peptidase-4 (DPP-4) is one of the widely explored novel targets for Type 2 diabetes mellitus patients (T2DM). The goal of DPP-4 inhibitors is to preserve the endogenous glucagon-like peptide (GLP)-1 activity by inhibiting the DPP-4 action [6] [7]. The DPP-4 inhibitors are weight neutral, well tolerated, and give better glycaemic control over a longer duration of time compared to existing conventional therapies. The journey of DPP-4 inhibitors in the market started with the launch of sitagliptin in 2006 onwards on to vildagliptin in 2022. Along with helping to control blood sugar it also boosts weight loss, GLP-1s seem to have major benefits [8]. Research has also investigated that drugs in these groups may also lower the risk of heart diseases such as heart failure, stroke, and kidney disease [9]. Nanotechnology opens the possibility for a wide variety of biological research topics and medical uses at the molecular and cellular levels. The biosynthesis of nanoparticles has been proposed as a cost-effective and environmentally friendly alternative to chemical and physical methods. This innovative approach might significantly enhance diabetics' quality of life [10]. **Diabetes Mellitus and its Types** 

Diabetes can be divided into Type I and Type II based on their etiological factors. Low or very low levels of circulating insulin are the major issues in type-I diabetes mellitus (IDDM/Juvenile onset), which is brought on by autoimmune beta cell death in pancreatic islets [11]. Whereas, in Type- II diabetes, beta cells have an aberrant glucoceptor that only reacts to high glucose concentrations also called Non-Insulin Dependent Diabetes Mellitus (NIDDM/ Maturity Onset Diabetes Mellitus) [12]. The "Down Regulation" of insulin receptors, which reduces the sensitivity of peripheral tissues to insulin, is another name for this disorder. A defective insulin molecule, an increased concentration of circulating antagonists, and abnormalities in the target tissue all contribute to resistance to insulin action [13]. Insulin receptors sensitization and desensitization sometimes get altered due to some biochemical or cellular changes in the pancreas because of various etiological risk factors through GLUT-4 which have the task to remove and balence glucose from blood plasma (Figure -1).

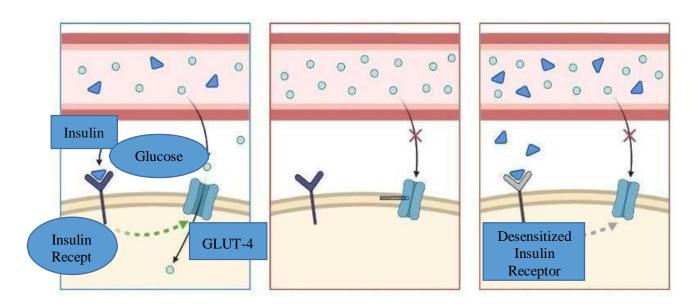


Figure No. -1 Pathogenesis of insulin receptor sensitization and desensitization in pancreatic beta cells to remove glucose from the bloodstream through GLUT-4 in a healthy person, Type-I and Type-II diabetic patients.

1. Healthy Person 2

2. Diabetes mellitus, type 1

3. Diabetes mellitus, type

Insulin insulin binds to receptors and triggers the opening of glucose transporters in fats/muscle cells, allowing glucose removal from the blood stream.

Insulin is not produced by beta cells in the pancreas and hence glucose is not removed from the bloodstream.

Prolonged overproduction of insulin leads to desensitization of the insulin receptors and hence glucose is not removed from the bloodstream.

# **Etiopathogenesis**

Different inflammatory mediators like Ικ- β, NF-κβ, TLR 4, MAPK, TNF, ROS, Macrophage, IL-1ß and IL-6 are released in this disorder which complicate a wide range of painful and inflammatory consequences, like stroke, retinopathy, neuropathy, and nephropathy [14]. Figure 02 shows h o w oxidative stress, mitochondrial dysfunction, hepatic glucose production, glucose uptakes, vesicular insulin resistance, and neurotransmitter dysfunction all increase in diabetics. They also diminish incretin effects, moreover, several inflammatory mediators are generated, including IkB-NF, NF-kB, TLR 4, MAPK, TNF, ROS, Macrophage, IL-1, and IL-6 [15]. Numerous unpleasant and inflammatory effects are brought on by these inflammatory mediators, including retinopathy, neuropathy, nephropathy, coronary artery disease, and peripheral arterial disease, which can result in stroke (Figure -2).

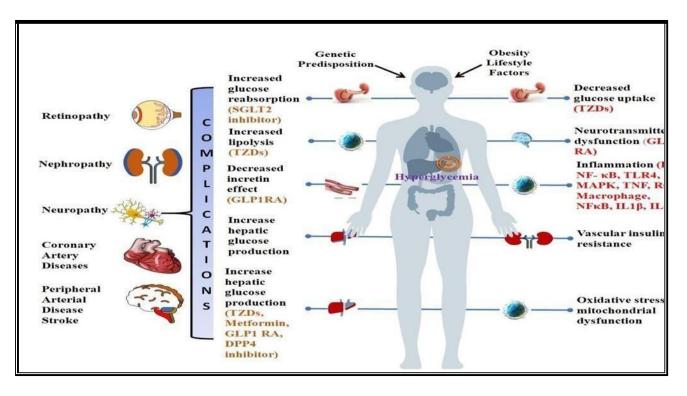


Figure -2 Pathophysiology of diabetes mellitus, risk factors, biomarkers, complications, targeted drugs, and their mechanism of action.

The primary factors for oxidative stress and mitochondrial dysfunction are genetic predisposition and obesity lifestyle, which increase glucose reabsorption, decrease incretin effects, increase hepatic glucose production, decrease glucose uptakes, vesicular insulin resistance, and neurotransmitter dysfunction [16].

Free radicals are implicated in the pathogenesis of diabetes [5] and, more importantly, in the onset of diabetic complications [17]. Free radicals are capable of causing damage to cellular molecules, DNA, proteins, and lipids, altering cellular functions. Acute or chronic hyperglycemia in diabetes increases ROS production and activates beta-cell apoptosis [18]. Under physiological conditions such as hypertension, ischemic heart disease, and diabetes, as well as during the aging process, reactive oxygen species (ROS) play a role in intracellular signal transduction of oxidative stress [19]. Numerous pathological processes involving protein and lipid peroxidation and DNA damage are well-known to be induced by oxidative stress [20]. Calcium cycling between the sarcoplasmic reticulum (SR) and the cytosol via the sarco-/endoplasmic reticulum Ca-ATPase (SERCA) pump, inositol-1,4,5-triphosphate receptor (IP3R), and Ryanodine receptor (RyR), plays a major role in agonist-induced intracellular calcium dynamics in vascular smooth muscle cells (VSMC) of diabetic patients [21]. The ATP-sensitive K+ channel (K ATP channel) detects metabolic alterations in the beta-cell of the pancreas, coupling metabolism to electrical activity and, ultimately, insulin secretion. When K ATP channels open, beta-cells hyperpolarize and suppress insulin secretion [22]. Ca2+ ion is an essential signaling molecule for the function of pancreatic beta-cells. Ca2+ is also essential for multiple beta-cell pathways, including insulin secretion, transcription, metabolism, endoplasmic reticulum function, and stress response. A novel approach could substantially improve diabetics' quality of life [23].

The below-mentioned diagram (Figure -3) depicts the interactions of insulin receptors and their route in controlling blood serum concentrations. It also depicts how oxidative stress and reactive

oxygen species inside beta islet cell becomes the etiological cause of Diabetes mellitus by some biochemical changes at a molecular level.

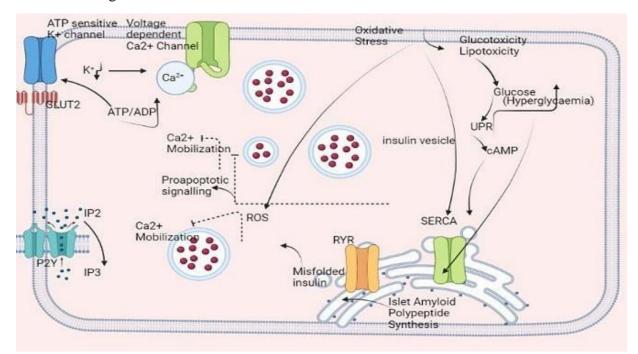


Figure -3 Pancreatic  $\beta$ -cell dysfunctioning results in the manifestation of Hyperglycaemia. Biochemical changes induced by oxidative stress up to ion

#### **Risk Factors**

Family history, polycystic ovarian syndrome (PCOS), low HDL, hypertension, age, ethnicity, diet, physical activity, overweight, and smoking are the risks associated with this metabolic disorder (Figure -4). In women age, waist, body mass index (BMI), hypertension, low education, and living environment; in men age, BMI, and hypertension were independently associated with an increased prevalence of diabetes [24]. Numerous risk factors contribute to the development of diabetes mellitus are diet, family history, overeating or obesity, smoking, ethnicity, genetics, and hypertension [25].



Figure 4: Risk Factors Accountable in a Diabetic Victim.

# **Global Prevalence**

Diabetes-related deaths accounted for 3.1 percent of all deaths in India in 2016, up from 0.98 percent in 1990. A recent study found that diabetes mortality will double in the upcoming five years and these patients in India invest for their care total of approximately 10,000 rupees in urban areas and 6260 rupees in rural regions per year [26]. It was found that around 462 million individuals have TD2, and by 2025, this figure may probably rise up to 600 million. It was also analyzed that between the age group of 20 and 79 years, 537 million people (or 1 in 10) have diabetes mellitus [27]. This number may be projected to be 643 million by 2030 and 783 million by 2045. China is the country with the highest rate of diabetic cases in the world, where about 141 million people are affected [28]. Table Chart 1 lists the nations with the greatest estimated incidence of diabetes.

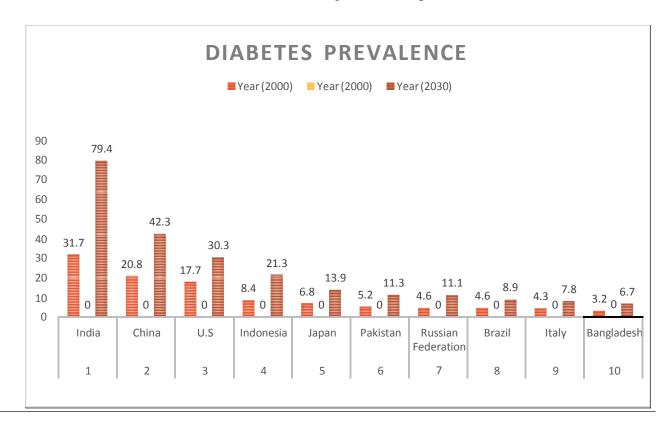


Table Chart 1. List of nations where diabetes is thought to be most prevalent

#### **DPP-4** Inhibitors

DPP-4 inhibitors, a class of oral diabetic medications, inhibit the DPP-4 enzyme, which is a unique enzyme present on the cell surface and deactivates the maximum number of polypeptides (GLP-1) and glucose-dependent insulinotropic polypeptides [29]. DPP-4 inhibitors are superior therapies based on the GLP-1 receptor; they reduce stomach emptying and food intake, increase insulin secretion dependent on blood glucose, and suppress glucagon post-meal release. GLP-1 (glucagon-like peptide 1) is an intestinal hormone produced in response to diet by inducing hypoglycemia and stimulating insulin secretion from the pancreas. DPP-4 inhibitors may be used as monotherapy in patients who are intolerant to metformin or have contraindications to its use, such as those with chronic renal disease and a significant risk for hypoglycemia [30].

The contraption of DPP-4 Inhibitors is explained in Figure-5 below, which shows how, after eating, our GI Track is employed to block or release GLP-1 from pancreatic beta cells, which activate and release insulin. DPP IV hydrolyzes incretin, a key regulatorof postprandial insulin secretion. This hormone aids diabetics in controlling their blood sugar. A DPP-4 inhibitor, sitagliptin boosts insulin production and lowers the overproduction of glucose in the liver. Raising the levels of active incretins, it prolongs the effect of GLP-1 and GIP. It prevents the DPP-4 enzyme from increasing the activity of GLP-1 and GIP hormones, which causes insulin secretion to longer lasting [31].

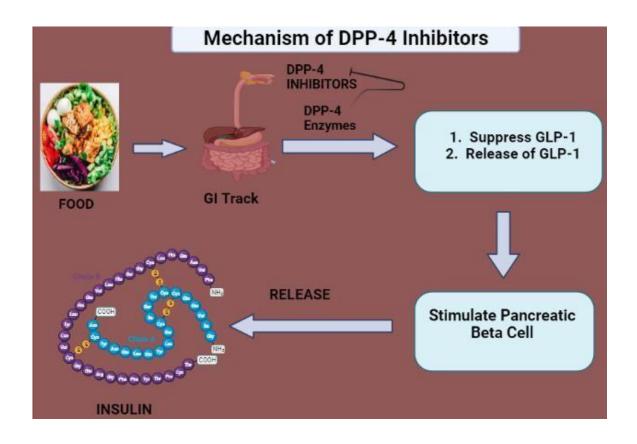


Figure -5: Sitagliptin's mechanism of action is depicted in a figure along with other DPP-4 inhibitor medications and GLP-1.

The best quality of DPP-4 inhibitors is even their adverse effect is also helpful as it leads to weight loss in victims. Therefore, in this review, we have focused on the pharmacokinetic and pharmacodynamic status of DPP-4 inhibitor herbal drugs both in animal and human patient research studies.

Whereas, Sulfonylurea (Biguanides) primarily attempts to block ATP-sensitive K+ channels in the plasma membrane of beta cells shown in Figure -6 to start a series of events in the pancreas. Insulin is released when sulfonylurea binding changes the resting potential of the cell and causes calcium influx [32]. Sulfonylureas and other conventional hypoglycaemic drugs have 2numerous adverse effects like hypotension, increase hunger, weight gain, dizziness, confusion, heartburn, stomach upset etc [33].

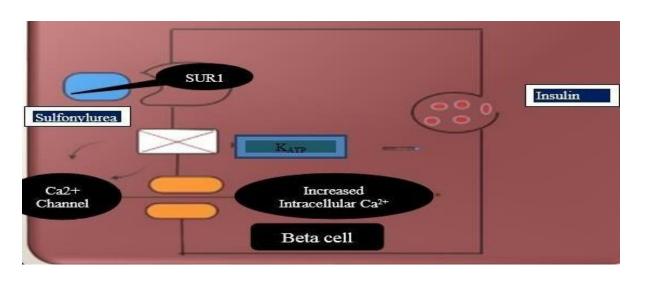


Figure -6 Diagram illustrating the operating procedure of Sulfonyl Urea in Pancreatic  $\beta$ - Cell by deliverance of insulin through the actuation of ATP-sensitive K<sup>+</sup> and Ca<sup>2+</sup> ion channels inside pancreatic islets.

# **Herbal Drugs**

In contrast to conventional medicine, the use of botanicals for the treatment and prevention of diseases, including diabetes, has a long history. Herbal medicine has been utilized for centuries to treat a variety of health conditions. If someone wishes to avoid prescription medications and invasive treatments as much as possible, herbal medicine may be a suitable alternative [34]. There are several benefits to using herbal medicine to treat diabetic illness, including reduced adverse effects, lower costs, improved overall health, and self-healing [35]. However, the selection of botanicals may depend on many variables, such as the stage of progression of diabetes, the categories of comorbidities the patients have, their availability, affordability, and their safety profile [36]. Numerous herbal treatments are recommended for diabetes and diabetic complications. However, there are very few herbal treatments for diabetes on the market, so more research is required to demonstrate their efficacy and ensure their safety [37].

#### Nanotechnology

The conventional treatment options for type 2 diabetes mellitus (T2DM) are extensive. However, conventional medications have some disadvantages that can impact the treatment's efficacy. Low bioavailability and immediate drug release are the primary disadvantages of the ground in the development of novel delivery modalities that could enhance the efficacy of anti-diabetic regimens [38]. Pharmacies and other medical corporations use nanotechnology applications worldwide in a range of biotechnology, bioengineering, and biomechanics disciplines, replacing the current scenario of researchers pursuing this field as an alternative drug delivery system [38]. As a result, nanotechnology is now one of the most intriguing academic disciplines. Nanomedicine and Nano delivery utilize nanoscale materials as diagnostic instruments for the targeted delivery of therapeutic agents. This field makes it possible to design biomaterials with the desired Nanoscale size, shape, and morphology due to increased bioavailability, prolonged release, and decreased dosing, thereby improving patient compliance [39]. This novel approach could significantly improve diabetics' quality of life. Existing oral treatments for T2DM

necessitate an increase in administration frequency. In tandem with the onset of adverse side effects, patient adherence to treatment decreases. Nanotechnology has found favor in recent years [40].

This review gives a thorough update to address the worldwide crisis, stressing the global situation and the statistical incidence. It became a need for future diabetic laypeople, as it ranks the fourth most prevalent cause of death in the nation. So there should be more and more research and production of novelherbal medicines in this era.

#### **Biomarkers**

The acronym biomarker pertains to a trait that may be objectively tested and assessed as a predictor of pathogenic biological processes, healthy biological processes, pharmacological reactions to therapeutic interventions, or chemicals that seem to be biological manifestations of a disease. Understanding the illness spectrum is made easier by biomarkers [41, 42]. Biomarkers are generally utilized in epidemiological research to understand the risk factors and problems linked to illnesses, identify the etiological and pathophysiological variables, and diagnose the disease early [43].

Many blood indicators for diabetic pathology are present. Blood analysis is necessary to determine the pathological and physiological aspects of the illness [44, 45]. The acute phase response is triggered by two crucial pro-inflammatory cytokines, TNF- $\alpha$  and IL-6, which boost the production of CRP and AGA among other proteins. TNF- $\alpha$  serum and urine concentrations are greater in DM3 patients compared to normoalbuminuric subjects or healthy controls. According to recent studies, serum TNFR1 and TNFR2 concentrations may be utilized to predict how diabetics' renal illness may develop in the future [46, 47].

#### **Biomarkers in Body Fluids**

Many biomarkers that show pancreatic beta-cell dysfunction and disease development may be identified in blood plasma, blood serum, and urine. These biomarkers include Lipoprotein Lipase, LDL, VLDL, IDL, Apo proteins, leptin, TNF-, and IL [48]. All of the biomarkers for diabetes mellitus are included in Table 02 below, along with their mediators, molecules, samples, and methodologies [49]. The diverse biomolecule levels in the various bodily fluids, such as blood plasma, serum, and urine, become unbalanced in diabetes mellitus, a liver and pancreatic condition [50]. Also, table 2 lists the adverse responses as well as the clinical result and numerous problems. Elevated TNF- and IL-6 impair the body's system and contribute to diabetic nephropathy and other cardiovascular risk factors [51]. The table describes the biomolecules and mediator changes in various biofluid samples using various methodologies. According to the results of the available literature, diabetic beta cell dysfunction is characterized by an imbalance in the levels of lipoprotein lipase, apolipoproteins E, cholesterol, and TNF- [52].

Since the metabolism of amino acids is significantly affected in pre-diabetes and continues to change as T2DM progresses, amino acids have been suggested as potential diagnostic indicators [53]. Due to greater blood concentrations in T2DM patients, tryptophan and branched-chain amino acids (BCAAs, which include valine, leucine, and isoleucine) may be particularly helpful as T2DM biomarkers [54]. In T2DM/pre-diabetes patients and healthy controls, there were statistically equal amounts of LPC (C18:2), palmitic acid (C16:0), alanine, citrulline, glutamate, glycine, isoleucine, leucine, lysine, phenylalanine, proline, serine, tyrosine, and valine amino acids [55]. Pre-diabetes and T2DM are very different from one another, showing that different disease stages are connected to unique and distinct metabolic biomarker profiles [56].

Sr.	Biomarkers	Mediators	Molecules	mples	Methods	Comments	Reference
No.	(Biofluids)			•			
			Soluble leptin-R	,		Biomarkers	
		Leptin system,	Interleukin (IL-	-	ELISA	correlation structure	
1.	Bloodplasma	Monoclonal	6), Hs- CRP, C-	Blood	Immunol	was disturbed before	[57]
		sandwich	Peptide,	Plasma	turbidime	the clinical diagnosis	
			HbA1C		tricassay		
			Lipo proteir				[58]
			lipase,				
			Apolipoprotein				
		Lipids, proteins	E,			An increase in	
	Blood serum	for lipids	Apolipoprotein	Blood	-	Copolipoprotein	
		metabolism	A1, A2,	serum		develops coronary	2
			Apolipoprotein			artery disease, MI	
			B, Cholesterol 1				
2.			esters				
			transferase,				
			proteins				
			nicroglobulin (a1	erum,	-	plicate liverdisease	[59]
			m)	urine			
		Cytokine	tin, TNF-α,IL-6	Blood			[60]
		related proteins		serum	-	-	[61]
		mmune-		Blood			[62]
		related	MCP-1	serum	-	-	[63]
		proteins					
			α1- Complement				
			factor, Alpha-1,				
			antichymo				[64]
			trypsin				[65]
		Defense	precursors,			Urine	[66]
		response	Antithrombin-				[67]
3	Urine		III, α-2,				[68]
			Glycoprotein-I,				
			Vitronectin				
			Precursors				

Table 2:- Biomarkers of Diabetes mellitus along with their mediators, molecules, sample, and their methodology

	21.22		Mi-RNA	[7]
n-codingRNA	21-23	iRNA	progression from	-
	Nucleotides	-	-	[73
			T2DM	[74
				[7:
				[70
	Transferrin		Elevation	[7]
Glomerular	Type- IV	Urine -	-	[78
			diabetic	
	Collgen		nephropathy	[7
				[8
				[8
			TNF- α elevation &	[8]
Inflammato	TNF-α	Urine -	IL-6 ccomplicate	[8]
Ry		/serum	diabetic	[84
			nephropathyy	
			Immunological	[8
		Blood	biomarkers lead to	[8
Genotype	HLA-DR/-DQ	plasma -	the development&	
			1 0	[8]
			T1DM	
Proteomic	GAD IGRP IL-0	6Blood -		[8
	SOB-R	plasma		[9

# **Principles of Preclinical Studies**

Before a trial on humans, medications must first undergo preclinical testing on animals. To be able to establish the pharmacological and toxic effects of a therapeutic molecule, preclinical testing utilizes the advantages of both in-vitro and in-vivo testing [91]. Preclinical research is intended to ascertain the initial dosage, safe dose, adverse drug responses, and toxicity of a drug product, which often includes a novel medical device, prescription medications, and diagnostics [92] [93]. The effects of synthetic drug compounds evaluated on rats are listed in Table 3; their effects and treatment results are noted for various time horizons (7 days, 21 days, 8 weeks, etc.) [94].

