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#### PHYTOCHEMICAL SCREENING AND α-GLUCOSIDASE INHIBITORS ACTIVITY OF MITRAGYNA SPECIOSA KORTH. 3 Muhammad Nivomdecha<sup>[a]</sup>, Kittisak Muandao<sup>[a]</sup>, Sucharat Sanongkiet<sup>[a]</sup> and 4 Chanjira Jaramornburapong<sup>[b]\*</sup> 5 6 7 [a]. Department of Chemistry, Faculty of Science, Silpakorn University, Nakhon Pathom, 73000, Thailand. 8 [b]. Program of Chemistry, Faculty of Science and Technology, Nakhon Pathom Rajabhat University, Nakhon 9 Pathom, 73000, Thailand. 10 11 (<sup>\*</sup>Corresponding author's e-mail: chanjira.jara@gmail.com) 12 Abstract 13 The objective of this research was to study the basic phytochemical compounds of extracts from different 14 leaves of the kratom using 5 solvents namely hexane, dichloromethane, ethyl acetate, ethanol and water. The 15 bioactive phytochemicals in red vein kratom leaves included terpenoids, saponins, tannins, flavonoids, and 16 alkaloids. And the green vein kratom leaves included terpenoids, saponins, flavonoids, and alkaloids. According to 17 the $\alpha$ -glucosidase inhibitors activity (1U/mL) using *in vitro* kratom leaf extract, it was found that the red vein kratom 18 leaves had a higher inhibitory effect on the $\alpha$ -glucosidase enzyme than the green vein kratom leaves. It was also 19 observed that ethyl acetate extract of red vein kratom with $\alpha$ -glucosidase inhibitors activity (IC<sub>50</sub> =17.28 mg/mL) 20 was similar to that of acarbose positive control ( $IC_{50} = 15.74 \text{ mg/mL}$ ) (there was a statistically significant difference 21 (p<0.5). 22 23 **Keywords:** Crude extract, phytochemicals, kratom, $\alpha$ -glucosidase inhibitors activity 24 25 **INTRODUCTION** 26 Kratom (Mitragynaspeciosa) is a perennial plant in the family Rubiaceae, native to Southeast Asia (SEA). Thailand has a long history of kratom use.<sup>1</sup>Kratom is used to listed in the Narcotic Drugs Act, Category 5, 1979, in 27 28 Thailand.<sup>2</sup>Which under Section 3 refers only to *M. speciosa*. Kratom leaves or extracts are typically eaten for pain 29 and other medical treatments, including as an aid to agricultural labor <sup>1</sup>.Kratom and kratom alkaloids are classified 30 as central nervous system (CNS) disorders.<sup>3</sup>Kratom contains the main alkaloid, mitragynine, which accounts for approximately 66% of the total alkaloids in kratom leaves <sup>4</sup>.Preliminary studies of mitragynine suggest that it may 31 be developed into an analgesic drug.<sup>4, 5</sup>Kratom has thus become a natural product that can be commercially available 32 33 over the Internet.<sup>6, 7</sup>The US Drug Enforcement Administration lists kratom as a regulated drug for use. In case studies, many Americans are using kratom to treat themselves with various conditions.<sup>8-12</sup>Although there is limited 34 35 evidence supporting the benefits of kratom in the treatment of various ailments, kratom is believed to be curative.

According to Thai medicine textbooks, Kratom leaves have properties to suppress and treat diabetes.<sup>13, 14</sup>Diabetes is 36 37 caused by abnormal secretion of the hormone insulin, resulting in hyperglycemia.<sup>15</sup>At present, the trend of diabetes is increasing rapidly, which is expected to increase to 592 million by 2035.<sup>16</sup>To treat diabetes, doctors prescribe 38 39 medications that reduce or prevent the absorption of glucose such as acarbose, myglitol, and voglibose. These drugs 40 act by inhibiting  $\alpha$ -glucosidase, an enzyme that plays a key role in catalyzing the hydrolysis of starch into glucose. 41 The absorption of glucose from the small intestine into the bloodstream is reduced. In the article titled "Does kratom 42 have medicinal properties?", which studied the wisdom and popularity of using kratom by folk healers through 43 interviews, it was found that they used kratom for medicinal purposes. The belief of indigenous healers arose from 44 the experience of healing but not the study of scientific methods. The researcher was interested in studying 45 phytochemical compounds in different leaves of kratom such as red vein kratom leaves and green vein kratom 46 leaves. In addition, a comparative study was conducted on the inhibitory activity of  $\alpha$ -glucosidase, an enzyme that 47 plays an important role in catalyzing the hydrolysis of starch into glucose of red vein kratom leaves and green vein 48 kratom leaves.

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### 50 MATERIALS AND METHODS

#### 51 Equipment and chemicals

The red vein kratom leaves and green vein kratom leaves obtained from Ratchaburiprovince, Thailand which were preliminarily examined by means of identifying kratom leaves in the control group by using kratom leaves from the same source(**Figure 1**). After that, the experiment was carried out when the initial identity was checked, starting from cleaning the kratom leaves with clean water, then drying them at 30 °C until completely dry (weighing until the weight was stable) and grinding them thoroughly.

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Red vein kratom leaves Green vein kratom leaves

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Figure 1. The red vein kratom leaves and green vein kratom leaves.

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61 The reagents consisted of 0.02M *p*-nitrophenyl- $\alpha$ -D-glucopyranoside(*p*-NPG), 0.10 M sodium phosphate 62 buffer (pH 6.8),0.10 M sodium phosphate buffer (pH 6.8) containing $\alpha$ -glucosidase (1 U/mL), 1.0 M sodium 63 carbonate(Na<sub>2</sub>CO<sub>3</sub>), 80% methanol or dimethyl sulfoxide (DMSO), crude extract of kratom leaves, acarbose

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64 (positive control), hexane, dichloromethane, ethyl acetate, ethanol, distilled water, standard laboratory glassware, EZ
 65 Read 2000 Microplate Reader.

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#### 67 Preparation of crude extract from kratom leaves

Preparation of crude extracts using solvents were as follows: hexane,dichloromethane,ethyl acetate, ethanol and distilled water.In the experiment, a volume of 200 mL of solvent was used. Initially, 20 g ofkratom leaf powder were weighed and then extracted with 200 mL of solvent per time. In each experiment, the extracted crude extract was filtered and vaporated by rotary evaporator until the solventwas completely evaporated, then the crude extract was weighed and calculated as a percentage.

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#### 74 Test of kratom phytochemical compounds <sup>17</sup>

#### 75 Anthraquinones test

About 0.2 g of crude extract was weighed, then 10 mL of 10% sulfuric acid ( $H_2SO_4$ ) solution was added and heated on a waterbath for 5 minutes, then filtered while hot. After that, let the solution cool to room temperature, add 2-3 drops of 10% ammonia( $NH_3$ ) solution and notice a reddish-pink color indicating that anthraquinone was found.

80 Terpenoids test using Salkowski test

About 