**Table 3:** Preclinical studies, their effects, treatment, and results are noted for various time horizons

Sr.	Herbal	Animals	Dose	Duratio	Side effects	Mechanism/ Molecular Targets	Refe
No	Drugs			n of			renc
•				study			e
1.	Acacia	36 female	100,	21 days	Unpleasant	Hypoglycaemia by initiating	[95]
	arabica	albino	200		mouth sensation,	release of insulin from pancreatic	

							1
		rats	mg/kg		morning sickness, slight diarrhea, and bloating.	$\beta$ cell, Reduction in blood glucose, TC and LDL	
2.	Aegle marmelo sa (Methan olic extract)	Wistar rats	250, 500 mg/kg	21 days	Diarrhea, indigestion, constipation and dysentery	Weight gain, blood sugar, lipid profile, C-Peptide, HbA1c, insulin secretion, and pancreatic insulin levels are all affected by the decline in beta cell density and disturbance of the normal architecture of the pancreatic beta cell.	[96]
3.	Allium cepa	Rats	300 mg/kg	45 days	Indigestion, bleeding disorders	Reduce levels of blood sugar, elev ate insulin levels, and lessen the pr oduction of advanced glycation en d products (AGE) Regulate cholesterol levels, blood sugar, and the liver's hexokinase, g lucose-6-phosphatase, and HMG- co-A-reductase enzymes	[97]
4.	Allium sativum	Rabbits	Aqueou s homoge nate of garlic 10ml/k g/day	2 months	Bad breath. Body odor. Burping. Heartburn. Upset stomach	Extremely reduced AST, urea, uric acid, total cholesterol, and other blood markers Hypoglycemia, increased hepatic metabolism, insulin release, hepatic glycogen, and free amino acids	[98]
5.	Aloe vera and barbeden sis	Mice	200 mg/kg	2 weeks	skin irritation, hives, cramping, and diarrhea	Boosts the body's tissues' receptivity to insulin, increasing the effectiveness of the hormone. encourage the pancreas to produce or release more insulin	[99]
6.	Azadirac hta indica (hydroal coholic extract)	Rats	400 mg/kg	30 days	death, vomiting, abdominal pain, sleepiness, blood disorders, seizures, loss of consciousness, coma, brain diseases, and brain disorders	adjusted the changing concentrations of GLUT4 proteins, serum insulin, lipid profile, blood sugar, and insulin signaling molecules. Method to increase glucose absorption and glycogen synthesis in the rat hemi diaphragm	[100]
7.	Caesalpi nia bondcell a (85% ethanolic	Rats	250, 500 mg/kg	8 weeks	abdominal pain, colic, leprosy, fever	By restoring free radical scavenging capacity, blocking glucose absorption causes hypoglycaemic and hypolipidaemic activity, which	[101]

[	extract)					reduces oxidative stress in	
	extract)					pancreatic cells.	
8.	Coccinia	Rats	500 mg/	6 weeks	None	restore lipoprotein lipase, lower	[102
0.	indica	Rais	kg	U WEEKS	NONE	lactate dehydrogenase, glucose-6-	-
	mulca		кд				]
						1 1 '	
						hypoglycemia	
						Alloxan increases superoxide	
						dismutase, decreases oxidative	
						stress, and increases lipid levels in	
-		-				the RBS, kidney, and heart.	5100
9.	Eugenia	Rats	400mg/	2 weeks	bodily pains,	free radical scavenging, as shown	[103
	jambola		kg		fever, phlegm	by increased oxidative stress,	]
	na				buildup in the	raised catalase, glutathione	
	(Decocti				lungs, and	peroxidase, glutathione-s-	
	on				coughing	transferase, and SOD activity, a	
	extract					73.5% decrease in sugar, a rise in	
	of jamun					blood insulin levels, and an	
	pulp)					inhibition of insulinase activity in	
						the liver and kidney	
10.	Mangfer	Rats	200	30 min.		boost muscle and liver glycogen	[104
	a indica		µg/mL		stomach pain,	stores, significantly reduce alpha-	]
	(oral				indigestion and	amylase activity, and significantly	
	aqueous				diarrhoea	improve glucose absorption in	
	extract)					diabetic groups to affect the	
						glycogen production pathway.	
11.	Momord	Langurs	200	21 days	boost muscle and	enhancing the sensitivity of	[105
	ica	(s.c.)	mg/kg		liver glycogen	insulin, Sugar, amino acids, TC,	]
	charantia				stores,	TG, and total lipids are all reduced	
	(extract				significantly	Hepatic fructose-1,6-	
	of fruit				reduce alpha-	biphosphatase, glucose-6-	
	pulp,				amylase activity,	phosphatase, and glucose-6-	
	seeds)				and significantly	phosphate dehydrogenase	
	,				improve glucose	activities are all inhibited.	
					absorption in		
					diabetic groups		
					to affect the		
					glycogen		
					production		
					pathway.		
12.	Ocimum	Rats	200mg/	30days	severe headache,	substantial reductions in fasting	[106
	sanctum		kg	-	hallucinations,	and postprandial blood glucose	]
	(aqueous		-		convulsions,	levels, improved pancreatic beta-	
	extract				irregular	cell activity, and insulin secretion	
	of				heartbeat, and	Skeletal muscle and hepatic	
	leaves)				extreme	glycogen levels decline by 68 and	
					dizziness	75%, respectively, but renal	
L						in the second se	

						glycogen content increases tenfold.260	
13.	Phyllant hus amarus (methan olic extract of plant)	Rats	600 mg/kg	21 days	increased risk of bleeding	extract improved adipogenesis in 3T3-L1 fat cells and raised deoxyglucose absorption in C2C12 muscle cells. strong antioxidant effects and a reduction in sugar levels	[107 ]
14.	Pterocar pus marsupi um (Tannate s extract	Male albino rats	250, 1000 mg/kg	10 days	None	stimulated the insulin secretion and glucose uptake Pancreatic beta cells regranulation, hypolipidaemics, stimulate oxygen uptale in fats cells and tissues, increase glycogen content of rat diaphragm	[108 ]
15.	Trigonel la foenum graecum	Wistar male rats	1.6 g/kg	60 min	Dizziness and headaches are sporadic, along with symptoms of the digestive system such as diarrhea and nausea.	Limit the activity of fructose-1,6- bisphosphate, glucose-6- phosphatase, and the liver Reduce blood sugar levels in both healthy and diabetic rats while enhancing glucose metabolism normalize the levels of liver, skeletal muscle, and cardiac CK in these tissues	[109]
16.	Tinospor a cordifoli a (Guduch iroot extract)	Rats	200, 400 mg/kg	30 days	headache or nasal pain	Strengthening the liver's ability to store glycogen or reducing its ability to release glucose. decrease in blood sugar and triglycerides, hypoglycemic activity, and improvement in glucose tolerance	[110 ]
17.	Urtica diocia	Wistar male rats	100, 200, 300 mg/kg	21 days	minor stomach discomfort, bloating, diarrhea, perspiration, and hives or rash	Leaves have insulin secretagogue, PPAR gamma agonistic, and alpha-glucosidase inhibitory effects Reduction in fasting insulin resistance index and increase sensitivity of tissues	[111]
18.	Anacardi um occident ale leaves extract	Rats	100 mg/kg	30 days	bloating, constipation, weight gain, and joint swelling	levels of glycated hemoglobin, insulin, fasting blood sugar, and lipid markers have decreased. a decrease in fasting sugar, a rise in serum insulin, and an improvement in skeletal muscle glycogenesis	[112]

					1		r
19.	S.	Mouse	1000	28 days	constipation and	significantly decreased fasting	[113
	cochinch		mg/kg		excessive	blood glucose	]
	inesis				urination	Decrease homeostatic model score	
						on insulin resistance index,	
						increase sensitivity of tissues	
20.	Helicter	Rats	200,	28 days		enhancing microcirculation,	[114
	us		400		hepatotoxicity,	raising insulin secretion,	]
	angustif		mg/kg		nephrotoxicity	controlling glycemic metabolism,	
	olia				and	lowering cholesterol, removing	
	(ethanol				hypersensitivity	free radicals, and all of the above	
	extract)				JI	Lower HOMA-IR and blood sugar	
	childer)					levels, higher insulin levels	
21.	Pleurotu	Rats	100,	4 weeks	hives, fever,	alkaline phosphatase (ALP),	[115
21.	s	Rats	200,	+ WCCR5	chills, soreness,	aspartate aminotransferase (AST),	1
			400		, , ,	and alanine aminotransferase	1
	ostseatus				itching, or		
	(aqueous		mg/kg		swelling in the mouth or throat	(ALT) all showed increased liver function.	
	extract)				mouth or throat		
						raising insulin levels, and	
- 22	A.C. 1'		100	10.1	1 .	enhancing cellular sensitivity	5116
22.	Afzelia	Rats	100,	10 days	drowsiness,		[116
	africana		200		dizziness,	Pancreatic tissue regeneration	]
			mg/kg		hypotension or a	stimulate inintracellular glucose	
					headache	transport, increase glucose uptake	
						by various cells	
23.	Uvaria	Sprague	100,	8 days	kidney stones	Extracts improved HDL-C, lipid	[117
	chamae	dawley	250,			profiles, and blood glucose	]
	roots	Rats	400			levels.Hypoglycaemic effects in	
			mg/kg			dose dependent manner,	
						regeneration of islet of	
						Langerhans, enhance insulin	
						sensitivity	
24.	Camellia	Rats	60, 120	7 days	jitteriness,	Examining their impact on cellular	[118
	sinesis		and		nausea, skin	glucose absorption and fat	]
			480 mg		rashes, and liver	accumulation, ameliorates insulin	
			/ml		toxicity	secretion, inhibits the DPP-IV	
						enzyme, enhances glucose	
						tolerance, and increases active	
						GLP-1	
						-amylase inhibition, decreased	
						starch digestion and absorption,	
						and sod inhibition. dependably	
						transporting glucose	
25.	Cinnamo	Wistar	150,	7 days	diarrhea,	Nrf2, PI3K/Akt and MAPK	[119
	num	albino	200		vomiting,	pathways activate	]
	zeylanic	rats	mg/kg		dizziness,	Inhibit $\alpha$ -glucosidase activity,	1
1	ZUYIAIIIU	Tais	mg/ Kg		U122111055,	minut u-glucosluase activity,	

	um				drowsiness	chronic malabsorption of carbohydrate, suppression of post meal glucose	
26.	Callistep hus chinesis flower (ethylace tate extract)	Rats	2 mg/ml	7 days	GI disturbances	decreasing fasting glucose levels, increasing insulin release by triggering beta cells or sensitizing insulin Inhibit $\alpha$ -glucosidase due to quercitin	[120 ]
27.	Corchor us olitorius (jute)	Rats	100, 200 mg/kg	120 minutes	swelling of the mouth or lips, and respiratory problems	stimulation of insulin secretion, increasing $\beta$ -cell proliferation, thus promoting insulin sensitivity Manage postpramdial hyperglycaemia, inhibit $\alpha$ -amylase and $\alpha$ -glucosidse	[121 ]
28.	Holarrhe na antidyse ntrica	Rats	300 mg/kg	21 days	Nausea · Flatulence · Constipation · Anxiety, nervousness, and sleeplessness, · Vertigo	stimulation of histaminergic receptors and $Ca^{++}$ antagonist mechanisms Inhibit $\alpha$ -amylase, $\alpha$ -glucosidse in dose dependent manner	[122]
29.	Ficus deltoidea	Mouse	125, 250 and 500 mg/kg/ d	4 weeks	skin allergies, kidney and liver function	regulating blood sugar, blood pressure and cholesterol levels Inhibit intestinal $\alpha$ -glucosidse and improve insulin mediated glucose uptake into adipocyte	[123 ]
30.	Olea europaea L (alcoholi c extract)	Male Wis tar rats	200, 400 mg/kg	10 weeks	dizziness, lightheadedness, stroke, and even kidney failure	decrease post-prandial blood glucose Reduced starch breakdown and absorption in vivo, lowers fasting plasma insulin level and Hb in placebo patients	[124 ]
31.	(Free and bound phenolic extract) Glycine max L	Rats	200, 500 mg/kg	28 days	nausea, vomiting, and upset stomach	strong nutritional value, a low glycemic index, and the capacity to reduce blood sugar levels inhibition of glucosidase and amylase	[125]
32.	Catharan thus roseus	Mice	500, 1000 and 2000 ppm	20 weeks	nausea, vomiting, hair loss, hearing loss, dizziness, bleeding, nerve	prolonged action in reduction of blood glucose Enhance intestinal glucose uptake, increase glucose-6-phosphate dehydrogenase, facilitate	[126 ]

	aromatic um						
	Carria						
	verum and				dizziness		
57.	mum	11100	mg/kg	I WOOKS	vomiting,	modulate insulin signaling	]
39.	Cinnamo	Mice	150	4 weeks	other allergic reactions diarrhea,	adipose tissues, decrease fats oxidation Increase of hepatic glycogenesis,	[133
38.	Gastrodi a elata	Rats	20 mg/kg	8 weeks	skin allergies, hair loss , and	Reduce insulin resistance through decrease in fats accumulation in	[132 ]
37.	Astragal us membra naceus	Mouse	200 mg/kg	3 weeks	rash, itching, nasal symptoms, or stomach discomfort	Increase better glycemic control by increasing insulin sensitivity, Akt activation and upregulation of GLUT-4	
36.	Dioscore a polysach aries	Rats	100 mg/kg	3 weeks	stomach, and headache	Improve insulin resistance, diminution of phosphorylation of ERK, increase of GLUT-4 transporters	
35.	Coccinia grandis (ethanoli c extract)	Male Wistar rats	5 mg/kg	9 days	Nausea, headache, and drowsiness	a dose-dependent rise in blood insulin level	[129 ]
34.	olic and aqueous extract) Forsythi a suspense (Ethyl acetae fraction of methano l extract)	Male kunning mice	50, 100 and 200 mg/kg	4 weeks	Fever. · Gonorrhea,Heart disease HIV/AIDS, Nausea and vomiting, Pain and swelling	Reduced levels of hepatic lipid, triglycerides, acid phosphatase, alkaline phosphatase, aspartate transaminase, and total cholesterol were also seen. Significant improvements were seen in glucose tolerance.	[128]
33.	Chlorox ylon swieteni a (Methan	Male albino wistar rats	250 mg/kg	45 days	None	Moderate reduction in blood glucose and glycosylated Hb level	[127 ]
					problems, seizures, liver damage	glycaemic control, catalyze the oxidation of malate to oxaloacetate, utilize glucose levels	

	chinesis		mg/kg		on the skin,	anti-oxidative effects	J
					swelling of the		
					lips and the		
					throat and		
41	<b>D</b>	24	10	0 1	diarrhea		5105
41.	Ervatani	Mice	10	8 weeks	constipation, dry		[135
	a		mg/kg		mouth, and	regeneration of pancreatic beta	]
	microph				insomnia	cell, decrease fibrosis of islet cell	
	ylla					and increase level of insulin	
42.	Anoecto	Mice	100,	25 days	hepatotoxicity,	Attribute beta cell repair or	[136
	chilus		300		reproductive	regeneration, beta cell survival	]
	roxburgh		mg/kg		toxicity,		
	ii				pruritus, rash,		
10	~	-			urticaria		5107
43.	Gymne	Rats	0.84µg/	3 weeks	headache,	boosts insulin production,	[137
	ma		ml		nausea,	encourages islet cell regeneration,	]
	sylvestre				lightheadedness,	and improves glucose uptake	
					shakiness and		
					dizziness		
44.	Cartham	ICR mice	120	6 weeks	bleeding issues	Enhance insulin secretion,	[138
	us		mg/kg		such stomach or	suppress $\alpha$ -glucosidase activity	]
	tinctoriu				intestinal ulcers,		
	S				hemorrhagic		
	(hydroal				illnesses, or		
	coholic				clotting		
	extract)	~	100		abnormalities		51.00
45.	Momord	Sprague	100		vaginal	Increase insulin secretion from	[139
	ica	Dawley	mg/kg		bleeding,	beta cells, inhibit glucose	]
	charantia	rats			contractions, and	reabsorption and peripheral	
					other	glucose in gut, suppression of	
					gastrointestinal	gluconeogenic enzyme	
					symptoms		
46.	Panax	Mouse	150	12	gastrointestinal	Reduction in insulin resistance,	[140
	ginseng		mg/kg	weeks	discomfort,	improve beta cell functions,	]
					euphoria,	protect beta cell apoptosis,	
					sleeplessness,	antioxidant activity by up	
					headaches, high	regulation of glutathione	
					or low blood		
					pressure,		
					mastalgia, and		
					vaginal bleeding		
47.	Curcuma	Mouse	250	2 weeks	stomach upset,	Lower insulin resistance and	[141
	longa		µg/ml		nausea,	inhibit $\alpha$ -glycosidase activity	]
					dizziness, or		
					diarrhea		

48.	Zingiber	Mouse	250	4 weeks	belly discomfort,	Islet cell protection, increase	[142
	officinal		mg/kg		diarrhea, or	insulin receptor signalling	1
	e		0.0		heartburn, as		
					well as a burning		
					sensation in the		
					mouth or throat		
49.	Ribes	Rats	100,	7 days	Headache.	Decrease blood sugar level,	[143
	nigrum		500		Diarrhea.	enhance glucose tolerance,	]
			mg/kg		Gas and belching	inhibition of $\alpha$ -glucosidase and $\alpha$ -	
					Gas and berchning	amylase acivity, delay	
						carbohydrate digestion and	
						absorption in intestine	
50.	Daucus	Rats	250,	14 days	kidney damage	Reduce diabetic nephropathy,	[144
	carota		500		and nerve	decrease serum LDL, VH levels	]
	linn.		mg/kg		problems		
51.	Boswelli	Mouse	10 mg/	21 days	stomach pain,	Increase mRNA, protein	[145
	a serrate		kg		nausea, diarrhea,	expression of PPAR-y, increase	]
					headache,	lipoprotein lipase	
					heartburn, and	Increase glucose uptake in insuin	
					itching	sensitive cells, increase stimulation	
						of insulin pathways mediators	
52.	Myrtus	Rats	25,		feeling nauseous	Reduced serum levels of glucose,	[146
	commun		50µg/m		hypotension, and	triglycerides, urine volume, and	]
	is		1		problems with	protein due to -glucosidase	
					blood flow	inhibitory action	
53.	Postulac	Male	250	4 weeks	Kidney stones	decreases blood sugar and controls	[147
	a	wistar	mg/kg			how blood lipids and glucose are	]
	oleracea	rats				metabolized	

#### Herbs used in diabetes management

For many years, researchers have used plants as a significant resource to create new medications for human illnesses. This section [148] contains just a small percentage of the hundreds of plants that have been studied for diabetes. The many phytochemical qualities of the herbs are also covered in the paragraph that follows, which will aid researchers in determining whether to utilize a certain herb to treat a particular medical condition [149].

There are several methods to minimize the bad effects of diabetes and the difficulties that follow, but herbal formulations are preferred since they are less costly and have less side effects [150] [151]. Approximately 60% of individuals use conventional medications manufactured from medicinal plants. Diabetes is a dangerous condition that has a broad range of effects on individuals globally [152]. It has been shown to be a serious health problem in India, especially in urban areas [153]. There is a list of medicinal plants with recognized antidiabetic characteristics and their favorable impact on herbal diabetes therapies [154]. In the past, medicinal plants have mostly been used to discover new solutions for human health conditions [155]. In the past, several plants have been recommended for the treatment of diabetes. Moreover, several researches have noted that

some plants contain anti-diabetic properties [156]. While there is little proof of their therapeutic utility, these claims are often supported by animal models and even in vitro investigations [157]. The emphasis of the present research was on medicinal herbs, whose hypoglycemic effects have been verified by several preclinical and clinical investigations on diabetic patients [158].

### 1.1 Acacia arabica (Babool)

In the Unani system of medicine, Babool (Acacia Arabica) is regarded as a plant with medicinal properties for various human body systems. The plant's medicinal properties include the bark, root, gum, leaves, pod, and seeds. Acacia Arabica (Babool) has been studied pharmacologically and phytochemically [159]. The creation of contemporary pharmaceuticals from Acacia arabica for the treatment of different ailments may be stressed, as pharmacologists eagerly anticipate the production of novel medications from natural sources. Numerous phytoconstituents, vital components of this plant's therapeutic significance, are present [160]. The purpose of the research was to determine if mice that had been given alloxan to cause diabetes may benefit from aqueous leaf extracts of Acacia nilotica. The findings demonstrate the anti-diabetic effect of aqueous leaf extracts from A. nilotica. Herbal extracts have been reported to be more efficient when administered intraperitoneally than orally [161].

Additionally, phenols, alkaloids, flavonoids, tannins, and saponins were found in the aqueous leaf extracts of A. nilotica by qualitative and quantitative phytochemical screening [162]. Acacia Arabica extract is utilized as an antioxidant, antihyperlipidemic, and hypoglycemic drug to study its effects on streptozotocin-induced diabetic rats [163]. The results of this investigation indicate that Acacia arabica extract possesses hypoglycemic, hypolipidemic, and antioxidant capabilities; as a result, its potential for treating diabetes in people may be explored [164].

#### 1.2 Aegle marmelosa

In Ayurveda, aegle marmelosa leaf extract is used to cure diabetes. It has been suggested that Aegle marmelosa contains anti-diabetic and antioxidant effects. The findings demonstrate that Aegle marmelosa extract significantly lowered alloxan-induced oxidative stress and reduced blood sugar [165]. The prominent medicinal plant Aegle marmelosa, often known as the bael tree, is used to treat diabetes in the Ayurvedic and Siddha systems of medicine as well as in folk remedies [166]. Additionally, the feasibility of using calluses made from A. marmelosa leaf explants for the treatment of diabetes was investigated [167] in comparison to regular plant material. The research found that the in vitro callus culture of A. marmelosa had the same potential for managing diabetes as the original leaf extract [168].

#### 1.3 Datura stramonium

A folk remedy used in Ethiopia, Datura stramonium Linn, has been demonstrated to have antidiabetic action in an in vitro research [169]. The research also aimed to evaluate hydromethanolic seed extract's anti-diabetic effects in mice. According to the research, Datura stramonium Linn hydromethanolic seed extract demonstrated strong antihyperglycemic and free radical scavenging capabilities [170]. Nature has given us a plethora of phytochemicals that may have an antidiabetic effect. Datura stramonium has traditionally been used to treat diabetes mellitus [171]. The researcher's wants to see whether the plant's hydromethanolic seed extract had any kind of antidiabetic properties in *in vivo* experiments [172]. The study revealed that Datura stramonium hydromethanolic seed extract possessed significant antihyperglycemic and antioxidant activity [173].

# 1.4 Piper longum (Piper longum)

All diabetic medications, including insulin and oral diabetics, have adverse effects. According to the WHO, it is challenging to identify new antidiabetic natural herbal drugs with minor or no side

effects [174]. In light of this, the goal of the present investigation was to determine if the aqueous root extract of Piper longum (PlrAqe) had any antihyperglycemic or hyperlipidemic effects on diabetic rats (STZ) produced by streptozotocin. The results show that in STZ-induced diabetic rats, plant extract may reduce diabetes complications. [175]. As a result, this species might be considered a potential source for identifying new oral antihyperglycemic agents with the introduction of novel and innovative, transformative technologies such as recombinant DNA [176]. According to the pharmacological profile, the plant has antidiabetic, antihyperlipidemic properties, hepatoprotective, neuroprotective, and antioxidant properties. It is cardioprotective, antibacterial, aphrodisiac relieves respiratory disorders and can also be used as a digestive agent [177].

### 1.5 Coriandrum sativum

The herb coriander, also known as Coriandrum sativum, has long been used to treat diabetes. The aim of this study was to analyze existing studies on the efficacy of coriander in treating diabetes in animal models using a database. The publications were found using PubMed [178]. It was confirmed that there is a high level of interestin diabetes research. It has also been discovered that approximately 6.92 percent of the articles containing the keyword "Coriandrum sativum" are related to diabetes [179]. It is possible to conclude and suggest that coriander may be appropriate for attempting to lower the plasma glucose level of diabetic animals [180]. To show the phytochemical makeup of a petroleum ether extract of C. sativum (CPE) seeds, we employed chromatographic, spectroscopic, and healthy spectrometric analyses in this work [181]. Finally, using the STZ-NAD model, we were able to correctly predict that the properties of bioactive components CPE would impede the development of DM. Further research on C. sativum is necessary since it has the potential to be used as an adjuvant for anti-diabetic treatment [182].

# 1.6 Zingiber officinale (Zingiber officinale)

One of the antioxidant supplements from the diet that are being evaluated as possibilities for treating diabetes is ginger (Zingiber officinale). The therapeutic benefits increase insulin sensitivity and shield the body from the negative consequences of diabetes [183]. The results show that ginger has hypoglycemic potential and lowers diabetic complications, thus it should be taken into consideration while treating this condition [184]. The antidiabetic activity of Zingiber officinale aqueous extract against Streptozotocin-induced diabetes in Sprague Dawley rats is the subject of a research. The results show that Zingiber officinale aqueous extract possesses hypoglycemic characteristics [185].

# 1.7 Cinamomum zeylanicum

An evergreen tree in the Lauraceae family called cinnamon (Cinnamomum zeylanicum) has been picked for generations in Asian nations. Many different ailments may be treated with cinnamon bark [186]. Numerous essential oils and tannins may be found in cinnamon. It possesses strong antiulcerogenic, anti-ulcerogenic, antipyretic, anti-diabetic, and antioxidant effects [187]. Administration of cinnamon essential oil dramatically decreased blood glucose levels in alloxaninduced diabetic mice. This effect may have been caused by the reversal of insulin resistance or by an increase in insulin production by mending injured pancreatic -cells [188]. Additionally, flavonoids are known to serve as insulin secretagogues and heal injured -cells in rats given alloxan [189].

# 1.8 Datura stramonium

A well-known medicinal plant from the Americas, Datura stramonium Linn (Astenagra in Amharic), belongs to the genus Datura and the family Solanaceae [190]. Following diabetes induction, measurements of the diabetic mice's fasting blood glucose levels were made once a week [191]. The seed extract substantially (p0.0l) decreased fasting BGL at dosages of 100 mg/kg, 200 mg/kg, and 400 mg/kg [192]. The hydromethanolic seed extract of Datura stramonium exhibited significant blood glucose-lowering and antioxidant activities. For the bioassay-guided fractionation, isolation, and characterisation of active substances with glucose-lowering activity, more study is required [193] [194].

# 1.9 Eugenia jambolana

In Indian traditional medicine, E. jambolana is often used to treat a range of illnesses. The kernel, leaves, and septum of E. jambolana, among other components, show antihyperglycemic effects [195]. It has been shown that the ethyl acetate portion of E. jambolana seed has antihyperglycemic activity [196]. The high solubility of bioactive phytomolecules such gallic acid and polyphenolic compounds may be the cause of the antihyperglycemic activity of E. jambolana's ethyl acetate fraction in both short- and long-duration models [197]. The ethyl acetate fraction of E. jambolana seed has strong anti-diabetic effect in experimental model rats at the genetic level [198].

#### 1.10 Trigonella foenumgraecum L.

All diabetic treatments, including oral hypoglycemics and insulin, have the potential to have negative side effects [199]. In accordance with WHO recommendations, it may be difficult to find novel anti-diabetic medications made from medicinal plants that have minimal or no negative effects [200]. In this regard, the goal of the research was to evaluate the effects of Trigonella foenumgraecum L. root aqueous extract (PlrAqe) on hyperlipidemia and hyperglycemia in streptozotocin-induced diabetic rats (STZ) [201]. The results support the hypothesis that the plant extract may control hyperglycemia and the consequences of diabetes in STZ-induced diabetic rats. This plant may thus be considered as a possible source for the creation of innovative oral hypoglycemic drugs with the introduction of fresh and inventive transformative technologies like recombinant DNA [202].

# 1.11 Linnaeus Ficus religiosa

The religious ficus, Ficus religiosa The peepal tree, commonly known as Linn, is a member of the Moraceae family. In Ayurvedic medicine, its bark is used to cure diabetes [203]. It has antidiabetic qualities due to the presence of phytosterolin in the root bark and -sitosterol-D-glucoside in the stem bark [204]. All of the animals' blood glucose levels were significantly lowered by the three

treatments. In comparison to 25mg/kg, the impact was more prominent at 50 and 100mg/kg. The FRAE also revealed a considerable rise in body weight, liver and skeletal muscle glycogen content, serum insulin, and rats with STZ-induced diabetes [205] [206]. The medication may be tested for the chemical constituent in charge of the action and utilized as an adjuvant in diabetes treatment [207].

# 1.12 Gymnema sylvestris (Gymnema sylvestris)

Central and peninsular India are home to the important Asclepiadaceae plant known as Gymnema Sylvestre [208]. The plant's extracts are potent natural diabetes medications. One of the plant's most important antidiabetic components is gymnemic acid, which gives the plant its therapeutic usefulness [209]. It possesses powerful anti-diabetic and anti-obesity characteristics, limits glucose absorption, and lowers body weight, according to several research. Gymnema has the potential to repair pancreatic beta-cells, making it a potential medication for the management of diabetes mellitus and its consequences. Gymnemic acids A and B have been shown to have antiviral properties against the influenza virus [210] [211]. A potential utility in avoiding the development of dental plaque has also been researched, however formal trials to support this usage are missing [212].

# 1.13 Hagenia abyssinica (Hagenia abyssinica)

Hagenia abyssinica (Bruce) J. F. Gmel is a member of the Rosaceae family's monospecific genus Hagenia [213]. It is also known as kosso and African rosewood. H. abyssinica extract ameliorates metabolic abnormalities associated with diabetes and may reduce the risk of hyperglycemia-related complications [214]. Consequently, it appears that the beneficial effects of H. abyssinica on diabetes are due to the synergistic effects of its bioactive compounds, including phenols, triterpenoids, flavonoids, saponins, and anthraquinones [215].

# 1.14 Momordica charantia L.

Momordica charantia L., also known as bitter gourd, melon, and karela, is an annual ascending plant in the Cucurbitaceae family. As research has progressed, numerous phytochemicals, such as saponins, polysaccharides, triterpenes, proteins, vitamins, minerals, flavonoids, ascorbic acid, and steroids, have been discovered [216]. In addition, antioxidant, hypoglycemic, antitumor, antibacterial, skin care, anthelmintic, neuroprotective, anti-inflammatory, antiviral, immune-modulatory, wound healing promoting, antimutagenic, antiulcer, liver protection, and antiobesity properties have been established [217]. The concentration of the saponin-rich component of Momordica charantia L. stimulated insulin secretion in MIN6 pancreatic cells [218]. The survival of RIN-m5F pancreatic cells treated with high glucose and MC aqueous extract was substantially greater than that of untreated glucotoxicity cells, according to an experiment [219].

# 1.15 O. sanctum

This plant is predominantly found in tropical and subtropical regions, including India. It is widely employed in the treatment of diabetes due to its hypoglycemic and antihyperlipidemic properties [220]. Glibenclamide plus O. Sanctum was more efficacious than oral hypoglycemic agents alone at reducing fasting and postprandial blood glucose levels in type 2 diabetes patients [221]. Glibenclamide and O. Sanctum decreased Glycosylated Haemoglobin (HBA1c) levels significantly [222]. The acute antidiabetic effects of O. sanctum ethanolic extracts were investigated in rodents with type 1 and type 2 diabetes induced by chemical means [223]. The effects of extracts on glucose absorption, intestinal disaccharide activity, gastrointestinal motility in rats with type 2 diabetes,

glucose uptake, and insulin action in 3T3- L1 cells were evaluated [224]. The extract (1.25 g/kg bw) significantly enhanced oral glucose tolerance and suppressed blood glucose elevation in both normal rats and rats with type 2 diabetes [225].

# 1.16 Piper longum

Piper longum has potent hypoglycemic and anti-lipid peroxidative effects in animals with alloxaninduced diabetes [226]. In our research, untreated diabetic rodents had elevated serum levels of urea and creatinine, which are significant indicators of renal impairment [227]. Following PlrAqe treatment, the concentrations of urea and creatinine in diabetic rodents were significantly reduced. PlrAqe appears to safeguard the kidneys of diabetic rodents [228] [229].

# 1.17 Zingiber officinale (Zingiber officinale)

Ginger (Zingiber officinale, Zingiberaceae) is among the most popular species in the world [230]. It has a lengthy history of use as a herbal medicine to treat a variety of maladies, including vertigo, pain, dyspepsia, and cold-induced syndromes, dating back to its Southeast Asian origins and subsequent dissemination to Europe [231]. Recent research indicates that ginger has anti-cancer, anticoagulant, anti-inflammatory, and analgesic properties [232]. Maintaining normal blood glucose levels is dependent on the amount of insulin present in the blood. Ginger increases serum insulin levels, enhances insulin sensitivity, and decreases blood glucose levels during fasting [233]. Table 4 provides a concise summary of the phytochemical properties, origin, bioactive components, structure, and effects of various herbal medications. The phytochemical properties of the herbs are also listed, allowing researchers to determine which herb should be used to treat a particular medical

**Table 4:-** Effects of Herbal Drugs, their phytoconstituents, source, chemical structure and their Pharmacological activity listed below in table

Sr No		Synony ms	Source (Part Use)	Bioactive Components	Structures	Animal models		Refere nce
1.	Acacia arabica (AA)	Babool, Unani Tibbi	Roots,	Phenols, alkaloids, flavonoi ds, Tannins, Saponin s	OH ,		Antidiabetic, antihyper lipidaemic, antioxidants	[234] [235]
2.	Aegle marmelosa (AM)	Bael Tree	Fruit pulp. leaf	Alkaloids, carbohy drates, glycosides, flavonoids, tannins, Coumarin, sterol, triterpen es		Alloxan induced diabetic rats	Antioxida nts, antibacteri al, antifungal, antiprolife rative against human tumor cell line K562, analgesic, anti- inflammat ory, anti- arthritis, hepatoprot ective, cytoprotec tive, antidiarrh oeal	[236]

3.		Bulbour plant	Dried bulb	Querciti n-1, cycloalli n-2, s- methyl- L- cysteine -		induced diabetic Rattus norvegicu s	hypolipida emic	[238]
4.	Aloe ver a (AV)	Aloe barbede nsis	leaves	Phenolic compou nd, flavanoids	O=N+ OH	STZ induced	Hypolipid aemia, improve ce llular integrity, C VD, kidney failure	[239]
5.	Terminalia arjuna (TA)	Arjuna Tree	Stem, Bark	Arjunic acid, Termini c acid, Glycosi des, Tannins , Saponin s, flavones			Antioxida nt status in liver and kidneys, Lipid lowering	
6.	Cinnamonu m zeylanicum (CZ)		Inner bark	Coumarin	O H	Streptozo cin	Antioxida nts, lipid- lowering, Blood glucose lowering	

7.	Coriandrum sativum (CS)	Chinese parsley cilantro	Fresh leaves	Phenolic compou nds	HO, $CH_3$ $CH_2$ $H_3C$ $CH_3$	Alloxan induced diabetes in experime ntal animals	Antidiabet ic, hepatoprot ective activity, Antiperoxi dative, hypolipida emic	
8.	Datura stramonium (DS)	Jimson weed, Thorn apple	Seed extract	Flavanoids, anthraq uinone glycosid es, saponin s, Terpeno ids, tannins		STZ induced diabetes in mice	Antioxida nt potential, hypoglyce mic effect	
9.	Eugenia jambolana [EJ]	Syzygiu m jambola na, Indian blackber ry, Jamun	Seed extract, Fruit pulp	-		STZ induced diabetic male albino rats	Antihyper glycaemic activity, antioxidan t	[244]
10.	Trigonella foenumgraec um L (TFG)	Fenugre ek plant		Flavano ids, stilbene s glycosid es	R <sub>1</sub> O OH OOR <sub>2</sub>	diet-fed	Hypolipid aemic, antidiabeti cs	[245]

11.		tree,	Leaves, fruits	s-allyl cysteine sulphoxi de, l- chiro- inositol		Antidiabet ics, anxiolytic s, antioxidan ts	[246]
12.	Gymnema sylvestris R.Br. (GS)	Periploc aof woods, Gurmar	leaves	Gymne mic acid	STZ		[247]
13.	Hagenia abyssinica (HA)	African redwood		Gymne mic acid	STZ induced	antioxidan ts	[248]
14.	Momordica charantia L (MC)		Fresh fruit juice	momordicin, Charanti n, chorine, cryptox anthin, cucurbit in	monohydr ate	effects	[249]

15.		Ocimum tenuiflor um, Holy basil or tulsi	leaves	Cirsilineol, cirsimar tin, isothym osin	OH O O O O O O O O O O O O O O O O O O O	STZ induced diabetes in Wistar rats		[250]
16.	Piper longum (PL)			Piperine		STZ induced	renal	[251]
17.	Zingiber officinale (ZO)	Amomu m	Fresh & dried rhizom e	Gingerol		STZ induced type-1	Lowers glucose & lipids levels, antitumor, antimicrob ial	

# **Current Ongoing Clinical Trials**

Ahead the FDA approves a novel drug or medical device for use by the general public [253], it is subjected to extensive testing in clinical trials on patients. Clinical trials are crucial to the development of novel diabetes and other disease treatments [254]. There have been very few clinical trials reported for the treatment of diabetes mellitus [255]. This could be attributed to a lack of complete understanding of the disease's underlying mechanism, which prevent the patients from receiving complete relief [256].

As this metabolic disorder rapidly progressing among the urban diabetic populations therefore maximum therapeutic alternatives approximately 60 have been approved by the FDA [257]. From 2015 through 2020, 375 clinical studies including about 100 antidiabetic drugs are registered. Ten of these treatments already have commercial authorization from regulatory bodies in other countries and might possibly apply for FDA approval [258]. One-fourth of these drugs are in phase III studies. Additionally clinical trials on 100 drugs are carried on beyond insulin, metformin and other drug combinations like sitagliptins etc [259]. DPP-4 inhiboitors and SGLT-2 inhibitors have gained a special importance in pharmaceutical market which are less expensive alternatives in developing

countries [260]. Since the FDA authorized human insulin (Humulin) in 1982, 59 different antidiabetic medications have been approved [261]. The authorized medications include 23 original pharmacological combinations of two or more antidiabetic medicines and 36 novel molecular entities as monotherapies [262]. A major clinical development is in process for diabetes management which is in phase III trials.

Clinical trial is a very important step to ensure the safety and effective use of a medicine among human beings. It needs the completion and analysis of safe, effective results from phase I through phase III by USFDA regulatory agencies [263]. US Food and Drug Administration's authorize 100 new medicines every year [264].

Some of the completed or ongoing trials on antidiabetic herbs (DPP-4 Inhibitors) are listed below in table no. 05 with their complete profile.

Table 05: Clinical	Trial	of Herbal	Drugs	in	various	Phases	for	Diabetes	mellitus	Metabolic
Disorder										

Sr. No	Herbal Drug	Sampl e size	Duration of Study	Purpose	Phase	Status	Design	Study Year	Refere nce
• 1.	American ginseng	770	$\geq$ 30 days	Treatment	Phase 2	Completed	R	3 July, 2013	[265]
2.	Cinnamon zeylenicum	25	12 weeks	Treatment	Phase 3	completed	PC, R	21 Feb, 2016	[266]
3.	Trigonella foenum graecum	154	90 days	Treatment	Phase 1	completed	R	11 oct, 2016	[267]
4.	Allium sativum L.	110	5 weeks	Treatment	Phase 4	completed	R	7 July, 2020	[268]
5.	Bitter melon	95	10 weeks	Treatment	Phase 2	completed	R	26 jan, 2015	[269]
6.	Azadirachta indica	80	12 weeks	Treatment	Phase 1	completed	R, DB, PC	17 Nov, 2020	[270]
7.	Onion cepa	28	8 weeks	Treatment	Phase 1	completed	R	3 june, 2016	[271]
8.	Plantago psyllium	30	40 days	Treatment	Phase 2	completed	R	2 may, 2017	[272]
9.	Siberian ginseng	15	5 days	Treatment	Phase 2	completed	R	Sep, 2018	[273]
10.	Tinospora cardifolia	60	15 days	Treatment	Phase 3	completed	R	1 Feb, 2016	[274]
11.	Urtica diocia	46	3 months	Treatment	Phase 1	completed	R, DB, PC	2013	[275]
12.	Nigella sativa	57	3 months	Treatment	Phase 1	completed	PC	23 Feb, 2015	[276]
13.	Coccinia grandis	79	3 months	Treatment	Phase 3	Recruiting	R, DB, PC	Jan, 2021	[277]

14.	Cinnamonu	997	3 months	Treatment	Phase 2	completed	R	Juna, 2020	[278]
	m verum &								
	Cassia								
	aromaticum								
15.	Silybum	60	90 days	Treatment	Phase 4	completed	R, DB,	6 Feb, 2017	[279]
	marianum						PC		
16.	Carthamus	67	2 weeks	Treatment	Phase 3	Recruiting	R, DB,	10 Jan, 2022	[280]
	tinctorius						PC		
17.	Berberis	75	8 weeks	Treatment	Phase 3	Recruiting	R	5 July, 2021	[281]
	vulgaris								
18.	Momordica	23	8 weeks	Treatment	Phase 2	completed	R, DB,	15 Oct, 2014	[282]
	charantia						PC		
19.	Ilex	973	16 weeks	Treatment	Phase 2	completed	R	June, 2019	[283]
	paraguariens								
	is								
20.	Boswellia	71	12 weeks	Treatment	Phase 4	completed	R	March, 2014	[284]
	serrata								
21.	Postulaca	32	12 weeks	Treatment	Phase 3	completed	R, DB,	8 Feb, 2016	[285]
	oleracea						PC		

**Acroniums**: CO; Cross-Over, DB; Double-Blind, PC; Placebo-controlled, NR; Non-Randomized, R; Randomized Requirements for a Drug Delivery System.

There are lots of methodologies through which a drug is able to produce its maximum efficacy on target tissue with in an optimum concentration range [286]. Below or above optimum concentration will not be able to produce its therapeutic effect at all. Different drug carriers like soluble polymers, micro particles, synthetic polymers, microcapsules, micelles, lipoproteins and liposomes are used to target the drug loaded system at the site of interest [287]. Multiple mechanisms, including diffusion through the carrier matrix, diffusion through the carrier wall, and a combined diffusion or attrition process, are involved in the drug's release [288]. Currently, medication targeting systems are being developed to decrease medication loss and degradation, prevent adverse side effects, and increase drug bioavailability [289].

Using nanoscale materials as diagnostic instruments or to transport therapeutic medications to specific targeted regions in a controlled manner is the field of nano delivery systems [290], which is still immature but expanding rapidly. By delivering targeted and site-specific medications, nanotechnology has a number of benefits for the treatment of chronic human diseases. Recently, numerous noteworthy applications of nanomedicine have emerged [291][292]. Microparticles are defined as having dimensions measured in micrometers (typically 1 to 1000 m) and possessing distinct structural characteristics, such as low bioavailability, poor water solubility, and lack of blood-brain barrier (BBB) permeability. Recent advances in the understanding of drug pharmacokinetic and pharmacodynamic behavior have led to the creation of novel drug delivery systems, which are new systems that adopt a more logical approach to the design of the ideal drug

#### delivery system [293] [294].

In controlled release drug delivery systems, the drug is released at a predetermined rate based on the intended therapeutic concentration and the pharmacokinetic properties of the drug. Nanoparticles of polymer transport insulin [295]. These biodegradable polymers containing a polymeric insulin matrix are encapsulated in a membrane containing nanoholes that permit glucose oxidation [296]. From implantable electronic devices to single polymer chains, drug delivery systems must be biocompatible with both body processes and the substance to be delivered. DDS modify the biodistribution and pharmacokinetics of the associated drug: that is, the time-dependent proportion of the administered dose in the various organs of the body [297].

Niosomes have been used as a system to increase the bioavailability of drugs with limited aqueous solubility and to make active drugs available at the site of action for a prolonged period of time [298]. When sodium deoxycholate was used as a surfactant, it was discovered that insulin-loaded niosomes were stable in the presence of proteolytic enzymes of the gastrointestinal tract [299]. In order to make these systems available on a global scale, it is necessary to consider their safety profile, pharmacology, environmental effects during formulation, and prospective effects on health care [300].

Moreover, the developed formulations must be comprehensively optimised in a variety of animal species in order to reduce the occurrences of clinical failure in a variety of human subjects [301]. The development of robust glucose-sensitive nanoparticles as well as integrated glucose-sensing and insulin-delivering nanoformulations [302] could be investigated as an additional therapeutic possibility. These ongoing advancements in nanotechnology offer optimism for the development of an effective anti-diabetic treatment in the near future [303]. In addition to reducing the need for repeated administration to combat noncompliance, novel drug delivery methods contribute to an increase in therapeutic value by reducing toxicity and increasing bioavailability [304].

#### Phytonanotherapy in the Management of Diabetes

The planet is home to a wide variety of native medicinal flora. The World Health Organization (WHO) prioritizes healthcare that is environmentally benign, non-hazardous, and cost-effective, such as the use of medicinal plants to treat a variety of diseases [305]. Unique to phytonanotherapy, the synergistic properties of plant and metal NPs offer clinically bioequivalent effects to many synthetic medications with minimal adverse effects [306]. This could provide an effective alternative treatment for chronic diseases, circumventing the disadvantages of synthetic monotherapy and permitting medicinal plant therapy to coexist with existing synthetic treatments [307]. The pharmaceutical formulation consists of nanoparticles of solid lipids. Particulate systems, such as nanoparticles, are utilized as a physical method for modifying and enhancing the pharmacokinetic and pharmacodynamic properties of diverse drug molecules [308]. It is possible to create nanoparticles through chemical, physical, and biological processes [309]. Although the chemical method of synthesis requires little time to produce a large number of nanoparticles, capping agents are required to stabilize the nanoparticles' size [310].

Phytotherapeutics require a systematic strategy to providing the components in a sustained way in order to maximise patient compliance and minimise repetitive dosing [311]. This can be done by creating innovative drug delivery systems (NDDSs) for herbal components [312]. NDDSs not only lessen the need for repeated administration to overcome noncompliance, but they also

contribute to therapeutic efficacy [313]. Via reducing toxicity and improving bioavailability Nanoparticles (NPs) are loaded with therapeutic substances for delivery to target cells. In addition, metal nanoparticles appear to be less toxic than mineral compounds and serve multiple functions within the body [314].

**Metallic nanoparticles:** Nanotechnology is creating nanoparticles of copious metal oxides like gold, silver, zinc, magnesium, titanium, etc., with sizes ranging from 1 to 100 nm. Because of their non-invasiveness and site-specificity, the use of metallic nanoparticles is becoming the most appealing and promising phytonanotherapy option [315]. Metallic nanoparticles have significant potential for improving diabetes care because they allow for the oraldelivery of insulin to a specific site, increasing its bioavailability and pharmacological efficacy [316]. A metallic nanoparticle's optical properties are primarily determined by its surface plasmon resonance, where plasma on refers to the collective oscillation of free electrons within the metallic nanoparticle. Physical, chemical, and biological methods are used to produce nanoparticles [317] [318].

#### Nanoparticles of zinc oxide

Anti-diabetic, antibacterial, anticancer, antifungal, drug delivery, and anti-inflammatory characteristics are among the many biological uses for ZnO NPs. Zinc is necessary for insulin production, secretion, and storage and is crucial for insulin structure [319].

# Magnesium (Mg)

Magnesium (Mg) is an essential ion in glucose regulation. Mg is also implicated in several enzymes involved in phosphorylation and glucose metabolism, and it may potentially play a role in insulin production [320].

#### Cerium oxide nanoparticles

In the periodic table, the lanthanide series contains numerous rare earth elements, including Ce. CeO2 nanoparticles have demonstrated promise as a potential treatment for oxidative diseases, surpassing several existing treatments for brain injury [321].

# **Copper ions**

Copper is a transition metal that participates in numerous biological reactions. Cu NPs possess exceptional antioxidative properties and inhibit alpha-amylase and alpha-glucosidase in animals.

Moreover, Cu NPs substantially reduced diabetics' functional cardiovascular abnormalities. These NPs may increase nitric oxide bioavailability and reduce oxidative stress in the vascular endothelium [322].

#### Selenium nanoparticles (NPs)

Selenium is a trace element found in nearly all plant life. A selenium deficiency in the body has been linked to numerous diseases, including diabetes. Se NPs' antioxidant properties are also less hazardous than selenium itself [323]. Due to their ability to scavenge various peroxides, protect lipids and cellular macromolecules from oxidative membrane damage, and increase glutathione peroxidase and thioredoxine reductase levels, Se nanoparticles possess antioxidant properties [324].

#### Silver nanoparticles

In numerous medical, food storage, textile coating, and environmental applications, AgNPs have been utilized as anti-bacterial agents. AgNPs have been utilized as antibacterial agents in a variety of applications spanning from the disinfection of medical devices and household appliances to water treatment [325]. AgNPs exhibited potent antibacterial activity against Escherichia coli.

Synthesis of silver nanoparticles and their applications [326] [327].

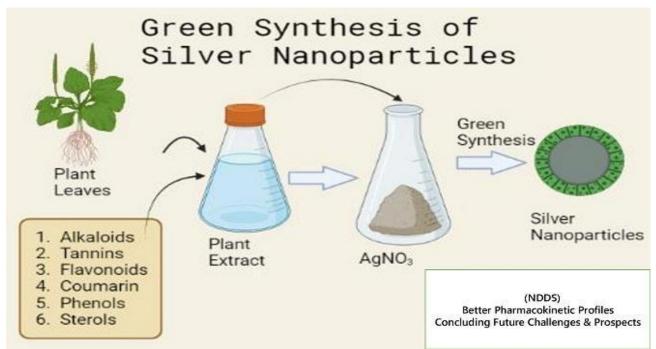


Figure -7: Emergence of Silver Nanoparticles using AgNO<sub>3</sub> as reducing Agent.

Silver Nanoparticles of herbal extract utilize various AgNO3 concentrations; the samples' visual appearance changed shortly after incorporation of the plant extract, indicating that a reduction reaction occurred. Initially, the reacting mixture is a faintly yellowish liquid that eventually turns dark reddish brown. As the reaction continued, the solutions changed color, becoming orange, red, and brown. This is a significant indicator of the origin or development of silver nanoparticles. In addition, the formulation was confirmed using UV spectroscopy at 420 nm.

**Pharmacokinetic parameters for herbal drugs & their nanoparticles:-** In numerous in vitro and in vivo models, the pharmacokinetic and pharmacodynamic properties of anti-diabetic medications were evaluated using innovative drug delivery systems (NDDS) [328]. The NDDS technology, which is simple, fast, cost-effective, and environmentally favorable, improves preclinical output. It provides enhanced in vitro drug release, enhanced drug efficacy, enhanced glucose uptake, enhanced insulin sensitivity, enhanced collagen fiber density, enhanced glucokinase enzyme activity, and enhanced GLUT-4 activity [329].

Moreover, it increases the levels of TNF, IL-4, IL-6, IL-10, GK, IRA, GLUT-2, SH, SOD, VEGF, and TGF while decreasing the levels of IL-1b, -glucosidase, PKLR, MDA, MAPK, ROS, and renal fibrosis. Compared to other dosage formulations, it improves therapeutic efficacy, solubility, stability, and bioavailability [330]. Additionally, it reduces body weight and glucose levels. Phytonanotherapy reduces hyperglycemia-induced apoptosis, oxidative stress, DNA damage, and impaired glucose uptake in L6 skeletal muscle cells more effectively and with a lower dose than other treatments [331] [332]. The many herbal drugs In Table 6, which summarizes nanoparticle formulations, various NDDS and in vitro results are briefly discussed together with phytoconstituents of the plant.

Sr.					References
No.	Plants Used	Phytoconstituents	NDDS	Outcome	
1.	Ocimum basilicum, Ocimum Pharmacokinetic parameters for herbal drugs & their nanoparticles sanctum	Alpha-pinene, beta- myrcene, 4- Hexen- 1- ol acetate	AgNPs	Extract AgNPs shows 79- 89% inhibition for in vitro α- glycosidase whereas 49- 66% inhibition ofExtract	[333]
2.	Aloe vera	Aloin	Aloe loaded chitosan NP	Aloe loaded NP shows maximum in vitro drug release during cancer treatment to improve effectiveness and reduce adverse effects.	[334]
3.	Taxus brevifolia	Taxanes	Liposomes	Inclusion into liposomes led to decreased tissue toxicity & increased efficacy of drug	[335]
4.	Leonotis leonurus	Labdane	Nanostructured lipidCarrier	↑insulin sensitivity ↑Glucose uptake	[336]
5.	Dendrocalamu s hamilton	Cellulose, lignin	Nanobiocomposite (cellulose nanocrystals and AgNPs)	$\begin{array}{ccc} \downarrow TNF\alpha & \downarrow IL6 \\ \uparrow PDGF & \uparrow FGF \\ \uparrow VEGF & \uparrow \\ TGF\beta & \uparrow Fiber \\ density & of \end{array}$	[337]

Table No.- 06 Pharmacokinetic Parameters of Novel Herbal Drug Nanoparticles Formulation

				collagen	
				↓blood glucose	
6.	Pouteria sapota	Allene carotenoid	Synthesized green	↑serum insulin	[338]
	-		Ag nanoparticle	↓ alpha amylase	
				†glucose uptake	
				↓FBS ↓HbA1c	
7.	Stevia	Steviol glycoside	Chitosan	• •	[339]
	rebaudiana		nanoparticles	↑ CAT ↑GSH	
			1	↑ consumption	
			PLGA	of glucose	
8.	Syzygium	Delphinidin,	nano-encapsulated	Ũ	[340]
	cumini	cyaniding	1	†GLUT4 ↓	
				plasma glucose,	
				↓NF-κB ↓	
				iNOS	
9.	Syzygium	Delphinidin,	Green synthesized	↓cell size ↓lipi	d [341]
	cumini	cyaniding	Ag nanoparticle	peroxidation	
				↓blood glucose	
10.	Eysenhardtia	Dihydroxychalcone	Green synthesized	↓insulin	[342]
11.	-	Linarin			[343]
	Polystachya		Ag nanoparticle	secretion ↓TC	
			Green synthesized	↓blood glucose	
12.	Musa	Myrcene	Ag nanoparticle	↓HbA1c ↑insulin	[344]
	paradisiaca			↑glycogen	
		Oxalic acid,	Green synthesized	↓Blood glucose	
13.	Cassia fistula	oxyanthroquinone	Au nanoparticle	↓HbA1c↓ LDL-C	[345]
			-	↑ HDL-C	
	Gymnema		Green synthesized	↓ TNFα ↓ILe	5
14.	sylvestre	Gymnemic acid	Au nanoparticle	↓CRP ↓HbA1c	[346]
			_	LDL-C ↑HDL-C	
			Green synthesized	↓ blood glucose	
15.	Sambucus	Anthocyanin,	Au nanoparticle	↓MDA	[347]
	nigra	cyanin		$\downarrow COX2$	
16.	Marsilea	Trans-Farnesol	Green synthesized	↑ glucos	e [348]
	quadrifolia		Au nanoparticle	utilization	
				↓Blood glucose	
17.	Chamaecostus	α-tocopherol	Green synthesized	↑body weigh	t [349]
	cuspidatus		Au nanoparticle	↓super oxide anion	1
				↓lipid peroxidation	
	Stevia			↓blood sugar	s
18.	rebaudiana	stevioside	TiO2 nanomaterial	↑insulin ↓HbA1	c [350]

19.	Moringa	Gallic acid	Green synthesized	↓ α-amylase	[351]
17.	oleifera		ZnO nanoparticle	$\downarrow \alpha$ -glucosidase	[001]
	Tamarindus		Green synthesized	$\downarrow \alpha$ -amylase	
20.	indica	Tartaric acid	ZnO nanoparticle	$\downarrow \alpha$ -glucosidase.	[352]
	Hibiscus	Allo-hydroxy citric	Green synthesized	$\downarrow$ TNF- $\alpha$ $\downarrow$ IL-1b	
21.	subdariffa	acid lactone	ZnO nanoparticle	$\downarrow IL- 6 \uparrow IL-4 \uparrow IL-10$	[353]
22.	Copaifera sp.	Copaiba oil	Nano encapsulated	↑SH ↑Gpx ↑SOD ↑Nfr2 ↓RV hypertrophy	[354]
23.	Bambusa bambos	Cellulose, lignin	Nanobiocomposite (cellulose nanocrystals and AgNPs)	↓TNFα ↓IL6 ↑PDGF ↑FGF ↑VEGF ↑ TGFβ ↑Fiber density of collagen	[355]
24.	Argyreia nervosa	Aryl esters, scopoletin	Green synthesized Ag nanoparticle	$\downarrow \alpha$ -amylase $\downarrow \alpha$ -glucosidase	[356]
25.	Cinnamomum litseifolium	Cinnamaldehyde, cinnamic acid	Nanoemulsion	↓α-amylase ↓α- glucosidase	[357]
26.	Costus speciosus	costunolide	PLGA nano-encapsulated	glucose in the blood, insulin (I&II), and GLUT4 GLUT2	[358]
27.	Ficus religiosa	lanosterol	Solid lipid nanoparticle	↓ blood glucose ↑insulin level	[359]
28.	Momordica charantia	charantin	Nanoemulsion	$\uparrow$ SOD, $\uparrow$ GPx, $\downarrow$ MDA	[360]
29.	Zingiber officinale	gingerols	Green synthesized AgNPs	↓ blood sugar	[361]
30.	Silybum marianum	silibinin	Synthesized green ZnO nanoparticle	↓FBS, TC, TAG ↑insulin, HDL-C	[362]
31.	The Nasturtium officinale	myristicin	Green nanoparticles of ZnO synthesized	↓FBS, TC, TAG, insulin, HDL-C, and TAG	[363]
32.	Punica granatum	Oleanolic acid	Green synthesized Au nanoparticle	↓MAPK/NF- κB/STAT3 ↓ RAGENOX- 4/p47phox ↓ROS ↑Nrf2 ↓renal fibrosis	[364]

Section A-Research paper

	r				[]
33.	Morus alba	prenylflavin	Synthesized green Ag nanoparticle	↓deterioration in retinal cell layer ↓ Aluminum and carbohydrates	[365]
34.	Smilax glabra	smiglabrone	Green synthesized Au nanoparticle	↓TNFα and IL-β ↓leptin ↑adiponectin ↓body mass index and blood glucose levels	[366]
35.	Curcuma longa	curcumin	Curcumin-self- nanophospholipid dispersions	enhance therapeutic effectiveness, solubility, stability, and bioavailability	[367]
36.	Vaccinium macrocarpon	Resveratrol	lipid nanocarriers	enhanced bioavailability, targeted targeting, advanced therapeutic effectiveness, and improved patient compliance	[368]
37.	Matricaria chamomilla	Naringenin	Self- nanoemulsified naringenin	enhance medication absorption, release, and oral bioavailability	[369]
38.	Scutellaria baicalensis	Baicallin	Baicalin- entrapped nanoliposome	increased biodistribution and oral bioavailability	[370]
39.	Reseda luteola	Luteolin	Solid lipid nanoparticles containing luteolin	Enhance solubility, biological half- life and bioavailability	[371]
40.	Mangifera indica Rheum	Mangiferin	Self-assembled phospholipidic nanomicelles containing mangiferin Solid lipid nanoparticles containing luteolin Emodin-loaded	improve the biopharmaceutical attributes improve oral	[372]
			Emoum-ioadeu	mprove oral	

41.	officinale	Emodin	nanoemulsion	bioavailability	[373]
			Rosmarinic acid-	high heat stability,	
42.	Rosemarinus	Rosmarinic acid	chitosan	in vitro release, and	[374]
	offficinalis		nanoparticles	effective	
				antioxidant action	
				improve berberine's	
	European		Berberine-	oral bioavailability,	
43.	barberry	Berberine	assembled	stability, and anti-	[375]
			solid lipid	diabetic activity	
			nanoparticles	over	
				the free medication	
			Nanostructured	increased small	
44.	Centella	Asiatic acid	lipid carriers	intestine	[376]
	asiatica		containing	penetration and	
			PEGylated asiatic	transport	
			Acid	capacity	
			Chitosan-gum	increased	
45.	Glycyrrhiza	Glycyrrhizin	arabic nanoparticles	bioavailability and	[377]
	glabra		assembled with	long-term release	
			glycyrrhizin		
			enriched with -	Improved	
46.	Prunus	α-Eleostearic Acid	Eleostearic acid	formulation for the	[378]
	mahaleb		nanoemulsion	treatment of	
				diabetes and	
	<u>a</u> . 11 .			toxicosis.	
47	Scutellaria	G ( 11 )	Scutellarin-loaded	increased	[270]
47.	barbata	Scutellarin	amphiphilic	bioavailability and	[379]
				long-term release	
				Improvements have	
	Cilebras			been made to water	
40	Silybum	C'il-manin	Silymarin-loaded	solubility,	[290]
48.	marianum	Silymarin	Soluplus-TPGS	biological stability,	[380]
			nanomicelles	P-gp inhibition,	
				gastrointestinal	
				absorption, and	
			Chitosan	cellular uptake. Better antioxidant	
49	Camelia	Catechins			[381]
47	sinesis	Catecinins	nanoparticles with	capacity than	[301]
	51110515		catechin grafting	native catechins	
				ameliorate hyperglycemia-	
				induced apoptosis,	
				muuccu apopiosis,	

Section A-Research paper

50.	Rasp berries	Pelargonidin	Pelargonidin- PLGA nanoparticles	oxidative stress, DNA damage, and glucose consumption impairment more effectively and at a lower dosage than natural pelargonidin	[382]
51.	Nigella sativa	Thymoquinone	Thymoquinone- loaded gum rosin nanocapsules	cure hyperglycemia, dyslipidemia, and haemoglobin glycosylation more effectively at a considerably lower dose	[383]
52.	Bamboo Tree	Ferulic acid	Nanoparticles of ferulic acid- chitosan	Excellent therapeutic efficacy in lowering blood glucose levels and	[384]
53.	Terminalia arjuna	Arjunic acid	Au NPs	Economically possible may be used in controlling human and agricultural pathogens.	[385]
54.	Curcuma longa	curcumin	sodiumalginate- gum arabic nanoparticles	Good therapeutic potential for the prevention and treatment of solid malignancies, including hepatic, breast, cervical, and cutaneous cancers.	[386]

# **Conclusions and Prospectives**

DPP-4 inhibitors are superior GLP-1 receptor-based therapies; they reduce gastric emptying and food intake, increase glucose-dependent insulin secretion, and suppress glucagon post-meal release. They increase endogenous GLP-1 concentrations without increasing the risk of hypoglycemia,

which stimulates insulin release and inhibits glucagon secretion [388]. They have a range of 0.5 to 1.0% HbA1c effectiveness potential, and their safety profile is very good. DPP-4 inhibitors are generally safe, successful in T2DM patients, and we anticipate that these will be able to assist patients in achieving their glycemic objectives in a supportive therapy environment [389] [390].

The therapeutic molecule made from herbal plants is effective in treating diabetes and other vascular problems. For its therapeutic purpose, these phytochemicals are more biocompatible than other synthetic compounds [391]. However, they have super biopharmaceutical and pharmacokinetic characteristics, which limit their therapeutic relevance. To solve this issue and increase treatment efficacy and patient compliance, a number of pharmacological initiatives have been considered. Nanotechnology has been identified as the most effective method to address this issue [392].

With the breadth of numerous applications, nanoscience and nanotechnology have advanced quickly in recent years. Metallic nanoparticles have a number of advantages in clinical practise, including superior biocompatibility and stability, low operating and capital costs, and minimal environmental impact [393]. The development of metallic nanoparticles with antioxidant properties appears to be an especially promising therapeutic option because it may allow for highly targeted or localised therapy [394]. Site-directed sustained delivery with enhanced curative potential, patient compliance, and the absence of undesirable toxic side effects and hypersensitivity reactions are crucial characteristics of herbal drugs [395]. The unique advantages of nano antioxidants for clinical application from the fact are that they can be made larger than the cutoff size for kidney filtration (10 nm), extending the circulation period in comparison to small molecules [396] [397]. There is a need to demonstrate increased bioavailability, target specificity, and the utilisation of various aspects of nanotechnology. Not only synthesised herbal-mediated metal nanoparticles biodegradable, biocompatible, and non-toxic properties, but they also have a greater capacity to penetrate biological cell membranes and act more rapidly, resulting in increased bioavailability as an alternative herbal treatment for diabetes [398] [399].

### Summary

In summary, nanoparticle formulations of plant-derived herbal medicinal compounds offer improved pharmacological and biopharmaceutical profiles, which remove all related barriers to provide the greatest therapeutic outcome [400]. This study focuses on the preclinically and clinically successful herbal anti-diabetic nanoscale formulations that use mechanisms, such as DPP-4 inhibition, to manage diabetes and its associated problems. In order to entirely eradicate this ailment, researchers are advised to continue their study on this subject.

ABBREVIATIONS DPP-4: Dipeptidyl peptidase-4

TGF: Transforming Growth Factor

CeO: Celenium Oxide NDDS: Novel Drug Delivery System ZnO: Zinc Oxide AgNP: Nanoparticles **ROS:** Reactive oxygen species FBS:Fasting blood sugar HDL:High density lipoprotein SOD:Superoxide Dismutase **GP:** Glycoproteins MDA:MassDrug Administeration GLUT: Glucose Transporter PDGF:Platelet DerivedGrowth Factor FGF:Fibroblast Growth Factor VEGF:VascularEndothelialGrowth Factor LDL:Low Density Lipoprotein **TC:Total Cholesterol** TAG:Tri Acyl Glycerol DDS:Drug Delivery System **BBB:Blood Brain Barrier** THC:Tetra Hydro Cannabinol **CBD**:Cannabidiol EGCG:EpiGalloCatechin-3-gallate PBT2:Polybutylene Terephthalate SD-809: Tablet for Chorea disease FDA:Food & Drug Administeration STZ: Streptozocin WHO: World Health Organization CPE: Petroleum Extract of Ether Inos: Inducible nitricoxide synthase NrF-2: Nuclear factor erythroid 2-related factor 2 FBS: Fasting Blood Sugar CAT: Anti-immunotoxin cell TGF: Transforming growth factor NDDS: Novel Drug Delivery System COX-2: Carboxylase enzyme **STAT:** Instantly

PLGA:Poly lactic co-glycollic acid DNA:Deoxy ribonucleic acid TNF-α:Tumour Necrosis Factor IL:Inter Leukin MAPK: Mitogen activated Protein Kinase NF: Nuclear Factor kappa B HOMA-IR: NMR: Homeostatic Model Assessment of insulin resistance PPAR-y: Peroxisome proliferator activated receptor URT1: Upper respiratory track -1 TNFR2: Tumor necrosis factor receptor TG:Tri Glyceride TC:Total Cholesterol LL:Lipoprotein-lipase CVD:Cardio Vascular Disease HBA1c: Haemoglobin A1c DM:Diabetes mellitus MDA: Mass Drug administeration **CAT:**Computerized Axial Tomography **CPE:**Continuing Education Pharmacy Dinucleotide NAD:Nicotinamide Adenine **GLP:Glucagon Like Peptidase** PABA:Para Amino Benzoic Acid **DHF:Design History File** T2DM:Type-2 Diabetes mellitus LPO:Lipid peroxidation

### References

[1] Mukhtar, Y., Galalain, A., & Yunusa, U. (2020). A modern overview on diabetes mellitus: a chronic endocrine disorder. *European Journal of Biology*, 5(2), 1-14. [2] Mohamed, J., Nafizah, A. N., Zariyantey, A. H., & Budin, S. (2016). Mechanisms of diabetes-induced liver damage: the role of oxidative stress and inflammation. *Sultan qaboos university*  medical journal, 16(2), e132.

[3] Natarajan, E. (2020). *Plasma Fibinogen Level in Newly Detected Type 2 Diabetes Mellitus Patients in a Tertiary Care Centre* (Doctoral dissertation, Madras Medical College, Chennai).

[4] Verma, S., Gupta, M., Popli, H., & Aggarwal, G. (2018). Diabetes mellitus treatment using herbal drugs. *International Journal of Phytomedicine*, *10*(1), 1-10.

[5] Fisher, M. C., Alastruey-Izquierdo, A., Berman, J., Bicanic, T., Bignell, E. M., Bowyer, P., ... & Verweij, P. E. (2022). Tackling the emerging threat of antifungal resistance to human health. *Nature reviews microbiology*, 20(9), 557-571.

[6] Chin, Y. X., Chen, X., Cao, W. X., Sharifuddin, Y., Green, B. D., Lim, P. E., ... & Tang, Q. J. (2020). Characterization of seaweed hypoglycemic property with integration of virtual screening for identification of bioactive compounds. *Journal of functional foods*, *64*, 103656.

[7] Xin, Z., Kupczyk, E., Schmitt-Kopplin, P., & Mueller, C. (2022). Current and future approaches for in vitro hit discovery in diabetes mellitus. *Drug Discovery Today*.

[8] Patel, B. D., & Ghate, M. D. (2014). Recent approaches to medicinal chemistry and therapeutic potential of dipeptidyl peptidase-4 (DPP-4) inhibitors. *European journal of medicinal chemistry*, *74*, 574-605.

[9] Muntner, P., Davis, B. R., Cushman, W. C., Bangalore, S., Calhoun, D. A., Pressel, S. L., ...& Rahman, M. (2014). Treatment-resistant hypertension and the incidence of cardiovascular disease and end-stage renal disease: results from the Antihypertensive and Lipid-Lowering Treatment to Prevent Heart Attack Trial (ALLHAT). *Hypertension*, *64*(5), 1012-1021.

[10] Parveen, K., Banse, V., & Ledwani, L. (2016, April). Green synthesis of nanoparticles: Their advantages and disadvantages. In *AIP conference proceedings* (Vol. 1724, No. 1). AIP Publishing.

[11] Ozougwu, J. C., Obimba, K. C., Belonwu, C. D., & Unakalamba, C. B. (2013). The pathogenesis and pathophysiology of type 1 and type 2 diabetes mellitus. *J Physiol Pathophysiol*, 4(4), 46-57.

[12] Rohli, K. E., Boyer, C. K., Blom, S. E., & Stephens, S. B. (2022). Nutrient regulation of pancreatic islet  $\beta$ -cell secretory capacity and insulin production. *Biomolecules*, *12*(2), 335.

[13] Tchetina, E. V., Markova, G. A., & Sharapova, E. P. (2020). Insulin resistance in osteoarthritis: similar mechanisms to type 2 diabetes mellitus. *Journal of nutrition and metabolism*, 2020.

[14] Akash, M. S. H., Rehman, K., & Chen, S. (2013). Role of inflammatory mechanisms in pathogenesis of type 2 diabetes mellitus. *Journal of cellular biochemistry*, *114*(3), 525-531.

[15] Chun, P. (2021). Histone deacetylase inhibitors in medical therapeutics. In *Medical Epigenetics* (pp. 597-632). Academic Press.

[16] Qi, H., Casalena, G., Shi, S., Yu, L., Ebefors, K., Sun, Y., ... & Daehn, I. (2017). Glomerular endothelial mitochondrial dysfunction is essential and characteristic of diabetic kidney disease susceptibility. *Diabetes*, *66*(3), 763-778.

[17] Asmat, U., Abad, K., & Ismail, K. (2016). Diabetes mellitus and oxidative stress—A concise review. *Saudi pharmaceutical journal*, 24(5), 547-553.

[18] Aldubayan, M. A., Ahmed, A. S., Emara, A. M., Ahmed, A. A., & Elgharabawy, R. M. (2020). Sinapic acid attenuates cardiovascular disorders in rats by modulating reactive oxygen species and angiotensin receptor expression. *Oxidative Medicine and Cellular Longevity*, 2020.

[19] Wang, M., Liu, Y., Liang, Y., Naruse, K., & Takahashi, K. (2021). Systematic understanding 2165

of pathophysiological mechanisms of oxidative stress-related conditions—diabetes mellitus, cardiovascular diseases, and ischemia–reperfusion injury. *Frontiers in cardiovascular medicine*, *8*, 649785.

[20] Kesarwala, A. H., Krishna, M. C., & Mitchell, J. B. (2016). Oxidative stress in oral diseases. *Oral diseases*, 22(1), 9-18.

[21] Johny, J. P., Plank, M. J., & David, T. (2017). Importance of altered levels of SERCA, IP3R, and RyR in vascular smooth muscle cell. *Biophysical journal*, *112*(2), 265-287.

[22] Lv, W., Wang, X., Xu, Q., & Lu, W. (2020). Mechanisms and characteristics of sulfonylureas and glinides. *Current Topics in Medicinal Chemistry*, 20(1), 37-56.

[23] Garnham, J. O. (2018). *Mechanisms of exercise intolerance in chronic heart failure and type 2 diabetes mellitus* (Doctoral dissertation, University of Leeds).

[24] Satman, I., Omer, B., Tutuncu, Y., Kalaca, S., Gedik, S., Dinccag, N., ... & Tuomilehto, J. (2013). Twelve-year trends in the prevalence and risk factors of diabetes and prediabetes in Turkish adults. *European journal of epidemiology*, 28, 169-180.

[25] Kelishadi, R., & Poursafa, P. (2014). A review on the genetic, environmental, and lifestyle aspects of the early-life origins of cardiovascular disease. *Current problems in pediatric and adolescent health care*, 44(3), 54-72.

[26] Su, Z., Zou, Z., Hay, S. I., Liu, Y., Li, S., Chen, H., ... & Zhang, H. (2022). Global, regional, and national time trends in mortality for congenital heart disease, 1990–2019: An age-period-cohort analysis for the Global Burden of Disease 2019 study. *EClinicalMedicine*, 43.

[27] De Gaetano, M., McEvoy, C., Andrews, D., Cacace, A., Hunter, J., Brennan, E., & Godson, C. (2018). Specialized pro-resolving lipid mediators: modulation of diabetes-associated cardio-, reno-, and retino-vascular complications. *Frontiers in pharmacology*, *9*, 1488.

[28] Kropp, M., Golubnitschaja, O., Mazurakova, A., Koklesova, L., Sargheini, N., Vo, T. T. K. S., ... & Thumann, G. (2023). Diabetic retinopathy as the leading cause of blindness and early predictor of cascading complications—Risks and mitigation. *EPMA Journal*, *14*(1), 21-42.

[29] Makrilakis, K. (2019). The role of DPP-4 inhibitors in the treatment algorithm of type 2 diabetes mellitus: when to select, what to expect. *International journal of environmental research and public health*, *16*(15), 2720.

[30] Brunton, S. (2014). GLP-1 receptor agonists vs. DPP-4 inhibitors for type 2 diabetes: is one approach more successful or preferable than the other?. *International journal of clinical practice*, 68(5), 557-567.

[31] Dougherty, J. A., Guirguis, E., & Thornby, K. A. (2021). A systematic review of newer antidiabetic agents in the treatment of nonalcoholic fatty liver disease. *Annals of Pharmacotherapy*, 55(1), 65-79.

[32] Ojha, A., Ojha, U., Mohammed, R., Chandrashekar, A., & Ojha, H. (2019). Current perspective on the role of insulin and glucagon in the pathogenesis and treatment of type 2 diabetes mellitus. *Clinical pharmacology: advances and applications*, 57-65.

[33] Vesa, C. M., Popa, L., Popa, A. R., Rus, M., Zaha, A. A., Bungau, S., ... & Zaha, D. C. (2020). Current data regarding the relationship between type 2 diabetes mellitus and cardiovascular risk factors. *Diagnostics*, *10*(5), 314.

[34] Tiwari, P., Mishra, B. N., & Sangwan, N. S. (2014). Phytochemical and pharmacological properties of Gymnema sylvestre: an important medicinal plant. *BioMed research* 

international, 2014.

[35] Yuan, H., Ma, Q., Cui, H., Liu, G., Zhao, X., Li, W., & Piao, G. (2017). How can synergism of traditional medicines benefit from network pharmacology?. *Molecules*, 22(7), 1135.

[36] Baker, C., Retzik-Stahr, C., Singh, V., Plomondon, R., Anderson, V., & Rasouli, N. (2021). Should metformin remain the first-line therapy for treatment of type 2 diabetes?. *Therapeutic Advances in Endocrinology and Metabolism*, *12*, 2042018820980225.

[37] Alqahtani, A., Hamid, K., Kam, A., Wong, K. H., Abdelhak, Z., Razmovski-Naumovski, V., ... & Li, G. Q. (2013). The pentacyclic triterpenoids in herbal medicines and their pharmacological activities in diabetes and diabetic complications. *Current Medicinal Chemistry*, 20(7), 908-931.

[38] Simos, Y. V., Spyrou, K., Patila, M., Karouta, N., Stamatis, H., Gournis, D., ... & Peschos, D. (2021). Trends of nanotechnology in type 2 diabetes mellitus treatment. *Asian journal of pharmaceutical sciences*, *16*(1), 62-76.

[39] World Health Organization. (2017). *Human resources for medical devices, the role of biomedical engineers*. World Health Organization.

[40] Thomford, N. E., Senthebane, D. A., Rowe, A., Munro, D., Seele, P., Maroyi, A., & Dzobo, K. (2018). Natural products for drug discovery in the 21st century: innovations for novel drug discovery. *International journal of molecular sciences*, *19*(6), 1578.

[41] Strimbu, K., & Tavel, J. A. (2010). What are biomarkers?. Current opinion in HIV and AIDS, 5(6), 463–466. https://doi.org/10.1097/COH.0b013e32833ed177.

[42] Gromov, M. S., Rogacheva, S. M., Barulina, M. A., Reshetnikov, A. A., Prokhozhev, D. A., & Fomina, A. Y. (2021). Analysis of Some Physiological and Biochemical Indices in Patients with Covid-19 Pneumonia Using Mathematical Methods. Journal of evolutionary biochemistry and physiology, 57(6), 1394–1407. https://doi.org/10.1134/S0022093021060181.

[43] Wiersinga WJ, Rhodes A, Cheng AC, Peacock SJ, Prescott HC. Pathophysiology, Transmission, Diagnosis, and Treatment of Coronavirus Disease 2019 (COVID-19): A Review. JAMA. 2020;324(8):782–793. doi:10.1001/jama.2020.12839.

[44] Popko K, Gorska E, Stelmaszczyk-Emmel A, Plywaczewski R, Stoklosa A, Gorecka D, Pyrzak B, Demkow U. Proinflammatory cytokines II-6 and TNF-α and the development of inflammation in obese subjects. Eur J Med Res. 2010 Nov 4;15 Suppl 2(Suppl 2):120-2. doi: 10.1186/2047-783x-15-s2-120. PMID: 21147638; PMCID: PMC4360270.

[45] Gohda, T., Niewczas, M. A., Ficociello, L. H., Walker, W. H., Skupien, J., Rosetti, F., Cullere, X., Johnson, A. C., Crabtree, G., Smiles, A. M., Mayadas, T. N., Warram, J. H., & Krolewski, A. S. (2012). Circulating TNF receptors 1 and 2 predict stage 3 CKD in type 1 diabetes. Journal of the American Society of Nephrology : JASN, 23(3), 516–524. https://doi.org/10.1681/ASN.2011060628.

[46] Ghai, V., Baxter, D., Wu, X., Kim, T. K., Kuusisto, J., Laakso, M., ... & Wang, K. (2019). Circulating RNAs as predictive markers for the progression of type 2 diabetes. Journal of cellular and molecular medicine, 23(4), 2753-2768.

[47] Stevens, L. A., Coresh, J., Schmid, C. H., Feldman, H. I., Froissart, M., Kusek, J., ... & Levey, A.

S. (2008). Estimating GFR using serum cystatin C alone and in combination with serum creatinine:

a pooled analysis of 3,418 individuals with CKD. American journal of kidney diseases, 51(3), 395-406.

[48] Currie, G., McKay, G., & Delles, C. (2014). Biomarkers in diabetic nephropathy: Present and future. World journal of Diabetes, 5(6), 763.

[49] Narita, T., Hosoba, M., Kakei, M., & Ito, S. (2006). Increased urinary excretions of immunoglobulin g, ceruloplasmin, and transferrin predict development of microalbuminuria in patients with type 2 diabetes. Diabetes care, 29(1), 142-144.

[50] Kado, S., Aoki, A., Wada, S., Katayama, Y., Kugai, N., Yoshizawa, N., & Nagata, N. (1996). Urinary type IV collagen as a marker for early diabetic nephropathy. Diabetes research and clinical practice, 31(1-3), 103-108.

[51] Banu, N., Hara, H., Okamura, M., Egusa, G., & Yamakido, M. (1995). Urinary excretion of type IV collagen and laminin in the evaluation of nephropathy in NIDDM: comparison with urinary albumin and markers of tubular dysfunction and/or damage. Diabetes Research and Clinical Practice, 29(1), 57-67.

[52] Tuttle, K. R. (2005). Linking metabolism and immunology: diabetic nephropathy is an inflammatory disease. Journal of the American Society of Nephrology, 16(6), 1537-1538.

[53] Fournier, T. (2000). Medjoubi-N N., Porquet D. Alpha-1-Acid Glycoprotein. Biochim. Biophys. Acta, 1482(1-2), 157-171.

[54] Schlatzer, D., Maahs, D. M., Chance, M. R., Dazard, J. E., Li, X., Hazlett, F., ... & Snell-Bergeon,

J. K. (2012). Novel urinary protein biomarkers predicting the development of microalbuminuria and renal function decline in type 1 diabetes. Diabetes care, 35(3), 549-555.

[55] Yanagawa T, Taniguchi A, Fukushima M, Nakai Y, Nagasaka S, Ohgushi M, Matsumoto K, Kuroe A, Ohya M, Seino Y. Leptin, triglycerides, and interleukin 6 are independently associated with C-reactive protein in Japanese type 2 diabetic patients. Diabetes Res Clin Pract. 2007 Jan;75(1):2-6. doi: 10.1016/j.diabres.2006.04.019. Epub 2006 Jun 9. PMID: 16764962.

[56] Feingold KR. Introduction to Lipids and Lipoproteins. [Updated 2021 Jan 19]. In: Feingold KR, Anawalt B, Blackman MR, et al., editors. Endotext [Internet]. South Dartmouth (MA): MDText.com, Inc.; 2000-. Available from: https://www.ncbi.nlm.nih.gov/books/NBK305896/.

[57] Bajaj, J. S., Fan, S., Thacker, L. R., Fagan, A., Gavis, E., White, M. B., Heuman, D. M., Fuchs, M., & Fiehn, O. (2019). Serum and urinary metabolomics and outcomes in cirrhosis. PloS one, 14(9), e0223061. https://doi.org/10.1371/journal.pone.0223061

[58] Zhang, J. M., & An, J. (2007). Cytokines, inflammation, and pain. International anesthesiology clinics, 45(2), 27–37. https://doi.org/10.1097/AIA.0b013e318034194e.

[59] European Association For The Study Of The Liver. (2012). EASL clinical practice guidelines: Wilson's disease. Journal of hepatology, 56(3), 671-685.

[60] Deshmane, S. L., Kremlev, S., Amini, S., & Sawaya, B. E. (2009). Monocyte chemoattractant protein-1 (MCP-1): an overview. Journal of interferon & cytokine research : the official journal of the International Society for Interferon and Cytokine Research, 29(6), 313–326. https://doi.org/10.1089/jir.2008.0027.

[61] Singh, S., Anshita, D., & Ravichandiran, V. (2021). MCP-1: Function, regulation, and involvement in disease. International immunopharmacology, 101(Pt B), 107598. https://doi.org/10.1016/j.intimp.2021.107598.

[62] Larsson A, Palm M, Hansson LO, Basu S, Axelsson O. Reference values for alpha1-acid glycoprotein, alpha1-antitrypsin, albumin, haptoglobin, C-reactive protein, IgA, IgG and IgM during pregnancy. Acta Obstet Gynecol Scand. 2008;87(10):1084-8. doi: 10.1080/00016340802428146. PMID: 18792844.

[63] Fritz H. Proteinase inhibitors in severe inflammatory processes (septic shock and experimental endotoxaemia): biochemical, pathophysiological and therapeutic aspects. Ciba Found Symp. 1979;(75):351-79. doi: 10.1002/9780470720585.ch20. PMID: 399895.

[64] Tino Hochepied; Franklin G Berger; Heinz Baumann; Claude Libert (2003).  $\alpha$ 1-Acid glycoprotein: an acute phase protein with inflammatory and immunomodulating properties. 14(1), 0–34. doi:10.1016/s1359-6101(02)00054-0.

[65] Hu Y, Hosseini A, Kauwe JS, Gross J, Cairns NJ, Goate AM, Fagan AM, Townsend RR, Holtzman DM. Identification and validation of novel CSF biomarkers for early stages of Alzheimer's disease. Proteomics Clin Appl. 2007 Nov;1(11):1373-84. doi: 10.1002/prca.200600999. Epub 2007 Oct 16. PMID: 21136637.

[66] Rahman, M. Mahafuzur; Zetterberg, Henrik; Lendel, Christofer; Härd, Torleif (2015). Binding of Human Proteins to Amyloid- $\beta$  Protofibrils. ACS Chemical Biology, 10(3), 766–774. doi:10.1021/cb5008663.

[67] C. I. Jhala; U. V. Shah; T. K. Shah; B. K. Naik; J. D. Dafda (1998). A study of serum lipid profile part-1: Establishment of normal reference values of serum lipid levels in healthy vegetarian population of Gujarat., 13(1), 1–7. doi:10.1007/bf02873435.

[68] Nain, M., Gupta, A., Malhotra, S. et al. High-density lipoproteins may play a crucial role in COVID-19. Virol J 19, 135 (2022). https://doi.org/10.1186/s12985-022-01865-4.

[69] Sidorkiewicz, I., Niemira, M., Maliszewska, K., Erol, A., Bielska, A., Szalkowska, A., Adamska-Patruno, E., Szczerbinski, L., Gorska, M., & Kretowski, A. (2020). Circulating miRNAs as a Predictive Biomarker of the Progression from Prediabetes to Diabetes: Outcomes of a 5-Year Prospective Observational Study. Journal of clinical medicine, 9(7), 2184. https://doi.org/10.3390/jcm9072184.

[70] Ghai, V., Baxter, D., Wu, X., Kim, T. K., Kuusisto, J., Laakso, M., Connolly, T., Li, Y., Andrade-Gordon, P., & Wang, K. (2019). Circulating RNAs as predictive markers for the progression of type 2 diabetes. Journal of cellular and molecular medicine, 23(4), 2753–2768. https://doi.org/10.1111/jcmm.14182.

[71] Pordzik, J., Jakubik, D., Jarosz-Popek, J. et al. Significance of circulating microRNAs in diabetes mellitus type 2 and platelet reactivity: bioinformatic analysis and review. Cardiovasc Diabetol 18, 113 (2019). https://doi.org/10.1186/s12933-019-0918-x.

[72] Ghoreishi, E., Shahrokhi, S.Z., Kazerouni, F. et al. Circulating miR-148b-3p and miR-27a-3p

can be potential biomarkers for diagnosis of pre-diabetes and type 2 diabetes: integrating experimental and in-silico approaches. BMC Endocr Disord 22, 207 (2022). https://doi.org/10.1186/s12902-022-01120-5.

[73] Ghai, V., Baxter, D., Wu, X., Kim, T. K., Kuusisto, J., Laakso, M., Connolly, T., Li, Y., Andrade-Gordon, P., & Wang, K. (2019). Circulating RNAs as predictive markers for the progression of type 2 diabetes. Journal of cellular and molecular medicine, 23(4), 2753–2768. https://doi.org/10.1111/jcmm.14182.

[74] Bartnikas T. B. (2012). Known and potential roles of transferrin in iron biology. Biometals : an international journal on the role of metal ions in biology, biochemistry, and medicine, 25(4), 677–686. https://doi.org/10.1007/s10534-012-9520-3.

[75] Sánchez-Hidalgo, J. J., Suárez-Cuenca, J. A., Lozano-Nuevo, J. J., García-López, V. H., Leal-Gutiérrez, M. G., León-Angel, S. A., Ramírez-Villa, M. L., Rodea-Rubio, M. E., González-Hernández, J. E., Canela-Mayoral, J. A., Murillo-Heredia, E., Vera-Gómez, E., Hernández-Patricio, A., Zamora-Alemán, C. R., Domínguez-Pérez, G. A., Gutiérrez-Buendia, J. A., & Mondragón-Terán, P. (2021). Urine transferrin as an early endothelial dysfunction marker in type 2 diabetic patients without nephropathy: a case control study. Diabetology & metabolic syndrome, 13(1), 128. https://doi.org/10.1186/s13098-021-00745-1.

[76] Wang, C., Li, C., Gong, W., & Lou, T. (2013). New urinary biomarkers for diabetic kidney disease. Biomarker research, 1(1), 9. https://doi.org/10.1186/2050-7771-1-9.

[77] Fiseha, T. Urinary biomarkers for early diabetic nephropathy in type 2 diabetic patients. Biomark Res 3, 16 (2015). https://doi.org/10.1186/s40364-015-0042-3.

[78] Swaminathan, S.M., Rao, I.R., Shenoy, S.V. et al. Novel biomarkers for prognosticating diabetic kidney disease progression. Int Urol Nephrol 55, 913–928 (2023). https://doi.org/10.1007/s11255-022-03354-7.

[79] Purohit, S., Sharma, A., Zhi, W., Bai, S., Hopkins, D., Steed, L., Bode, B., Anderson, S. W., Reed, J. C., Steed, R. D., & She, J. X. (2018). Proteins of TNF- $\alpha$  and IL6 Pathways Are Elevated in Serum of Type-1 Diabetes Patients with Microalbuminuria. Frontiers in immunology, 9, 154. https://doi.org/10.3389/fimmu.2018.00154.

[80] Araújo, L. S., Torquato, B. G. S., da Silva, C. A., Dos Reis Monteiro, M. L. G., Dos Santos Martins, A. L. M., da Silva, M. V., Dos Reis, M. A., & Machado, J. R. (2020). Renal expression of cytokines and chemokines in diabetic nephropathy. BMC nephrology, 21(1), 308. https://doi.org/10.1186/s12882-020-01960-0.

[81] Arau'jo LS, da Silva MV, da Silva CA, Borges MdF, Palhares HMdC, Rocha LP, et al. (2020) Analysis of serum inflammatory mediators in type 2 diabetic patients and their influence on renal function. PLoS ONE 15(3): e0229765. https:// doi.org/10.1371/journal.pone.0229765.

[82] Shashi Prabha Singh, Preeti Sharma, Pradeep Kumar, A.K. Mathur, Tapan Mahapatra. Role of cytokines IL-1, IL-6 and TNF- $\alpha$  in the pathogenesis of diabetic nephropathy. IAIM, 2019; 6(6): 141-150.

[83] Morran, M. P., Vonberg, A., Khadra, A., & Pietropaolo, M. (2015). Immunogenetics of type 1 diabetes mellitus. Molecular aspects of medicine, 42, 42–60. https://doi.org/10.1016/j.mam.2014.12.004.

[84] Noble, J. A., & Valdes, A. M. (2011). Genetics of the HLA region in the prediction of type 1 diabetes. Current diabetes reports, 11(6), 533–542. https://doi.org/10.1007/s11892-011-0223-x.

[85] Zajec A, Trebušak Podkrajšek K, Tesovnik T, Šket R, Čugalj Kern B, Jenko Bizjan B, Šmigoc Schweiger D, Battelino T, Kovač J. Pathogenesis of Type 1 Diabetes: Established Facts and New Insights. Genes. 2022; 13(4):706. https://doi.org/10.3390/genes13040706.

[86] Morran, M. P., Vonberg, A., Khadra, A., & Pietropaolo, M. (2015). Immunogenetics of type 1 diabetes mellitus. Molecular aspects of medicine, 42, 42–60. https://doi.org/10.1016/j.mam.2014.12.004.

[87] Yi, L., Swensen, A. C., & Qian, W. J. (2018). Serum biomarkers for diagnosis and prediction of type 1 diabetes. Translational research : the journal of laboratory and clinical medicine, 201, 13–25. https://doi.org/10.1016/j.trsl.2018.07.009.

[88] Shilpa Suneja;Sukanya Gangopadhyay;Vandana Saini;Rajni Dawar;Charanjeet Kaur; (2021). Emerging Diabetic Novel Biomarkers of the 21st Century . Annals of the National Academy of Medical Sciences (India), (), –. doi:10.1055/s-0041-1726613.

[89] Steinmetz, K.L., Spack, E.G. The basics of preclinical drug development for neurodegenerative disease indications. BMC Neurol 9 (Suppl 1), S2 (2009). https://doi.org/10.1186/1471-2377-9-S1-S2.

[90] Van Norman, G. A. (2019). Limitations of animal studies for predicting toxicity in clinical trials: is it time to rethink our current approach?. JACC: Basic to Translational Science, 4(7), 845-854.

[91] Bahadoran, Z., Mirmiran, P., Kashfi, K., & Ghasemi, A. (2020). Importance of systematic reviews and meta-analyses of animal studies: challenges for animal-to-human translation. Journal of the American Association for Laboratory Animal Science, 59(5), 469-477.

[92] Huang, W., Percie du Sert, N., Vollert, J., Rice, A.S.C. (2019). General Principles of Preclinical Study Design. In: Bespalov, A., Michel, M., Steckler, T. (eds) Good Research Practice in Non-Clinical Pharmacology and Biomedicine. Handbook of Experimental Pharmacology, vol 257. Springer, Cham. https://doi.org/10.1007/164\_2019\_277.

[93] Bundhun, Pravesh Kumar et al. "Adverse drug effects observed with vildagliptin versus pioglitazone or rosiglitazone in the treatment of patients with type 2 diabetes mellitus: a systematic review and meta-analysis of randomized controlled trials." BMC pharmacology & toxicology vol. 18,1 66. 23 Oct. 2017, doi:10.1186/s40360-017-0175-0.

[94] Liu, R., Zhang, P. Towards early detection of adverse drug reactions: combining pre-clinical drug structures and post-market safety reports. BMC Med Inform Decis Mak 19, 279 (2019). https://doi.org/10.1186/s12911-019-0999-1.

[95] Hegazy GA, Alnoury AM, Gad HG. The role of Acacia Arabica extract as an antidiabetic, antihyperlipidemic, and antioxidant in streptozotocin-induced diabetic rats.

[96] Jha, Deepak Kumar and Sharma, Pankaj Kumar and Haimed, Yousef Ahmed Saleh (2022) Anti-Type I Diabetic Activity of the Methanolic Extract of Aegle marmelos on Streptozotocin Induced Rat Model. Journal of Pharmaceutical Research International. pp. 1-14. ISSN 2456-9119.

[97] Galavi, A., Hosseinzadeh, H., & Razavi, B. M. (2021). The effects of Allium cepa L. (onion) and its active constituents on metabolic syndrome: A review. Iranian journal of basic medical sciences, 24(1), 3–16. https://doi.org/10.22038/ijbms.2020.46956.10843

[98] Sharma, N., Behl, T., Singh, S., Bansal, A., Singh, S. K., & Zahoor, I. (2021). Expatiating the therapeutic profile of garlic (Allium sativum): a bench to bedside approach. Biointerface Res. Appl. Chem, 11(6), 14225-14239.

[99] Amare, G. G., Meharie, B. G., & Belayneh, Y. M. (2020). Evaluation of Antidiabetic Activity of the Leaf Latex of Aloe pulcherrima Gilbert and Sebsebe (Aloaceae). Evidence-based complementary and alternative medicine : eCAM, 2020, 8899743. https://doi.org/10.1155/2020/8899743.

[100] Satyanarayana, K., Sravanthi, K., Shaker, I. A., & Ponnulakshmi, R. (2015). Molecular approach to identify antidiabetic potential of Azadirachta indica. Journal of Ayurveda and integrative medicine, 6(3), 165–174. https://doi.org/10.4103/0975-9476.157950.

[101] Iftikhar, A., Aslam, B., Iftikhar, M., Majeed, W., Batool, M., Zahoor, B., Amna, N., Gohar, H., & Latif, I. (2020). Effect of Caesalpinia bonduc Polyphenol Extract on Alloxan-Induced Diabetic Rats in Attenuating Hyperglycemia by Upregulating Insulin Secretion and Inhibiting JNK Signaling Pathway. Oxidative medicine and cellular longevity, 2020, 9020219.

https://doi.org/10.1155/2020/9020219.

[102] Quamri, M. A., Begum, S., Siddiqui, M. A., & Alam, M. A. (2017). Efficacy of Kanduri (Coccinia indica) in diabetes associated dyslipidemia-A randomized single blind standard controlled study. Age (Mean±SD), 43(7.845), 44-9.

[103] Ayyanar, M., Subash-Babu, P., & Ignacimuthu, S. (2013). Syzygium cumini (L.) Skeels., a novel therapeutic agent for diabetes: folk medicinal and pharmacological evidences. Complementary Therapies in Medicine, 21(3), 232-243.

[104] Ngo D-H, Ngo D-N, Vo TTN, Vo TS. Mechanism of Action of Mangifera indica Leaves for<br/>Anti-Diabetic Activity. Scientia Pharmaceutica. 2019; 87(2):13.<br/>https://doi.org/10.3390/scipharm87020013

[105] Banerjee, J., Chanda, R., & Samadder, A. (2019). Anti-diabetic activity of Momordica charantia or bitter melon: a review. Acta Scientific Pharmaceutical Sciences, 3, 24-30.

[106] Liu, Z., Gong, J., Huang, W., Lu, F., & Dong, H. (2021). The effect of Momordica charantia in the treatment of diabetes mellitus: A review. Evidence-Based Complementary and Alternative Medicine, 2021.

[107] Beidokhti, M. N., Andersen, M. V., Eid, H. M., Villavicencio, M. L. S., Staerk, D., Haddad, P. S., & Jńger, A. K. (2017). Investigation of antidiabetic potential of Phyllanthus niruri L. using assays for  $\alpha$ -glucosidase, muscle glucose transport, liver glucose production, and adipogenesis. Biochemical and biophysical research communications, 493(1), 869-874.

[108] Elavarasi, S., Revathi, G., Saravanan, K., & Averal, H. I. (2021). ANTIDIABETIC EFFECT OF MIXTURE OF CYATHEA NILGIRIENSIS (HOLTTUM) AND PTEROCARPUS MARSUPIUM ROXB. IN STREPTOZOTOCIN INDUCED DIABETIC RAT MODEL. International Journal of Pharmaceutical Sciences and Research, 12(4), 2147-57.

[109] Seyed Ali Hosseini, Khadijeh Hamzavi, Hoda Safarzadeh, Omidreza Salehi. (2023) Interactive effect of swimming training and fenugreek (Trigonella foenum graecum L.) extract on glycemic indices and lipid profile in diabetic rats. Archives of Physiology and Biochemistry 129:2, pages 349-353.

[110] Singh, C. S., Singh, A. K., Khandelwal, S., & Vishwkarma, R. (2013). Anti-Diabetic Activity of Ethanolic Extract of Tinospora Cordifolia Leaves. Int. J. of Drug Discovery & Herbal Research, 3(1), 601-604.

[111] Zahra Sadat Hussaini, Hakima Askndari, Kawsar Alami, Sayed Yousof Mousavi (2021). "Effect of Rheum Ribes and Urtica Dioica on type 2 diabetic rats", International Journal of Pharmaceutical and Phytopharmacological Research, 11(1), pp.63-69.

[112] Jaiswal, Y. S., Tatke, P. A., Gabhe, S. Y., & Vaidya, A. B. (2017). Antidiabetic activity of extracts of Anacardium occidentale Linn. leaves on n-streptozotocin diabetic rats. Journal of traditional and complementary medicine, 7(4), 421-427.

[113] Chewchinda, S., Leakaya, N., Sato, H., & Sato, V. H. (2021). Antidiabetic effects of Maclura cochinchinensis (Lour.) corner heartwood extract. Journal of Traditional and Complementary Medicine, 11(1), 68-74.

[114] Hu, X., Cheng, D., & Zhang, Z. (2016). Antidiabetic activity of Helicteres angustifolia root. Pharmaceutical biology, 54(6), 938-944.

[115] Zhang, Y., Hu, T., Zhou, H., Zhang, Y., Jin, G., & Yang, Y. (2016). Antidiabetic effect of polysaccharides from Pleurotus ostreatus in streptozotocin-induced diabetic rats. International journal of biological macromolecules, 83, 126-132.

[116] Oyedemi, S. O., Adewusi, E. A., Aiyegoro, O. A., & Akinpelu, D. A. (2011). Antidiabetic and haematological effect of aqueous extract of stem bark of Afzelia africana (Smith) on streptozotocin–induced diabetic Wistar rats. Asian Pacific journal of tropical biomedicine, 1(5), 353-358.

[117] Emordi, J. E., Agbaje, E. O., Oreagba, I. A., & Iribhogbe, O. I. (2016). Antidiabetic and hypolipidemic activities of hydroethanolic root extract of Uvaria chamae in streptozotocin induced diabetic albino rats. BMC complementary and Alternative Medicine, 16(1), 1-8.

[118] Abeywickrama, K. R. W., Ratnasooriya, W. D., & Amarakoon, A. M. T. (2011). Oral hypoglycaemic, antihyperglycaemic and antidiabetic activities of Sri Lankan Broken Orange Pekoe Fannings (BOPF) grade black tea (Camellia sinensis L.) in rats. Journal of ethnopharmacology, 135(2), 278-286.

[119] Wariyapperuma, W. N. M., Kannangara, S., Wijayasinghe, Y. S., Subramanium, S., & Jayawardena, B. (2020). In vitro anti-diabetic effects and phytochemical profiling of novel varieties of Cinnamomum zeylanicum (L.) extracts. PeerJ, 8, e10070.

[120] Nagaonkar, H. N., Dhawal, P. P., Barve, S. S., & Kakodkar, S. A. Evaluation of in vitro Anti-Diabetic Potential of Aster (Callistephus chinensis) Flower Waste Using Alpha-Amylase Inhibitory Assay and Glucose Uptake Assay.

[121] Patil, D. K., & Jain, A. P. (2019). In-vivo antidiabetic activity of methanolic extract of Corchorus olitorius for the management of type 2 diabetes. Journal of Pharmacognosy and Phytochemistry, 8(3), 3213-3218.

[122] Jamadagni, P. S., Pawar, S. D., Jamadagni, S. B., Chougule, S., Gaidhani, S. N., & Murthy, S. N. (2017). Review of Holarrhena antidysenterica (L.) Wall. ex A. DC.: Pharmacognostic, Pharmacological, and Toxicological Perspective. Pharmacognosy reviews, 11(22), 141–144. https://doi.org/10.4103/phrev.phrev\_31\_16.

[123] Nurnaeimah, Nik Mat, Nashriyah Suryati Mohd, Khamsah Badaluddin, Noor Afiza Yusoff, Nornasuha Sajili, Mohammad Hailmi Mahmud, Khairil Mohd Adnan, Ahmad Faris and Khandaker, Mohammad Moneruzzaman 2020. The Effects of Hydrogen Peroxide on Plant Growth, Mineral Accumulation, as Well as Biological and Chemical Properties of Ficus deltoidea. Agronomy, Vol. 10, Issue. 4, p. 599.

[124] Guex, C. G., Reginato, F. Z., de Jesus, P. R., Brondani, J. C., Lopes, G. H. H., & de Freitas Bauermann, L. (2019). Antidiabetic effects of Olea europaea L. leaves in diabetic rats induced by

high-fat diet and low-dose streptozotocin. Journal of ethnopharmacology, 235, 1-7.

[125] Chauhan, Samrat1; Gupta, Sumeet1,; Yasmin, Sabina2; Saini, Monika1. Antihyperglycemic and Antioxidant Potential of Plant Extract of Litchi chinensis and Glycine max. International Journal of Nutrition, Pharmacology, Neurological Diseases 11(3):p 225-233, Jul–Sep 2021. | DOI: 10.4103/ijnpnd\_ijnpnd\_13\_21.

[126] Cui, L., Liu, M., Chang, X., & Sun, K. (2016). The inhibiting effect of the Coptis chinensis polysaccharide on the type II diabetic mice. Biomedicine & Pharmacotherapy, 81, 111-119.

[127] Jayaprasad, B., Sharavanan, P. S., & Sivaraj, R. (2016). Antidiabetic effect of Chloroxylon swietenia bark extracts on streptozotocin induced diabetic rats. Beni-Suef University Journal of Basic and Applied Sciences, 5(1), 61-69.

[128] Zhang, Y., Feng, F., Chen, T., Li, Z., & Shen, Q. W. (2016). Antidiabetic and antihyperlipidemic activities of Forsythia suspensa (Thunb.) Vahl (fruit) in streptozotocin-induced diabetes mice. Journal of ethnopharmacology, 192, 256-263.

[129] Jamwal, A., & Kumar, S. (2019). Antidiabetic activity of isolated compound from Coccinia indica. Indian Journal of Pharmaceutical Education and Research, 53(1), 151-159.

[130] Povydysh MN, Titova MV, Ivkin DY, Krasnova MV, Vasilevskaya ER, Fedulova LV, Ivanov IM, Klushin AG, Popova EV, Nosov AM. The Hypoglycemic and Hypocholesterolemic Activity of Dioscorea deltoidea, Tribulus terrestris and Panax japonicus Cell Culture Biomass in Rats with High-Fat Diet-Induced Obesity. Nutrients. 2023; 15(3):656. https://doi.org/10.3390/nu15030656

[131] Ny, V., Houška, M., Pavela, R., & Tříska, J. (2021). Potential benefits of incorporating Astragalus membranaceus into the diet of people undergoing disease treatment: An overview. Journal of Functional Foods, 77, 104339.

[132] Yang HJ, Kim MJ, Kwon DY, Kim DS, Lee YH, Kim JE, Park S. Anti-Diabetic Activities of Gastrodia elata Blume Water Extracts Are Mediated Mainly by Potentiating Glucose-Stimulated Insulin Secretion and Increasing  $\beta$ -Cell Mass in Non-Obese Type 2 Diabetic Animals. Nutrients. 2016; 8(3):161. https://doi.org/10.3390/nu8030161.

[133] Ranasinghe, P., Galappaththy, P., Constantine, G. R., Jayawardena, R., Weeratunga, H. D., Premakumara, S., & Katulanda, P. (2017). Cinnamomum zeylanicum (Ceylon cinnamon) as a potential pharmaceutical agent for type-2 diabetes mellitus: study protocol for a randomized controlled trial. Trials, 18(1), 1-8.

[134] Alexander-Aguilera, A., Aguirre-Maldonado, I., Antolín, J. R., Toledo, L. N., Rodríguez, I. S., & Otero, M. G. S. (2019). Effect of Litchi chinensis on adipose and hepatic tissues in rats with obesity and non-alcoholic fatty liver disease (NAFLD). Journal of the Saudi Society of Agricultural Sciences, 18(3), 235-240.

[135] Hu, W., Yan, G., Ding, Q., Cai, J., Zhang, Z., Zhao, Z., ... & Zhu, Y. Z. (2022). Update of Indoles: Promising molecules for ameliorating metabolic diseases. Biomedicine & Pharmacotherapy, 150, 112957.

[136] Zhang, J. G., Liu, Q., Liu, Z. L., Li, L., & Yi, L. T. (2015). Antihyperglycemic activity of Anoectochilus roxburghii polysaccharose in diabetic mice induced by high-fat diet and streptozotocin. Journal of Ethnopharmacology, 164, 180-185.

[137] Abdullahi, Z., Magaji, Y., Vantsawa, P. A., Sheshe, S. M., & Alhaji, J. A. (2022). Anti-Diabetic Potential of Gymnema Sylvestre: In Vitro and in Silico Analysis. International Journal of Pharmaceutical and Bio Medical Science, 2(08), 233-248.

[138] Zhu, H., Wang, X., Pan, H., Dai, Y., Li, N., Wang, L., ... & Gong, F. (2016). The mechanism by which safflower yellow decreases body fat mass and improves insulin sensitivity in HFD-induced obese mice. Frontiers in Pharmacology, 7, 127.

[139] Xu B, Li Z, Zeng T, Zhan J, Wang S, Ho C-T, Li S. Bioactives of Momordica charantia as Potential Anti-Diabetic/Hypoglycemic Agents. Molecules. 2022; 27(7):2175. https://doi.org/10.3390/molecules27072175

[140] Chen W, Balan P, Popovich DG. Review of Ginseng Anti-Diabetic Studies. Molecules. 2019; 24(24):4501. https://doi.org/10.3390/molecules24244501

[141] Widowati, W., Liliana Wargasetia, T., Afifah, E., Mozef, T., Sari Widya Kusuma, H., Nufus, H., ... & Rizal, R. (2018). Antioxidant and antidiabetic potential of Curcuma longa and its compounds. Asian Journal of Agriculture and Biology, 6(2), 149-161.

[142] Pérez Gutiérrez, R. M., Muñiz-Ramirez, A., Garcia-Campoy, A. H., & Mota Flores, J. M. (2021). Evaluation of the Antidiabetic Potential of Extracts of Urtica dioica, Apium graveolens, and Zingiber officinale in Mice, Zebrafish, and Pancreatic β-Cell. Plants, 10(7), 1438.

[143] Sivalingam, G., & Sriram, N. (2013). Anti-diabetic activity of Ribes nigrum fruit extract in alloxan induced diabetic rats. International Journal of Pharmaceutical Sciences and Research, 4(3), 1196.

[144] Afzal, M., Kazmi, I., Kaur, R., Ahmad, A., Pravez, M., & Anwar, F. (2013). Comparison of protective and curative potential of Daucus carota root extract on renal ischemia reperfusion injury in rats. Pharmaceutical biology, 51(7), 856-862.

[145] Khan, A., Khan, I., Halim, S. A., Rehman, N. U., Karim, N., Ahmad, W., ... & Al-Harrasi, A. (2022). Anti-diabetic potential of  $\beta$ -boswellic acid and 11-keto- $\beta$ -boswellic acid: Mechanistic insights from computational and biochemical approaches. Biomedicine & Pharmacotherapy, 147, 112669.

[146] Okaiyeto, K., Kerebba, N., Rautenbach, F., Singh, S. K., Dua, K., & Oguntibeju, O. O. (2022). UPLC-ESI-QTOF-MS phenolic compounds identification and quantification from ethanolic extract of: In vitro antioxidant and antidiabetic potentials. Arabian Journal of Chemistry.

[147] Ramadan, B. K., Schaalan, M. F., & Tolba, A. M. (2017). Hypoglycemic and pancreatic protective effects of Portulaca oleracea extract in alloxan induced diabetic rats. BMC complementary and alternative medicine, 17, 1-10.

[148] Tang, Guozhi et al. "Design and synthesis of benzenesulfonamide derivatives as potent antiinfluenza hemagglutinin inhibitors." ACS medicinal chemistry letters vol. 2,8 603-7. 7 Jun. 2011, doi:10.1021/ml2000627.

[149] Salehi, Bahare et al. "Antidiabetic Potential of Medicinal Plants and Their Active Components." Biomolecules vol. 9,10 551. 30 Sep. 2019, doi:10.3390/biom9100551.

[150] Tran N, Pham B, Le L. Bioactive Compounds in Anti-Diabetic Plants: From Herbal Medicine to Modern Drug Discovery. Biology. 2020; 9(9):252. https://doi.org/10.3390/biology9090252.

[151] Oteng Mintah, S., Asafo-Agyei, T., Archer, M.-A., Atta-Adjei Junior, P., Boamah, D., Kumadoh, [108] Agyare, C. (2019). Medicinal Plants for Treatment of Prevalent Diseases. IntechOpen. doi: 10.5772/intechopen.82049.

[152] Modak M, Dixit P, Londhe J, Ghaskadbi S, Devasagayam TP. Indian herbs and herbal drugs used for the treatment of diabetes. J Clin Biochem Nutr. 2007 May;40(3):163-73. doi:

10.3164/jcbn.40.163. PMID: 18398493; PMCID: PMC2275761.

[153] L. M. Singh, Ankita Siddhanta, Ajay K. Singh, Shankar Prinja, Atul Sharma, Himanshu Sikka, Latashree Goswami, Potential Impact of the Insurance on Catastrophic Health Expenditures Among the Urban Poor Population in India, Journal of Health Management, 10.1177/09720634221088425, (097206342210884), (2022).

[154] Modak, M., Dixit, P., Londhe, J., Ghaskadbi, S., & Devasagayam, T. P. A. (2007). Indian herbs and herbal drugs used for the treatment of diabetes. Journal of clinical biochemistry and nutrition, 40(3), 163-173..

[155] Sofowora, Abayomi et al. "The role and place of medicinal plants in the strategies for disease prevention." African journal of traditional, complementary, and alternative medicines : AJTCAM vol. 10,5 210-29. 12 Aug. 2013, doi:10.4314/ajtcam.v10i5.2.

[156] Tran, Ngan et al. "Bioactive Compounds in Anti-Diabetic Plants: From Herbal Medicine to Modern Drug Discovery." Biology vol. 9,9 252. 28 Aug. 2020, doi:10.3390/biology9090252.

[157] Mak, Isabella Wy et al. "Lost in translation: animal models and clinical trials in cancer treatment." American journal of translational research vol. 6,2 114-8. 15 Jan. 2014.

[158] Alam, Safaet et al. "Antidiabetic Phytochemicals From Medicinal Plants: Prospective Candidates for New Drug Discovery and Development." Frontiers in endocrinology vol. 13 800714. 24 Feb. 2022, doi:10.3389/fendo.2022.800714.

[159] Roozbeh, Nasibeh, and Leili Darvish. "Acacia Nilotica: New Plant for Help in Pelvic Organ Prolapse." Journal of menopausal medicine vol. 22,3 (2016): 129-130. doi:10.6118/jmm.2016.22.3.129.

[160] Dias, Daniel A et al. "A historical overview of natural products in drug discovery." Metabolites vol. 2,2 303-36. 16 Apr. 2012, doi:10.3390/metabo2020303.

[161] Mukundi, Mwangi J (2015). Antidiabetic Effects of Aqueous Leaf Extracts of Acacia nilotica in Alloxan Induced Diabetic Mice. Journal of Diabetes & Metabolism, 6(7), –. doi:10.4172/2155-6156.1000568.

[162] Ghanghro, A. B., Ghanghro, I. H., Channa, M. J., Lanjwani, A. H., & Qureshi, S. (2015). Phytochemical Screening and Quantitative Biochemical Assessment of Acacia Nilotica (Flowers and Leaves). Sindh University Research Journal-SURJ (Science Series), 47(4).

[163] Hegazy GA, Alnoury AM, Gad HG. The role of Acacia Arabica extract as an antidiabetic, antihyperlipidemic, and antioxidant in streptozotocin-induced diabetic rats. Saudi Med J. 2013 Jul;34(7):727-33. PMID: 23860893.

[164] Pradeep Kumar Bhateja, Randhir Singh, "Antidiabetic Activity of Acacia tortilis (Forsk.) Hayne ssp. raddiana Polysaccharide on Streptozotocin-Nicotinamide Induced Diabetic Rats", BioMed Research International, vol. 2014, Article ID 572013, 9 pages, 2014. https://doi.org/10.1155/2014/572013.

[165] Sabu MC, Kuttan R. Antidiabetic activity of Aegle marmelos and its relationship with its antioxidant properties. Indian J Physiol Pharmacol. 2004 Jan;48(1):81-8. PMID: 15270373.

[166] Kumar, Vikas et al. "Umbelliferone  $\beta$ -D-galactopyranoside from Aegle marmelos (L.) corr. an ethnomedicinal plant with antidiabetic, antihyperlipidemic and antioxidative activity." BMC complementary and alternative medicine vol. 13 273. 20 Oct. 2013, doi:10.1186/1472-6882-13-273. [167] Bhatti R, Sharma S, Singh J, Ishar MP. Ameliorative effect of Aegle marmelos leaf extract on early stage alloxan-induced diabetic cardiomyopathy in rats. Pharm Biol. 2011 Nov;49(11):1137-

## 43. doi: 10.3109/13880209.2011.572077. PMID: 22014262.

[168] Tiwari, Brahm Kumar et al. "Efficacy of Composite Extract from Leaves and Fruits of Medicinal Plants Used in Traditional Diabetic Therapy against Oxidative Stress in Alloxan-Induced Diabetic Rats." ISRN pharmacology vol. 2014 608590. 4 Mar. 2014, doi:10.1155/2014/608590.

[169] Cherie Melaku, Bamlaku, and Gedefaw Getnet Amare. "Evaluation of Antidiabetic and Antioxidant Potential of Hydromethanolic Seed Extract of Datura stramonium Linn (Solanaceae)." Journal of experimental pharmacology vol. 12 181-189. 22 Jun. 2020, doi:10.2147/JEP.S258522.

[170] Yrga Adugna, Baye et al. "Evaluation of the Antidiabetic Activity of Hydromethanolic Roots Extracts of Rumex abyssinicus Jacq: (Polygonaceae) in Swiss Albino Mice." Evidence-based complementary and alternative medicine : eCAM vol. 2022 5193250. 30 Jun. 2022, doi:10.1155/2022/5193250.

[171] Sharma M, Dhaliwal I, Rana K, Delta AK, Kaushik P. Phytochemistry, Pharmacology, and Species—A Review. Antioxidants. Toxicology of Datura 2021: 10(8):1291. https://doi.org/10.3390/antiox10081291.

[172] Kouamé, N. G. M., Koffi, C., N'Zoué, K. S., Yao, N., Doukouré, B., & Kamagaté, M. (2019). Comparative antidiabetic activity of aqueous, ethanol, and methanol leaf extracts of Persea americana and their effectiveness in type 2 diabetic rats. Evidence-Based Complementary and Alternative Medicine, 2019.

[173] Al-Snafi, A. E. (2017). Medical importance of Datura fastuosa (syn: Datura metel) and Datura stramonium-A review. IOSR Journal of Pharmacy, 7(2), 43-58.

[174] Patel, D K et al. "An overview on antidiabetic medicinal plants having insulin mimetic Asian Pacific journal of tropical biomedicine vol. 2,4 (2012): property." 320-30. doi:10.1016/S2221-1691(12)60032-X.

[175] Nabi, Shaik Abdul et al. "Antidiabetic and antihyperlipidemic activity of Piper longum root aqueous extract in STZ induced diabetic rats." BMC complementary and alternative medicine vol. 13 37. 18 Feb. 2013, doi:10.1186/1472-6882-13-37.

[176] Nabi, S. A., Kasetti, R. B., Sirasanagandla, S., Tilak, T. K., Kumar, M. V. J., & Rao, C. A. (2013). Antidiabetic and antihyperlipidemic activity of Piper longum root aqueous extract in STZ induced diabetic rats. BMC complementary and alternative medicine, 13, 1-9.

[177] Abu-Odeh, Alaa M, and Wamidh H Talib. "Middle East Medicinal Plants in the Treatment of Diabetes: A Review." Molecules (Basel, Switzerland) vol. 26,3 742. 31 Jan. 2021, doi:10.3390/molecules26030742.

[178] Mahleyuddin, Nisa Najibah et al. "Coriandrum sativum L.: A Review on Ethnopharmacology, Phytochemistry, and Cardiovascular Benefits." Molecules (Basel, Switzerland) vol. 27,1 209. 30 Dec. 2021, doi:10.3390/molecules27010209.

[179] King, Aileen J F. "The use of animal models in diabetes research." British journal of pharmacology vol. 166,3 (2012): 877-94. doi:10.1111/j.1476-5381.2012.01911.x.

[180] Kajal, Anu, and Randhir Singh. "Coriandrum sativum seeds extract mitigate progression of diabetic nephropathy in experimental rats via AGEs inhibition." PloS one vol. 14,3 e0213147. 7 Mar. 2019, doi:10.1371/journal.pone.0213147.

[181] Kajal, A., & Singh, R. (2019). Coriandrum sativum seeds extract mitigate progression of diabetic nephropathy in experimental rats via AGEs inhibition. PloS one, 14(3), e0213147.

[182] Strugała P, Dzydzan O, Brodyak I, Kucharska AZ, Kuropka P, Liuta M, Kaleta-Kuratewicz

K, Przewodowska A, Michałowska D, Gabrielska J, Sybirna N. Antidiabetic and Antioxidative Potential of the Blue Congo Variety of Purple Potato Extract in Streptozotocin-Induced Diabetic Rats. Molecules. 2019; 24(17):3126. https://doi.org/10.3390/molecules24173126.

[183] Huang, Fang-Yan et al. "Dietary ginger as a traditional therapy for blood sugar control in patients with type 2 diabetes mellitus: A systematic review and meta-analysis." Medicine vol. 98,13 (2019): e15054. doi:10.1097/MD.00000000015054.

[184] Khandouzi, Nafiseh et al. "The effects of ginger on fasting blood sugar, hemoglobin a1c, apolipoprotein B, apolipoprotein a-I and malondialdehyde in type 2 diabetic patients." Iranian journal of pharmaceutical research : IJPR vol. 14,1 (2015): 131-40.

[185] Akhani SP, Vishwakarma SL, Goyal RK. Anti-diabetic activity of Zingiber officinale in streptozotocin-induced type I diabetic rats. J Pharm Pharmacol. 2004 Jan;56(1):101-5. doi: 10.1211/0022357022403. PMID: 14980006.

[186] Kojoma, M., Kurihara, K., Yamada, K., Sekita, S., Satake, M., & Iida, O. (2002). Genetic identification of cinnamon (Cinnamomum spp.) based on the trnL-trnF chloroplast DNA. Planta Medica, 68(01), 94-96.

[187] Rao, Pasupuleti Visweswara, and Siew Hua Gan. "Cinnamon: a multifaceted medicinal plant." Evidence-based complementary and alternative medicine : eCAM vol. 2014 (2014): 642942. doi:10.1155/2014/642942.

[188] Modak, Manisha et al. "Indian herbs and herbal drugs used for the treatment of diabetes." Journal of clinical biochemistry and nutrition vol. 40,3 (2007): 163-73. doi:10.3164/jcbn.40.163.

[189] Patel, D K et al. "An overview on antidiabetic medicinal plants having insulin mimetic property." Asian Pacific journal of tropical biomedicine vol. 2,4 (2012): 320-30. doi:10.1016/S2221-1691(12)60032-X.

[190] Soni, Priyanka et al. "Pharmacological properties of Datura stramonium L. as a potential medicinal tree: an overview." Asian Pacific journal of tropical biomedicine vol. 2,12 (2012): 1002-8. doi:10.1016/S2221-1691(13)60014-3.

[191] Chaudhry, Zunaira Z et al. "Streptozotocin is equally diabetogenic whether administered to fed or fasted mice." Laboratory animals vol. 47,4 (2013): 257-65. doi:10.1177/0023677213489548.

[192] Bai, Y. H., Shi, D. X., Lu, H. Y., Yang, K. B., Zhao, H. H., Lu, B. N., & Pang, Z. R. (2021). Hypoglycemic effects of Tibetan medicine Huidouba in STZ-induced diabetic mice and db/db mice. Chinese Herbal Medicines, 13(2), 202-209..

[193] Kifle, Z. D., Abdelwuhab, M., Melak, A. D., Meseret, T., & Adugna, M. (2022). Pharmacological evaluation of medicinal plants with antidiabetic activities in Ethiopia: A review. Metabolism open, 100174.

[194] Abdelgawad SM, Hetta MH, Ibrahim MA, Fawzy GA, El-Askary HI, Ross SA. Holistic Overview of the Phytoconstituents and Pharmacological Activities of Egyptian Riverhemp [Sesbania sesban (L.) Merr.]: A Review. Natural Product Communications. 2023;18(3). doi:10.1177/1934578X231160882.

[195] Jana, K., Bera, T. K., & Ghosh, D. (2015). Antidiabetic effects of Eugenia jambolana in the streptozotocin-induced diabetic male albino rat. Biomarkers and Genomic Medicine, 7(3), 116-124.

[196] Vora, A et al. "Eugenia jambolana extract reduces the systemic exposure of Sitagliptin and improves conditions associated with diabetes: A pharmacokinetic and a pharmacodynamic herbdrug interaction study." Journal of traditional and complementary medicine vol. 9,4 364-371. 3 Oct. 2018, doi:10.1016/j.jtcme.2018.10.001.

[197] Verma, Neeraj et al. "Antihyperglycemic and antihyperlipidemic activity of ethyl acetate fraction of Rhododendron arboreum Smith flowers in streptozotocin induced diabetic rats and its role in regulating carbohydrate metabolism." Asian Pacific journal of tropical biomedicine vol. 2,9 (2012): 696-701. doi:10.1016/S2221-1691(12)60212-3.

[198] Panda DK, Ghosh D, Bhat B, Talwar SK, Jaggi M, Mukherjee R. Diabetic therapeutic effects of ethyl acetate fraction from the roots of Musa paradisiaca and seeds of Eugenia jambolana in streptozotocin-induced male diabetic rats. Methods Find Exp Clin Pharmacol. 2009 Nov;31(9):571-84. doi: 10.1358/mf.2009.31.9.1435645. PMID: 20094640.

[199] Chatterjee, K., Ali, K. M., De, D., Panda, D. K., & Ghosh, D. (2012). Antidiabetic and antioxidative activity of ethyl acetate fraction of hydromethanolic extract of seed of Eugenia jambolana Linn through in-vivo and in-vitro study and its chromatographic purification. Free Radicals and Antioxidants, 2(1), 21-30.

[200] Shrestha, J. T. M., Shrestha, H., Prajapati, M., Karkee, A., & Maharjan, A. (2017). Adverse effects of oral hypoglycemic agents and adherence to them among patients with type 2 diabetes mellitus in Nepal. Journal of Lumbini Medical College, 5(1), 34-40.

[201] Rahman, M. M., Dhar, P. S., Anika, F., Ahmed, L., Islam, M. R., Sultana, N. A., ... & Rauf, A. (2022). Exploring the plant-derived bioactive substances as antidiabetic agent: an extensive review. Biomedicine & Pharmacotherapy, 152, 113217.

[202] Abdel-Barry JA, Abdel-Hassan IA, Al-Hakiem MH. Hypoglycaemic and antihyperglycaemic effects of Trigonella foenum-graecum leaf in normal and alloxan induced diabetic rats. J Ethnopharmacol. 1997 Nov;58(3):149-55. doi: 10.1016/s0378-8741(97)00101-3. PMID: 9421250.

[203] Gandhi, G Rajiv, and P Sasikumar. "Antidiabetic effect of Merremia emarginata Burm. F. in streptozotocin induced diabetic rats." Asian Pacific journal of tropical biomedicine vol. 2,4 (2012): 281-6. doi:10.1016/S2221-1691(12)60023-9.

[204] Rahman, M. M., Dhar, P. S., Anika, F., Ahmed, L., Islam, M. R., Sultana, N. A., ... & Rauf, A. (2022). Exploring the plant-derived bioactive substances as antidiabetic agent: an extensive review. Biomedicine & Pharmacotherapy, 152, 113217.

[205] Sikarwar, Mukesh S, and M B Patil. "Antidiabetic activity of Crateva nurvala stem bark extracts in alloxan-induced diabetic rats." Journal of pharmacy & bioallied sciences vol. 2,1 (2010): 18-21. doi:10.4103/0975-7406.62700.

[206] Joseph, B., & Raj, S. J. (2010). Phytopharmacological and phytochemical properties of three Ficus species-an overview. Int J Pharma Bio Sci, 1(4), 246-253.

[207] Nawaz, H., Waheed, R., & Nawaz, M. (2020). Phytochemical Composition, Antioxidant Potential, and Medicinal Significance of Ficus. IntechOpen. doi: 10.5772/intechopen.86562.

[208] Alimoradi, N., Firouzabadi, N., & Fatehi, R. (2021). Metformin and insulin-resistant related diseases: Emphasis on the role of microRNAs. Biomedicine & Pharmacotherapy, 139, 111662.

[209] Tiwari, Pragya et al. "Phytochemical and pharmacological properties of Gymnema sylvestre: an important medicinal plant." BioMed research international vol. 2014 (2014): 830285. doi:10.1155/2014/830285.

[210] Kanetkar, Parijat et al. "Gymnema sylvestre: A Memoir." Journal of clinical biochemistry and nutrition vol. 41,2 (2007): 77-81. doi:10.3164/jcbn.2007010.

[211] Oboh M, Govender L, Siwela M, Mkhwanazi BN. Anti-Diabetic Potential of Plant-Based

Pentacyclic Triterpene Derivatives: Progress Made to Improve Efficacy and Bioavailability. Molecules. 2021; 26(23):7243. https://doi.org/10.3390/molecules26237243.

[212] Khan, Farzana et al. "Comprehensive Review on Phytochemicals, Pharmacological and Clinical Potentials of Gymnema sylvestre." Frontiers in pharmacology vol. 10 1223. 29 Oct. 2019, doi:10.3389/fphar.2019.01223.

[213] Chen, Xiuqin et al. "Microbial Etiology and Prevention of Dental Caries: Exploiting Natural Products to Inhibit Cariogenic Biofilms." Pathogens (Basel, Switzerland) vol. 9,7 569. 14 Jul. 2020, doi:10.3390/pathogens9070569.

[214] Fan, Minxia et al. "Antioxidant and Anti-Proliferative Properties of Hagenia abyssinica Roots and Their Potentially Active Components." Antioxidants (Basel, Switzerland) vol. 9,2 143. 6 Feb. 2020, doi:10.3390/antiox9020143.

[215] Kifle, Zemene Demelash, and Yaschilal Muche Belayneh. "Antidiabetic and Antihyperlipidemic Effects of the Crude Hydromethanol Extract of Hagenia abyssinica (Rosaceae) Leaves in Streptozotocin-Induced Diabetic Mice." Diabetes, metabolic syndrome and obesity : targets and therapy vol. 13 4085-4094. 29 Oct. 2020, doi:10.2147/DMSO.S279475.

[216] Alimoradi, N., Firouzabadi, N., & Fatehi, R. (2021). Metformin and insulin-resistant related diseases: Emphasis on the role of microRNAs. Biomedicine & Pharmacotherapy, 139, 111662.

[217] Rotich, W. (2022). Botanical aspects, chemical overview, and pharmacological activities of 14 plants used to formulate a Kenyan Multi-Herbal Composition (CareVid<sup>TM</sup>). Scientific African, e01287.

[218] Richter E, Geetha T, Burnett D, Broderick TL, Babu JR. The Effects of Momordica charantia on Type 2 Diabetes Mellitus and Alzheimer's Disease. International Journal of Molecular Sciences. 2023; 24(5):4643. https://doi.org/10.3390/ijms24054643.

[219] Tan, S. P., Kha, T. C., Parks, S. E., & Roach, P. D. (2016). Bitter melon (Momordica charantia L.) bioactive composition and health benefits: A review. Food Reviews International, 32(2), 181-202.

[220] Tiwari, Pragya et al. "Phytochemical and pharmacological properties of Gymnema sylvestre: an important medicinal plant." BioMed research international vol. 2014 (2014): 830285. doi:10.1155/2014/830285.

[221] Keller, Amy C et al. "Saponins from the traditional medicinal plant Momordica charantia stimulate insulin secretion in vitro." Phytomedicine : international journal of phytotherapy and phytopharmacology vol. 19,1 (2011): 32-7. doi:10.1016/j.phymed.2011.06.019.

[222] Xu, Bilin et al. "Bioactives of Momordica charantia as Potential Anti-Diabetic/Hypoglycemic Agents." Molecules (Basel, Switzerland) vol. 27,7 2175. 28 Mar. 2022, doi:10.3390/molecules27072175.

[223] Alshatwi, Ali A, and P Subash-Babu. "Aloe-Emodin Protects RIN-5F (Pancreatic  $\beta$ -cell) Cell from Glucotoxicity via Regulation of Pro-Inflammatory Cytokine and Downregulation of Bax and Caspase 3." Biomolecules & therapeutics vol. 24,1 (2016): 49-56. doi:10.4062/biomolther.2015.056.

[224] Kooti, Wesam et al. "The role of medicinal plants in the treatment of diabetes: a systematic review." Electronic physician vol. 8,1 1832-42. 15 Jan. 2016, doi:10.19082/1832.

[225] Rambiritch, Virendra et al. "Glibenclamide in patients with poorly controlled type 2 diabetes: a 12-week, prospective, single-center, open-label, dose-escalation study." Clinical pharmacology :

advances and applications vol. 6 63-9. 4 Apr. 2014, doi:10.2147/CPAA.S54809

[226] Patel, D K et al. "Diabetes mellitus: an overview on its pharmacological aspects and reported medicinal plants having antidiabetic activity." Asian Pacific journal of tropical biomedicine vol. 2,5 (2012): 411-20. doi:10.1016/S2221-1691(12)60067-7

[227] Gebremeskel, Leake et al. "Evaluation of Antidiabetic Effect of Ethanolic Leaves Extract of Becium grandiflorum Lam. (Lamiaceae) in Streptozotocin-Induced Diabetic Mice." Diabetes, metabolic syndrome and obesity : targets and therapy vol. 13 1481-1489. 4 May. 2020, doi:10.2147/DMSO.S246996

[228] Hannan JM, Ali L, Khaleque J, Akhter M, Flatt PR, Abdel-Wahab YH. Aqueous extracts of husks of Plantago ovata reduce hyperglycaemia in type 1 and type 2 diabetes by inhibition of intestinal glucose absorption. Br J Nutr. 2006 Jul;96(1):131-7. doi: 10.1079/bjn20061819. PMID: 16870001.

[229] Sanni, Olakunle et al. "Anti-hyperglycemic and ameliorative effect of concentrated hot waterinfusion of Phragmanthera incana leaves on type 2 diabetes and indices of complications in diabetic rats." Journal of diabetes and metabolic disorders vol. 18,2 495-503. 14 Nov. 2019, doi:10.1007/s40200-019-00456-5.

[230] Atal, S., Agrawal, R. P., Vyas, S., Phadnis, P., & Rai, N. (2012). Evaluation of the effect of piperine per se on blood glucose level in alloxan-induced diabetic mice. Acta Pol Pharm, 69(5), 965-969.

[231] Idonije, B. O., Festus, O., & Oluba, O. M. (2011). Plasma glucose, creatinine and urea levels in type 2 diabetic patients attending a Nigerian teaching hospital. Research journal of medical sciences, 5(1), 1-3.

[232] Li, Wei et al. "Lycopene ameliorates renal function in rats with streptozotocin-induced diabetes." International journal of clinical and experimental pathology vol. 7,8 5008-15. 15 Jul. 2014.

[233] Winiarska, Agata et al. "Inflammation and Oxidative Stress in Diabetic Kidney Disease: The Targets for SGLT2 Inhibitors and GLP-1 Receptor Agonists." International journal of molecular sciences vol. 22,19 10822. 6 Oct. 2021, doi:10.3390/ijms221910822.

[234] Mao, Qian-Qian et al. "Bioactive Compounds and Bioactivities of Ginger (Zingiber officinale Roscoe)." Foods (Basel, Switzerland) vol. 8,6 185. 30 May. 2019, doi:10.3390/foods8060185.

[235] Bode AM, Dong Z. The Amazing and Mighty Ginger. In: Benzie IFF, Wachtel-Galor S, editors. Herbal Medicine: Biomolecular and Clinical Aspects. 2nd edition. Boca Raton (FL): CRC Press/Taylor & Francis; 2011. Chapter 7. Available from: https://www.ncbi.nlm.nih.gov/books/NBK92775/.

[236] Ali BH, Blunden G, Tanira MO, Nemmar A. Some phytochemical, pharmacological and toxicological properties of ginger (Zingiber officinale Roscoe): a review of recent research. Food Chem Toxicol. 2008 Feb;46(2):409-20. doi: 10.1016/j.fct.2007.09.085. Epub 2007 Sep 18. PMID: 17950516.

[237] Mohamad Hesam Shahrajabian, Wenli Sun & Qi Cheng (2019) Clinical aspects and health benefits of ginger (Zingiber officinale) in both traditional Chinese medicine and modern industry, Acta Agriculturae Scandinavica, Section B — Soil & Plant Science, 69:6, 546-556, DOI: 10.1080/09064710.2019.1606930.

[238] Bhupathyraaj, M., Halligudi, N., Sayigh, F. A. R. A., & Hakkak, M. (2022). Bioactives and

Pharmacology of Acacia Nilotica: A Review. Asian Journal of Applied Chemistry Research, 12(1), 24-29.

[239] Musa, H.H., Ahmed, A.A., Musa, T.H. (2019). Chemistry, Biological, and Pharmacological Properties of Gum Arabic. In: Mérillon, JM., Ramawat, K. (eds) Bioactive Molecules in Food. Reference Series in Phytochemistry. Springer, Cham. https://doi.org/10.1007/978-3-319-78030-6\_11.

[240] Rahman, Shahedur, and Rashida Parvin. "Therapeutic potential of Aegle marmelos (L.)-An overview." Asian Pacific Journal of Tropical Disease vol. 4,1 (2014): 71–77. doi:10.1016/S2222-1808(14)60318-2.

[241] Kianian, Farzaneh et al. "Pharmacological Properties of Allium cepa, Preclinical and Clinical Evidences; A Review." Iranian journal of pharmaceutical research : IJPR vol. 20,2 (2021): 107-134. doi:10.22037/ijpr.2020.112781.13946.

[242] Fredotović, Željana et al. "Chemical Composition and Biological Activity of Allium cepa L. and Allium × cornutum (Clementi ex Visiani 1842) Methanolic Extracts." Molecules (Basel, Switzerland) vol. 22,3 448. 11 Mar. 2017, doi:10.3390/molecules22030448.

[243] Hęś, M., Dziedzic, K., Górecka, D. et al. Aloe vera (L.) Webb.: Natural Sources of Antioxidants – A Review. Plant Foods Hum Nutr 74, 255–265 (2019). https://doi.org/10.1007/s11130-019-00747-5.

[244] Amalraj, Augustine, and Sreeraj Gopi. "Medicinal properties of Terminalia arjuna (Roxb.) Wight & Arn.: A review." Journal of traditional and complementary medicine vol. 7,1 65-78. 20 Mar. 2016, doi:10.1016/j.jtcme.2016.02.003.

[245] Ranasinghe, Priyanga et al. "Medicinal properties of 'true' cinnamon (Cinnamomum zeylanicum): a systematic review." BMC complementary and alternative medicine vol. 13 275. 22 Oct. 2013, doi:10.1186/1472-6882-13-275.

[246] Al-Snafi, A. E. (2016). A review on chemical constituents and pharmacological activities of Coriandrum sativum. IOSR Journal of Pharmacy, 6(7), 17-42.

[247] Soni, Priyanka et al. "Pharmacological properties of Datura stramonium L. as a potential medicinal tree: an overview." Asian Pacific journal of tropical biomedicine vol. 2,12 (2012): 1002-8. doi:10.1016/S2221-1691(13)60014-3

[248] Ayyanar, Muniappan, and Pandurangan Subash-Babu. "Syzygium cumini (L.) Skeels: a review of its phytochemical constituents and traditional uses." Asian Pacific journal of tropical biomedicine vol. 2,3 (2012): 240-6. doi:10.1016/S2221-1691(12)60050-1.

[249] Yadav UC, Baquer NZ. Pharmacological effects of Trigonella foenum-graecum L. in health and disease. Pharm Biol. 2014 Feb;52(2):243-54. doi: 10.3109/13880209.2013.826247. Epub 2013 Oct 9. PMID: 24102093.

[250] Chandrasekar, S B et al. "Phytopharmacology of Ficus religiosa." Pharmacognosy reviews vol. 4,8 (2010): 195-9. doi:10.4103/0973-7847.70918.

[251] Di Fabio G, Romanucci V, Di Marino C, Pisanti A, Zarrelli A. Gymnema sylvestre R. Br., an Indian medicinal herb: traditional uses, chemical composition, and biological activity. Curr Pharm Biotechnol. 2015;16(6):506-16. doi: 10.2174/138920101606150407112903. PMID: 25860062.

[252] Fan, Minxia et al. "Antioxidant and Anti-Proliferative Properties of Hagenia abyssinica Roots and Their Potentially Active Components." Antioxidants (Basel, Switzerland) vol. 9,2 143. 6 Feb. 2020, doi:10.3390/antiox9020143

[253] Bortolotti, Massimo et al. "Momordica charantia, a Nutraceutical Approach for Inflammatory Related Diseases." Frontiers in pharmacology vol. 10 486. 8 May. 2019, doi:10.3389/fphar.2019.00486

[254] Pattanayak, Priyabrata et al. "Ocimum sanctum Linn. A reservoir plant for therapeutic applications: An overview." Pharmacognosy reviews vol. 4,7 (2010): 95-105. doi:10.4103/0973-7847.65323

[255] Zaveri, M., Khandhar, A., Patel, S., & Patel, A. (2010). Chemistry and pharmacology of Piper longum L. International journal of pharmaceutical sciences review and research, 5(1), 67-76.

[256] de Lima, R. M. T., Dos Reis, A. C., de Menezes, A. A. P. M., Santos, J. V. D. O., Filho, J. W. G. D. O., Ferreira, J. R. D. O., ... & Melo-Cavalcante, A. A. D. C. (2018). Protective and therapeutic potential of ginger (Zingiber officinale) extract and [6]-gingerol in cancer: A comprehensive review. Phytotherapy research, 32(10), 1885-1907.

[257] Lipsky, M. S., & Sharp, L. K. (2001). From idea to market: the drug approval process. The Journal of the American Board of Family Practice, 14(5), 362-367.

[258] Long, Jianyan et al. "Overview of Clinical Trials on Type 2 Diabetes Mellitus: A Comprehensive Analysis of the ClinicalTrials.gov Database." Diabetes, metabolic syndrome and obesity : targets and therapy vol. 14 367-377. 26 Jan. 2021, doi:10.2147/DMSO.S288065.

[259] Hsia, Daniel S et al. "An update on sodium-glucose co-transporter-2 inhibitors for the treatment of diabetes mellitus." Current opinion in endocrinology, diabetes, and obesity vol. 24,1 (2017): 73-79. doi:10.1097/MED.00000000000311.

[260] Davis, H.E., McCorkell, L., Vogel, J.M. et al. Long COVID: major findings, mechanisms and recommendations. Nat Rev Microbiol 21, 133–146 (2023). https://doi.org/10.1038/s41579-022-00846-2.

[261] Dahlén, A. D., Dashi, G., Maslov, I., Attwood, M. M., Jonsson, J., Trukhan, V., & Schiöth, H. B. (2022). Trends in antidiabetic drug discovery: FDA approved drugs, new drugs in clinical trials and global sales. Frontiers in Pharmacology, 12, 4119.

[262] Deacon, C. F. (2018). A review of dipeptidyl peptidase-4 inhibitors. Hot topics from randomized controlled trials. Diabetes, Obesity and Metabolism, 20, 34-46.

[263] Tseng, C. H., Lee, K. Y., & Tseng, F. H. (2015). An updated review on cancer risk associated with incretin mimetics and enhancers. Journal of Environmental Science and Health, Part C, 33(1), 67-124.

[264] Quianzon, C. C., & Cheikh, I. (2012). History of insulin. Journal of community hospital internal medicine perspectives, 2(2), 18701.

[265] Shishtar, E., Sievenpiper, J. L., Djedovic, V., Cozma, A. I., Ha, V., Jayalath, V. H.,& Vuksan, V. (2014). The effect of ginseng (the genus panax) on glycemic control: a systematic review and meta-analysis of randomized controlled clinical trials. PloS one, 9(9), e107391.

[266] Sahib, A. S. (2016). Anti-diabetic and antioxidant effect of cinnamon in poorly controlled type-2 diabetic Iraqi patients: A randomized, placebo-controlled clinical trial. Journal of intercultural ethnopharmacology, 5(2), 108.

[267] S. N. Sankhwar, Pawan Kumar, Manashi Bagchi, Mehul Rungta, Debasis Bagchi. (2023) Safety and Efficacy of Furosap®, a Patented Trigonella foenum-graecum Seed Extract, in Boosting Testosterone Level, Reproductive Health and Mood Alleviation in Male Volunteers. Journal of the American Nutrition Association 42:1, pages 27-35.

[268] Soleimani, D., Paknahad, Z., & Rouhani, M. H. (2020). Therapeutic Effects of Garlic on Hepatic Steatosis in Nonalcoholic Fatty Liver Disease Patients: A Randomized Clinical Trial. Diabetes, metabolic syndrome and obesity : targets and therapy, 13, 2389–2397. https://doi.org/10.2147/DMSO.S254555

[269] Rahman, I. U., Khan, R. U., Rahman, K. U., & Bashir, M. (2015). Lower hypoglycemic but higher antiatherogenic effects of bitter melon than glibenclamide in type 2 diabetic patients. Nutrition journal, 14(1), 1-7.

[270] Pingali, U., Ali, M. A., Gundagani, S., & Nutalapati, C. (2020). Evaluation of the Effect of an Aqueous Extract of Azadirachta indica (Neem) Leaves and Twigs on Glycemic Control, Endothelial Dysfunction and Systemic Inflammation in Subjects with Type 2 Diabetes Mellitus - A Randomized, Double-Blind, Placebo-Controlled Clinical Study. Diabetes, metabolic syndrome and obesity : targets and therapy, 13, 4401–4412. https://doi.org/10.2147/DMSO.S274378

[271] Jafarpour-Sadegh, F., Montazeri, V., Adili, A., Esfehani, A., Rashidi, M. R., & Pirouzpanah, S. (2017). Consumption of fresh yellow onion ameliorates hyperglycemia and insulin resistance in breast cancer patients during doxorubicin-based chemotherapy: a randomized controlled clinical trial. Integrative Cancer Therapies, 16(3), 276-289.

[272] Zarvandi, M., Rakhshandeh, H., Abazari, M., Shafiee-Nick, R., & Ghorbani, A. (2017). Safety and efficacy of a polyherbal formulation for the management of dyslipidemia and hyperglycemia in patients with advanced-stage of type-2 diabetes. Biomedicine & Pharmacotherapy, 89, 69-75.

[273] Jeddy, N., Ravi, S., Radhika, T., & Sai Lakshmi, L. J. (2018). Comparison of the efficacy of herbal mouth rinse with commercially available mouth rinses: A clinical trial. Journal of oral and maxillofacial pathology : JOMFP, 22(3), 332–334. https://doi.org/10.4103/jomfp.JOMFP\_303\_18

[274] Kumar, V., Mahdi, F., Singh, R., Mahdi, A. A., & Singh, R. K. (2016). A clinical trial to assess the antidiabetic, antidyslipidemic and antioxidant activities of Tinospora cordifolia in management of type-2 diabetes mellitus. Int J Pharm Sci Res, 7(2), 757-64.

[275] Kianbakht, S., Khalighi-Sigaroodi, F., & Dabaghian, F. H. (2013). Improved glycemic control in patients with advanced type 2 diabetes mellitus taking Urtica dioica leaf extract: a randomized double-blind placebo-controlled clinical trial. Clin Lab, 59(9-10), 1071-6.

[276] Kaatabi, H., Bamosa, A. O., Badar, A., Al-Elq, A., Abou-Hozaifa, B., Lebda, F., ... & Al-Almaie, S. (2015). Nigella sativa improves glycemic control and ameliorates oxidative stress in patients with type 2 diabetes mellitus: placebo controlled participant blinded clinical trial. PloS one, 10(2), e0113486.

[277] Wasana, K. G. P., Attanayake, A. P., Weerarathna, T. P., & Jayatilaka, K. A. P. W. (2021). Efficacy and safety of a herbal drug of Coccinia grandis (Linn.) Voigt in patients with type 2 diabetes mellitus: A double blind randomized placebo controlled clinical trial. Phytomedicine, 81, 153431.

[278] Shinjyo, N., Waddell, G., & Green, J. (2020). A tale of two cinnamons: A comparative review of the clinical evidence of Cinnamomum verum and C. cassia as diabetes interventions. Journal of herbal medicine, 21, 100342.

[279] Khalili, N., Fereydoonzadeh, R., Mohtashami, R., Mehrzadi, S., Heydari, M., & Huseini, H. F. (2017). Silymarin, Olibanum, and Nettle, a mixed herbal formulation in the treatment of type II

diabetes: a randomized, double-blind, placebo-controlled, clinical trial. Journal of evidence-based complementary & alternative medicine, 22(4), 603-608.

[280] Ruyvaran, M., Zamani, A., Mohamadian, A., Zarshenas, M. M., Eftekhari, M. H., Pourahmad, S., ... & Nimrouzi, M. (2022). Safflower (Carthamus tinctorius L.) oil could improve abdominal obesity, blood pressure, and insulin resistance in patients with metabolic syndrome: A randomized, double-blind, placebo-controlled clinical trial. Journal of Ethnopharmacology, 282, 114590.

[281] Rasool Soltani, Syed Mustafa Ghanadian, Bijan Iraj, Alireza Homayouni, Tanin Shahmiveh Esfahani, Mojtaba Akbari, "The Effects of Berberis integerrima Fruit Extract on Glycemic Control Parameters in Patients with Type 2 Diabetes Mellitus: A Randomized Controlled Clinical Trial", Evidence-Based Complementary and Alternative Medicine, vol. 2021, Article ID 5583691, 7 pages, 2021. https://doi.org/10.1155/2021/5583691

[282] Park, S. H., Oh, M. R., Choi, E. K., Kim, M. G., Ha, K. C., Lee, S. K., ... & Chae, S. W. (2014). An 8-wk, randomized, double-blind, placebo-controlled clinical trial for the antidiabetic effects of hydrolyzed ginseng extract. Journal of Ginseng Research, 38(4), 239-243.

[283] Sahebkar-Khorasani, M., Jarahi, L., Cramer, H., Safarian, M., Naghedi-Baghdar, H., Salari, R., ... & Azizi, H. (2019). Herbal medicines for suppressing appetite: A systematic review of randomized clinical trials. Complementary therapies in medicine, 44, 242-252.

[284] Azadmehr, A., Ziaee, A., Ghanei, L., Fallah Huseini, H., Hajiaghaee, R., Tavakoli-Far, B., & Kordafshari, G. (2014). A Randomized Clinical Trial Study: Anti-Oxidant, Anti-hyperglycemic and Anti-Hyperlipidemic Effects of Olibanum Gum in Type 2 Diabetic Patients. Iranian journal of pharmaceutical research : IJPR, 13(3), 1003–1009.

[285] Wainstein, J., Landau, Z., Dayan, Y. B., Jakubowicz, D., Grothe, T., Perrinjaquet-Moccetti, T., & Boaz, M. (2016). Purslane extract and glucose homeostasis in adults with type 2 diabetes: A double-blind, placebo-controlled clinical trial of efficacy and safety. Journal of medicinal food, 19(2), 133-140.

[286] Rizvi, Syed A A, and Ayman M Saleh. "Applications of nanoparticle systems in drug delivery technology." Saudi pharmaceutical journal : SPJ : the official publication of the Saudi Pharmaceutical Society vol. 26,1 (2018): 64-70. doi:10.1016/j.jsps.2017.10.012.

[287] Tiwari, Gaurav et al. "Drug delivery systems: An updated review." International journal of pharmaceutical investigation vol. 2,1 (2012): 2-11. doi:10.4103/2230-973X.96920.

[288] Kamaly, N., Yameen, B., Wu, J., & Farokhzad, O. C. (2016). Degradable controlled-release polymers and polymeric nanoparticles: mechanisms of controlling drug release. Chemical reviews, 116(4), 2602-2663.

[289] Patra, Jayanta Kumar et al. "Nano based drug delivery systems: recent developments and future prospects." Journal of nanobiotechnology vol. 16,1 71. 19 Sep. 2018, doi:10.1186/s12951-018-0392-8.

[290] Sim, Serjay, and Nyet Kui Wong. "Nanotechnology and its use in imaging and drug delivery (Review)." Biomedical reports vol. 14,5 (2021): 42. doi:10.3892/br.2021.1418.

[291] Farjadian, Fatemeh et al. "Nanopharmaceuticals and nanomedicines currently on the market: challenges and opportunities." Nanomedicine (London, England) vol. 14,1 (2019): 93-126. doi:10.2217/nnm-2018-0120.

[292] Kadry, H., Noorani, B. & Cucullo, L. A blood-brain barrier overview on structure, function,

impairment, and biomarkers of integrity. Fluids Barriers CNS 17, 69 (2020). https://doi.org/10.1186/s12987-020-00230-3.

[293] Glassman, Patrick M, and Vladimir R Muzykantov. "Pharmacokinetic and Pharmacodynamic Properties of Drug Delivery Systems." The Journal of pharmacology and experimental therapeutics vol. 370,3 (2019): 570-580. doi:10.1124/jpet.119.257113.

[294] Mansour A, Romani M, Acharya AB, Rahman B, Verron E, Badran Z. Drug Delivery Systems in Regenerative Medicine: An Updated Review. Pharmaceutics. 2023; 15(2):695. https://doi.org/10.3390/pharmaceutics15020695.

[295] Trucillo P. Drug Carriers: Classification, Administration, Release Profiles, and Industrial Approach. Processes. 2021; 9(3):470. https://doi.org/10.3390/pr9030470.

[296] Adepu, Shivakalyani, and Seeram Ramakrishna. "Controlled Drug Delivery Systems: Current Status and Future Directions." Molecules (Basel, Switzerland) vol. 26,19 5905. 29 Sep. 2021, doi:10.3390/molecules26195905.

[297] Mansoor, Shazia et al. "Polymer-Based Nanoparticle Strategies for Insulin Delivery." Polymers vol. 11,9 1380. 22 Aug. 2019, doi:10.3390/polym11091380.

[298] Wang, Jinqiang et al. "Injectable Biodegradable Polymeric Complex for Glucose-Responsive Insulin Delivery." ACS nano vol. 15,3 (2021): 4294-4304. doi:10.1021/acsnano.0c07291.

[299] Kazi, Karim Masud et al. "Niosome: A future of targeted drug delivery systems." Journal of advanced pharmaceutical technology & research vol. 1,4 (2010): 374-80. doi:10.4103/0110-5558.76435

[300] Gharbavi, Mahmoud et al. "Niosome: A Promising Nanocarrier for Natural Drug Delivery through Blood-Brain Barrier." Advances in pharmacological sciences vol. 2018 6847971. 11 Dec. 2018, doi:10.1155/2018/6847971

[301]Coelho, Jorge F et al. "Drug delivery systems: Advanced technologies potentially applicable in personalized treatments." The EPMA journal vol. 1,1 (2010): 164-209. doi:10.1007/s13167-010-0001-x

[302] Stewart, Sarah A et al. "Implantable Polymeric Drug Delivery Devices: Classification, Manufacture, Materials, and Clinical Applications." Polymers vol. 10,12 1379. 12 Dec. 2018, doi:10.3390/polym10121379.

[303] Begines, Belén et al. "Polymeric Nanoparticles for Drug Delivery: Recent Developments and Future Prospects." Nanomaterials (Basel, Switzerland) vol. 10,7 1403. 19 Jul. 2020, doi:10.3390/nano10071403.

[304] DiSanto, Rocco Michael et al. "Recent advances in nanotechnology for diabetes treatment." Wiley interdisciplinary reviews. Nanomedicine and nanobiotechnology vol. 7,4 (2015): 548-64. doi:10.1002/wnan.1329.

[305] Veiseh, O., Tang, B., Whitehead, K. et al. Managing diabetes with nanomedicine: challenges and opportunities. Nat Rev Drug Discov 14, 45–57 (2015). https://doi.org/10.1038/nrd4477.

[306] The synergistic properties of plant and metal NPs are unique in phytonanotherapy because they offer clinically bioequivalent effects to many synthetic drugs, with minimum side effects.

[307] Anand K, Tiloke C, Naidoo P, Chuturgoon AA. Phytonanotherapy for management of diabetes using green synthesis nanoparticles. J Photochem Photobiol B. 2017 Aug;173:626-639. doi: 10.1016/j.jphotobiol.2017.06.028. Epub 2017 Jun 27. PMID: 28709077.

[308] Karimi, Ali et al. "Herbal versus synthetic drugs; beliefs and facts." Journal of 2186

nephropharmacology vol. 4,1 27-30. 1 Jan. 201

[309] Wang, L., Wang, N., Zhang, W. et al. Therapeutic peptides: current applications and future directions. Sig Transduct Target Ther 7, 48 (2022). https://doi.org/10.1038/s41392-022-00904-4.

[310] Scioli Montoto, Sebastián et al. "Solid Lipid Nanoparticles for Drug Delivery: Pharmacological and Biopharmaceutical Aspects." Frontiers in molecular biosciences vol. 7 587997. 30 Oct. 2020, doi:10.3389/fmolb.2020.587997.

[311] Iravani, S et al. "Synthesis of silver nanoparticles: chemical, physical and biological methods." Research in pharmaceutical sciences vol. 9,6 (2014): 385-406.

[312] Javed, R., Zia, M., Naz, S. et al. Role of capping agents in the application of nanoparticles in biomedicine and environmental remediation: recent trends and future prospects. J Nanobiotechnol 18, 172 (2020). https://doi.org/10.1186/s12951-020-00704-4

[313] Atreja, Ashish et al. "Strategies to enhance patient adherence: making it simple." MedGenMed : Medscape general medicine vol. 7,1 4. 16 Mar. 2005

[314] Ansari SH, Islam F, Sameem M. Influence of nanotechnology on herbal drugs: A Review. J Adv Pharm Technol Res. 2012 Jul;3(3):142-6. doi: 10.4103/2231-4040.101006. PMID: 23057000; PMCID: PMC3459443.

[315] Xu, Jia-Jie et al. "Metal nanoparticles as a promising technology in targeted cancer treatment." Drug delivery vol. 29,1 (2022): 664-678. doi:10.1080/10717544.2022.2039804.

[316] Dikshit PK, Kumar J, Das AK, Sadhu S, Sharma S, Singh S, Gupta PK, Kim BS. Green Synthesis of Metallic Nanoparticles: Applications and Limitations. Catalysts. 2021; 11(8):902. https://doi.org/10.3390/catal11080902.

[317] Li, Yan et al. "Advances in oral peptide drug nanoparticles for diabetes mellitus treatment." Bioactive materials vol. 15 392-408. 28 Feb. 2022, doi:10.1016/j.bioactmat.2022.02.025.

[318] Nie, Xin et al. "Oral Nano Drug Delivery Systems for the Treatment of Type 2 Diabetes Mellitus: An Available Administration Strategy for Antidiabetic Phytocompounds." International journal of nanomedicine vol. 15 10215-10240. 16 Dec. 2020, doi:10.2147/IJN.S285134.

[319] Srinoi P, Chen Y-T, Vittur V, Marquez MD, Lee TR. Bimetallic Nanoparticles: Enhanced Magnetic and Optical Properties for Emerging Biological Applications. Applied Sciences. 2018; 8(7):1106. https://doi.org/10.3390/app8071106.

[320] Li YV. Zinc and insulin in pancreatic beta-cells. Endocrine. 2014 Mar;45(2):178-89. doi: 10.1007/s12020-013-0032-x. Epub 2013 Aug 24. PMID: 23979673.

[321] Kostov, Krasimir. "Effects of Magnesium Deficiency on Mechanisms of Insulin Resistance in Type 2 Diabetes: Focusing on the Processes of Insulin Secretion and Signaling." International journal of molecular sciences vol. 20,6 1351. 18 Mar. 2019, doi:10.3390/ijms20061351.

[322] Rzigalinski, Beverly A et al. "Cerium oxide nanoparticles in neuroprotection and considerations for efficacy and safety." Wiley interdisciplinary reviews. Nanomedicine and nanobiotechnology vol. 9,4 (2017): 10.1002/wnan.1444. doi:10.1002/wnan.1444

[323] Faisal S, Al-Radadi NS, Jan H, Abdullah, Shah SA, Shah S, Rizwan M, Afsheen Z, Hussain Z, Uddin MN, Idrees M, Bibi N. Curcuma longa Mediated Synthesis of Copper Oxide, Nickel Oxide and Cu-Ni Bimetallic Hybrid Nanoparticles: Characterization and Evaluation for Antimicrobial, Anti-Parasitic and Cytotoxic Potentials. Coatings. 2021; 11(7):849. https://doi.org/10.3390/coatings11070849.

[324] Malyugina S, Skalickova S, Skladanka J, Slama P, Horky P. Biogenic Selenium

Nanoparticles in Animal Nutrition: A Review. Agriculture. 2021; 11(12):1244. https://doi.org/10.3390/agriculture11121244

[325] Kieliszek, M., Bano, I. & Zare, H. A Comprehensive Review on Selenium and Its Effects on Human Health and Distribution in Middle Eastern Countries. Biol Trace Elem Res 200, 971–987 (2022). https://doi.org/10.1007/s12011-021-02716-z.

[326] Zhang, Xi-Feng et al. "Silver Nanoparticles: Synthesis, Characterization, Properties, Applications, and Therapeutic Approaches." International journal of molecular sciences vol. 17,9 1534. 13 Sep. 2016, doi:10.3390/ijms17091534

[327] Zhang, Xi-Feng et al. "Silver Nanoparticles: Synthesis, Characterization, Properties, Applications, and Therapeutic Approaches." International journal of molecular sciences vol. 17,9 1534. 13 Sep. 2016, doi:10.3390/ijms17091534.

[328] Bruna, Tamara et al. "Silver Nanoparticles and Their Antibacterial Applications." International journal of molecular sciences vol. 22,13 7202. 4 Jul. 2021, doi:10.3390/ijms22137202

[329] Zhao, Ruichen et al. "Drug Delivery System in the Treatment of Diabetes Mellitus." Frontiers in bioengineering and biotechnology vol. 8 880. 29 Jul. 2020, doi:10.3389/fbioe.2020.00880

[330] Shamsi-Goushki, A., Mortazavi, Z., Mirshekar, M. A., Mohammadi, M., Moradi-Kor, N., Jafari-Maskouni, S., & Shahraki, M. (2020). Comparative effects of curcumin versus nanocurcumin on insulin resistance, serum levels of apelin and lipid profile in type 2 diabetic rats. Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy, 13, 2337

[331] Nouri, Z., Hajialyani, M., Izadi, Z., Bahramsoltani, R., Farzaei, M. H., & Abdollahi, M. (2020). Nanophytomedicines for the Prevention of Metabolic Syndrome: A Pharmacological and Biopharmaceutical Review. Frontiers in bioengineering and biotechnology, 8, 425. https://doi.org/10.3389/fbioe.2020.00425.

[332] Al-Ishaq, R. K., Abotaleb, M., Kubatka, P., Kajo, K., & Büsselberg, D. (2019). Flavonoids and Their Anti-Diabetic Effects: Cellular Mechanisms and Effects to Improve Blood Sugar Levels. Biomolecules, 9(9), 430. https://doi.org/10.3390/biom9090430

[333] Malapermal, V., Botha, I., Krishna, S. B. N., & Mbatha, J. N. (2017). Enhancing antidiabetic and antimicrobial performance of Ocimum basilicum, and Ocimum sanctum (L.) using silver nanoparticles. Saudi journal of biological sciences, 24(6), 1294–1305. https://doi.org/10.1016/j.sjbs.2015.06.026

[334] Chavda, V. P., Patel, A. B., Mistry, K. J., Suthar, S. F., Wu, Z. X., Chen, Z. S., & Hou, K. (2022). Nano-Drug Delivery Systems Entrapping Natural Bioactive Compounds for Cancer: Recent Progress and Future Challenges. Frontiers in oncology, 12, 867655. https://doi.org/10.3389/fonc.2022.867655

[335] Sercombe, L., Veerati, T., Moheimani, F., Wu, S. Y., Sood, A. K., & Hua, S. (2015). Advances and Challenges of Liposome Assisted Drug Delivery. Frontiers in pharmacology, 6, 286. https://doi.org/10.3389/fphar.2015.00286

[336] Odei-Addo, F., Shegokar, R., Müller, R. H., Levendal, R. A., & Frost, C. (2017). Nanoformulation of Leonotis leonurus to improve its bioavailability as a potential antidiabetic drug. 3 Biotech, 7(5), 344. https://doi.org/10.1007/s13205-017-0986-0

[337] Abdifetah, O., & Na-Bangchang, K. (2019). Pharmacokinetic studies of nanoparticles as a delivery system for conventional drugs and herb-derived compounds for cancer therapy: a systematic review. International journal of nanomedicine, 14, 5659–5677.

2189

### https://doi.org/10.2147/IJN.S213229

[338] Nouri, Z., Hajialyani, M., Izadi, Z., Bahramsoltani, R., Farzaei, M. H., & Abdollahi, M. (2020). Nanophytomedicines for the Prevention of Metabolic Syndrome: A Pharmacological and Biopharmaceutical Review. Frontiers in bioengineering and biotechnology, 8, 425. https://doi.org/10.3389/fbioe.2020.00425

[339] Dewanjee, S., Chakraborty, P., Mukherjee, B., & De Feo, V. (2020). Plant-Based Antidiabetic Nanoformulations: The Emerging Paradigm for Effective Therapy. International journal of molecular sciences, 21(6), 2217. https://doi.org/10.3390/ijms21062217

[340] Bitencourt, P. E., Cargnelutti, L. O., Stein, C. S., Lautenchleger, R., Ferreira, L. M., Sangoi, M., Denardi, L., Borges, R. M., Boligon, A., Moresco, R. N., Cruz, L., Zanette, R. A., Alves, S. H., & Moretto, M. B. (2017). Nanoparticle formulation increases Syzygium cumini antioxidant activity in Candida albicans-infected diabetic rats. Pharmaceutical biology, 55(1), 1082–1088. https://doi.org/10.1080/13880209.2017.1283338

[341] Soorya, C., Balamurugan, S., Ramya, S., Neethirajan, K., Kandeepan, C., & Jayakumararaj, R. (2021). Physicochemical, ADMET and Druggable properties of Myricetin: A Key Flavonoid in Syzygium cumini that regulates metabolic inflammations. Journal of Drug Delivery and Therapeutics, 11(4), 66-73.

[342] Abdifetah, O., & Na-Bangchang, K. (2019). Pharmacokinetic studies of nanoparticles as a delivery system for conventional drugs and herb-derived compounds for cancer therapy: a systematic review. International journal of nanomedicine, 14, 5659–5677. https://doi.org/10.2147/IJN.S213229

[343] Shariare, M.H., Rahman, M., Lubna, S.R. et al. Liposomal drug delivery of Aphanamixis polystachya leaf extracts and its neurobehavioral activity in mice model. Sci Rep 10, 6938 (2020). https://doi.org/10.1038/s41598-020-63894-9.

[344] Vijayakumar S, Vaseeharan B, Malaikozhundan B, Gopi N, Ekambaram P, Pachaiappan R, Velusamy P, Murugan K, Benelli G, Suresh Kumar R, Suriyanarayanamoorthy M. Therapeutic effects of gold nanoparticles synthesized using Musa paradisiaca peel extract against multiple antibiotic resistant Enterococcus faecalis biofilms and human lung cancer cells (A549). Microb Pathog. 2017 Jan;102:173-183. doi: 10.1016/j.micpath.2016.11.029. Epub 2016 Dec 1. PMID: 27916691.

[345] Daisy, P., & Saipriya, K. (2012). Biochemical analysis of Cassia fistula aqueous extract and phytochemically synthesized gold nanoparticles as hypoglycemic treatment for diabetes mellitus. International journal of nanomedicine, 7, 1189–1202. https://doi.org/10.2147/IJN.S26650

[346] Taghipour, Y. D., Hajialyani, M., Naseri, R., Hesari, M., Mohammadi, P., Stefanucci, A., Mollica, A., Farzaei, M. H., & Abdollahi, M. (2019). Nanoformulations of natural products for management of metabolic syndrome. International journal of nanomedicine, 14, 5303–5321. https://doi.org/10.2147/IJN.S213831

[347] Mota AH, Duarte N, Serra AT, Ferreira A, Bronze MR, Custódio L, Gaspar MM, Simões S, Rijo P, Ascensão L, Faísca P, Viana AS, Pinto R, Kumar P, Almeida AJ, Reis CP. Further Evidence of Possible Therapeutic Uses of Sambucus nigra L. Extracts by the Assessment of the In Vitro and In Vivo Anti-Inflammatory Properties of Its PLGA and PCL-Based Nanoformulations.

Pharmaceutics. 2020; 12(12):1181. https://doi.org/10.3390/pharmaceutics12121181

[348] Nouri, Z., Hajialyani, M., Izadi, Z., Bahramsoltani, R., Farzaei, M. H., & Abdollahi, M.

(2020). Nanophytomedicines for the Prevention of Metabolic Syndrome: A Pharmacological and Biopharmaceutical Review. Frontiers in bioengineering and biotechnology, 8, 425. https://doi.org/10.3389/fbioe.2020.00425

[349] Nouri, Z., Hajialyani, M., Izadi, Z., Bahramsoltani, R., Farzaei, M. H., & Abdollahi, M. (2020). Nanophytomedicines for the Prevention of Metabolic Syndrome: A Pharmacological and Biopharmaceutical Review. Frontiers in bioengineering and biotechnology, 8, 425. https://doi.org/10.3389/fbioe.2020.00425

[350] Dewanjee, S., Chakraborty, P., Mukherjee, B., & De Feo, V. (2020). Plant-Based Antidiabetic Nanoformulations: The Emerging Paradigm for Effective Therapy. International journal of molecular sciences, 21(6), 2217. https://doi.org/10.3390/ijms21062217

[351] Olawoye OS, Adeagbo BA, Bolaji OO. Moringa oleifera leaf powder alters the pharmacokinetics of amodiaquine in healthy human volunteers. J Clin Pharm Ther. 2018 Oct;43(5):626-632. doi: 10.1111/jcpt.12725. Epub 2018 Jun 19. PMID: 29920710.

[352] Malviya, R., Raj, S., Fuloria, S., Subramaniyan, V., Sathasivam, K., Kumari, U., Unnikrishnan Meenakshi, D., Porwal, O., Hari Kumar, D., Singh, A., Chakravarthi, S., & Kumar Fuloria, N. (2021). Evaluation of Antitumor Efficacy of Chitosan-Tamarind Gum Polysaccharide Polyelectrolyte Complex Stabilized Nanoparticles of Simvastatin. International journal of nanomedicine, 16, 2533–2553. https://doi.org/10.2147/IJN.S300991

[353] Shinta Ayu Nurfaradilla, Fadlina Chany Saputri, Yahdiana Harahap, "Pharmacokinetic Herb-Drug Interaction between Hibiscus sabdariffa Calyces Aqueous Extract and Captopril in Rats", Evidence-Based Complementary and Alternative Medicine, vol. 2020, Article ID 5013898, 8 pages, 2020. https://doi.org/10.1155/2020/5013898

[354] Carvalho, F. V., Ribeiro, L. N. M., Moura, L. D., Rodrigues da Silva, G. H., Mitsutake, H., Mendonça, T. C., Geronimo, G., Breitkreitz, M. C., & de Paula, E. (2022). Docetaxel Loaded in Copaiba Oil-Nanostructured Lipid Carriers as a Promising DDS for Breast Cancer Treatment. Molecules (Basel, Switzerland), 27(24), 8838. https://doi.org/10.3390/molecules27248838

[355] Javed, R., Ain, N. U., Gul, A., Arslan Ahmad, M., Guo, W., Ao, Q., & Tian, S. (2022). Diverse biotechnological applications of multifunctional titanium dioxide nanoparticles: An up-to-date review. IET nanobiotechnology, 16(5), 171-189.

[356] Subramanyam GK, Gaddam SA, Kotakadi VS, Palithya S, Penchalaneni J, Challagundla VN. Argyreia nervosa (Samudra pala) leaf extract mediated silver nanoparticles and evaluation of their antioxidant, antibacterial activity, in vitro anticancer and apoptotic studies in KB oral cancer cell lines. Artif Cells Nanomed Biotechnol. 2021 Dec;49(1):635-650. doi: 10.1080/21691401.2021.1996384. PMID: 34738487.

[357] Nouri, Z., Hajialyani, M., Izadi, Z., Bahramsoltani, R., Farzaei, M. H., & Abdollahi, M. (2020). Nanophytomedicines for the Prevention of Metabolic Syndrome: A Pharmacological and Biopharmaceutical Review. Frontiers in bioengineering and biotechnology, 8, 425. https://doi.org/10.3389/fbioe.2020.00425

[358] Surya, C., John, N. A. A., Pandiyan, V., Ravikumar, S., Amutha, P., Sobral, A. J., & Krishnakumar, B. (2019). Costus speciosus leaf extract assisted CS-Pt-TiO2 composites: Synthesis, characterization and their bio and photocatalytic applications. Journal of Molecular Structure, 1195, 787-795.

[359] Priyanka, K., Kosuru, R., Sharma, R. P., Sahu, P. L., & Singh, S. (2017). Assessment of 2190

pharmacokinetic parameters of lupeol in Ficus religiosa L. extract after oral administration of suspension and solid lipid nanoparticles to Wistar rats. Journal of Drug Delivery Science and Technology, 41, 58-67.

[360] Salunkhe R, Gadgoli C, Naik A, Patil N. Pharmacokinetic Profile and Oral Bioavailability of Diosgenin, Charantin, and Hydroxychalcone From a Polyherbal Formulation. Front Pharmacol. 2021 Apr 29;12:629272. doi: 10.3389/fphar.2021.629272. PMID: 33995027; PMCID: PMC8117003.

[361] Li LL, Cui Y, Guo XH, Ma K, Tian P, Feng J, Wang JM. Pharmacokinetics and Tissue Distribution of Gingerols and Shogaols from Ginger (Zingiber officinale Rosc.) in Rats by UPLC<sup>-</sup>Q-Exactive<sup>-</sup>HRMS. Molecules. 2019 Jan 31;24(3):512. doi: 10.3390/molecules24030512. PMID: 30708987; PMCID: PMC6384666.

[362] Di Costanzo, A., & Angelico, R. (2019). Formulation Strategies for Enhancing the Bioavailability of Silymarin: The State of the Art. Molecules (Basel, Switzerland), 24(11), 2155. https://doi.org/10.3390/molecules24112155

[363] Taghavinia, F., Teymouri, F., Farokhrouz, F., Bagherabad, E. H., Farjami, S., Karimi, E., ... & Shakeri, M. (2022). Nanoliposome-Loaded Phenolics from Nasturtium officinale Improves Health Parameters in a Colorectal Cancer Mouse Model. Animals, 12(24), 3492.

[364] Wang, R., Ding, Y., Liu, R., Xiang, L., & Du, L. (2010). Pomegranate: constituents, bioactivities and pharmacokinetics. Fruit, vegetable and cereal science and biotechnology, 4(2), 77-87.

[365] Mei M, Ruan JQ, Wu WJ, Zhou RN, Lei JP, Zhao HY, Yan R, Wang YT. In vitro pharmacokinetic characterization of mulberroside A, the main polyhydroxylated stilbene in mulberry (Morus alba L.), and its bacterial metabolite oxyresveratrol in traditional oral use. J Agric Food Chem. 2012 Mar 7;60(9):2299-308. doi: 10.1021/jf204495t. Epub 2012 Feb 22. PMID: 22225542.

[366] Li Y, Deng S, Zhao Y, Liu L, Zhao R. Smilax glabra Rhizoma affects the pharmacokinetics and tissue distribution of methotrexate by increasing the P glycoprotein mRNA expression in rats after oral administration. Mol Med Rep. 2017 Nov;16(5):7633-7640. doi: 10.3892/mmr.2017.7559. Epub 2017 Sep 20. PMID: 28944899.

[367] Gera, M., Sharma, N., Ghosh, M., Huynh, D. L., Lee, S. J., Min, T., Kwon, T., & Jeong, D. K. (2017). Nanoformulations of curcumin: an emerging paradigm for improved remedial application. Oncotarget, 8(39), 66680–66698. https://doi.org/10.18632/oncotarget.19164

[368] Yu CP, Yang MS, Hsu PW, Lin SP, Hou YC. Bidirectional Influences of Cranberry on the Pharmacokinetics and Pharmacodynamics of Warfarin with Mechanism Elucidation. Nutrients. 2021 Sep 16;13(9):3219. doi: 10.3390/nu13093219. PMID: 34579096; PMCID: PMC8470483.

[369] Yang R, Wang M, Ma X, Gao Q. Development of Silver nanoparticles green-formulated by Matricaria chamomilla as novel chemotherapeutic nanoformulation for the treatment of oral squamous cell carcinoma. Comb Chem High Throughput Screen. 2022 Nov 16. doi: 10.2174/1386207326666221116101621. Epub ahead of print. PMID: 36397627.

[370] Ibrahim A, Abdel Gaber SA, Fawzi Kabil M, Ahmed-Farid OAH, Hirsch AKH, El-Sherbiny IM, Nasr M. Baicalin lipid nanocapsules for treatment of glioma: characterization, mechanistic cytotoxicity, and pharmacokinetic evaluation. Expert Opin Drug Deliv. 2022 Nov;19(11):1549-1560. doi: 10.1080/17425247.2022.2139370. Epub 2022 Oct 31. PMID: 36287914.

[371] Ibrahim A, Abdel Gaber SA, Fawzi Kabil M, Ahmed-Farid OAH, Hirsch AKH, El-Sherbiny IM, Nasr M. Baicalin lipid nanocapsules for treatment of glioma: characterization, mechanistic cytotoxicity, and pharmacokinetic evaluation. Expert Opin Drug Deliv. 2022 Nov;19(11):1549-1560. doi: 10.1080/17425247.2022.2139370. Epub 2022 Oct 31. PMID: 36287914.

[372] Barakat, S., Nasr, M., Ahmed, R.F. et al. Recent Formulation Advances of Mangiferin. Rev. Bras. Farmacogn. 32, 871–882 (2022). https://doi.org/10.1007/s43450-022-00297-z

[373] Zhao D, Feng SX, Zhang HJ, Zhang N, Liu XF, Wan Y, Zhou YX, Li JS. Pharmacokinetics, tissue distribution and excretion of five rhubarb anthraquinones in rats after oral administration of effective fraction of anthraquinones from rheum officinale. Xenobiotica. 2021 Aug;51(8):916-925. doi: 10.1080/00498254.2021.1940353. Epub 2021 Jun 28. PMID: 34110981.

[374] Veenstra, J. P., Vemu, B., Tocmo, R., Nauman, M. C., & Johnson, J. J. (2021). Pharmacokinetic Analysis of Carnosic Acid and Carnosol in Standardized Rosemary Extract and the Effect on the Disease Activity Index of DSS-Induced Colitis. Nutrients, 13(3), 773. https://doi.org/10.3390/nu13030773

[375] Khoshandam A, Imenshahidi M, Hosseinzadeh H. Pharmacokinetic of berberine, the main constituent of Berberis vulgaris L.: A comprehensive review. Phytother Res. 2022 Nov;36(11):4063-4079. doi: 10.1002/ptr.7589. Epub 2022 Oct 11. PMID: 36221815

[376] Islam MR, Rahman MM, Dhar PS, Nowrin FT, Sultana N, Akter M, Rauf A, Khalil AA, Gianoncelli A, Ribaudo G. The Role of Natural and Semi-Synthetic Compounds in Ovarian Cancer: Updates on Mechanisms of Action, Current Trends and Perspectives. Molecules. 2023; 28(5):2070. https://doi.org/10.3390/molecules28052070

[377] Zhang XF, Liu ZG, Shen W, Gurunathan S. Silver Nanoparticles: Synthesis, Characterization, Properties, Applications, and Therapeutic Approaches. Int J Mol Sci 2016;17:E1534. [PMID: 27649147 DOI: 10.3390/ijms17091534.

[378] Duong TT, Isomäki A, Paaver U, Laidmäe I, Tõnisoo A, Yen TTH, Kogermann K, Raal A, Heinämäki J, Pham TM. Nanoformulation and Evaluation of Oral Berberine-Loaded Liposomes. Molecules. 2021 Apr 29;26(9):2591. doi: 10.3390/molecules26092591. PMID: 33946815; PMCID: PMC8125214.

[379] Feng X, Wang K, Cao S, Ding L, Qiu F. Pharmacokinetics and Excretion of Berberine and Its Nine Metabolites in Rats. Front Pharmacol. 2021 Jan 15;11:594852. doi: 10.3389/fphar.2020.594852. PMID: 33584274; PMCID: PMC7874128.

[380] Derosa, G., Bonaventura, A., Bianchi, L., Romano, D., D'angelo, A., Fogari, E., & Maffioli, P. (2013). Berberis aristata/Silybum marianum fixed combination on lipid profile and insulin secretion in dyslipidemic patients. Expert opinion on biological therapy, 13(11), 1495-1506.

[381] Saha, S., Mahar, R., Pal, D. (2023). Natural Products in Controlling and Treatment of Cancers and Genital Warts Caused by Different Viruses. In: Pal, D. (eds) Anti-Viral Metabolites from Medicinal Plants. Reference Series in Phytochemistry. Springer, Cham. https://doi.org/10.1007/978-3-030-83350-3\_24-1.

[382] Ye M, Fu S, Pi R, He F. Neuropharmacological and pharmacokinetic properties of berberine: a review of recent research. J Pharm Pharmacol. 2009 Jul;61(7):831-7. doi: 10.1211/jpp/61.07.0001. PMID: 19589224.

[383] Subramanian, A. P., Jaganathan, S. K., Manikandan, A., Pandiaraj, K. N., Gomathi, N., & Supriyanto, E. (2016). Recent trends in nano-based drug delivery systems for efficient delivery of

phytochemicals in chemotherapy. RSC advances, 6(54), 48294-48314.

[384] Nouri, Z., Hajialyani, M., Izadi, Z., Bahramsoltani, R., Farzaei, M. H., & Abdollahi, M. (2020). Nanophytomedicines for the prevention of metabolic syndrome: a pharmacological and biopharmaceutical review. Frontiers in Bioengineering and Biotechnology, 425.

[385] Salama, M., & Ezzat, S. M. (2020). Bioactive lead compounds and molecular targets for the treatment of heart disease. Phytochemicals as Lead Compounds for New Drug Discoverys.

[386] Gera, M., Sharma, N., Ghosh, M., Huynh, D. L., Lee, S. J., Min, T., Kwon, T., & Jeong, D. K. (2017). Nanoformulations of curcumin: an emerging paradigm for improved remedial application. Oncotarget, 8(39), 66680–66698. https://doi.org/10.18632/oncotarget.19164.

[387] Pasquini R, Scassellati-Sforzolini G, Villarini M, Moretti M, Marcarelli M, Fatigoni C, Kaur S, Kumar S, Grover IS. In vitro protective effects of Terminalia arjuna bark extracts against the 4nitroquinoline-N-oxide genotoxicity. J Environ Pathol Toxicol Oncol. 2002;21(1):33-44. PMID: 11934011.

[388] Gallwitz B. (2019). Clinical Use of DPP-4 Inhibitors. Frontiers in endocrinology, 10, 389. https://doi.org/10.3389/fendo.2019.00389

[389] Gomez-Peralta, F., Abreu, C., Gomez-Rodriguez, S., Barranco, R. J., & Umpierrez, G. E. (2018). Safety and Efficacy of DPP4 Inhibitor and Basal Insulin in Type 2 Diabetes: An Updated Review and Challenging Clinical Scenarios. Diabetes therapy : research, treatment and education of diabetes and related disorders, 9(5), 1775–1789. https://doi.org/10.1007/s13300-018-0488-z

[390] Tran, N., Pham, B., & Le, L. (2020). Bioactive Compounds in Anti-Diabetic Plants: From Herbal Medicine to Modern Drug Discovery. Biology, 9(9), 252.

https://doi.org/10.3390/biology9090252

[391] Nyakudya TT, Tshabalala T, Dangarembizi R, Erlwanger KH, Ndhlala AR. The Potential Therapeutic Value of Medicinal Plants in the Management of Metabolic Disorders. Molecules. 2020; 25(11):2669. https://doi.org/10.3390/molecules25112669.

[392] Din, F. U., Aman, W., Ullah, I., Qureshi, O. S., Mustapha, O., Shafique, S., & Zeb, A. (2017). Effective use of nanocarriers as drug delivery systems for the treatment of selected tumors. International journal of nanomedicine, 12, 7291–7309. https://doi.org/10.2147/IJN.S146315.

[393] Yaqoob, A. A., Ahmad, H., Parveen, T., Ahmad, A., Oves, M., Ismail, I. M. I., Qari, H. A., Umar, K., & Mohamad Ibrahim, M. N. (2020). Recent Advances in Metal Decorated Nanomaterials and Their Various Biological Applications: A Review. Frontiers in chemistry, 8, 341. https://doi.org/10.3389/fchem.2020.00341.

[394] Murthy S. K. (2007). Nanoparticles in modern medicine: state of the art and future challenges. International journal of nanomedicine, 2(2), 129–141.

[395] Singh, J., Dutta, T., Kim, KH. et al. 'Green' synthesis of metals and their oxide nanoparticles: applications for environmental remediation. J Nanobiotechnol 16, 84 (2018). https://doi.org/10.1186/s12951-018-0408-4.

[396] Lushchak, O., Zayachkivska, A., & Vaiserman, A. (2018). Metallic Nanoantioxidants as Potential Therapeutics for Type 2 Diabetes: A Hypothetical Background and Translational Perspectives. Oxidative medicine and cellular longevity, 2018, 3407375.

https://doi.org/10.1155/2018/3407375

[397] Tillman, L., Tabish, T. A., Kamaly, N., Moss, P., El-briri, A., Thiemermann, C.,..... & Yaqoob, M. M. (2022). Advancements in nanomedicines for the detection and treatment of diabetic

kidney disease. Biomaterials and Biosystems, 100047.

[398] Ranjha, M. M. A. N., Shafique, B., Rehman, A., Mehmood, A., Ali, A., Zahra, S. M., Roobab, U., Singh, A., Ibrahim, S. A., & Siddiqui, S. A. (2022). Biocompatible Nanomaterials in Food Science, Technology, and Nutrient Drug Delivery: Recent Developments and Applications. Frontiers in nutrition, 8, 778155. https://doi.org/10.3389/fnut.2021.778155

[399] Bonsignore, G., Patrone, M., Martinotti, S., & Ranzato, E. (2021). "Green" Biomaterials: The Promising Role of Honey. Journal of functional biomaterials, 12(4), 72. https://doi.org/10.3390/jfb12040072.

[400] Bonifácio, B. V., Silva, P. B., Ramos, M. A., Negri, K. M., Bauab, T. M., & Chorilli, M. (2014). Nanotechnology-based drug delivery systems and herbal medicines: a review. International journal of nanomedicine, 9, 1–15. https://doi.org/10.2147/IJN.S52634.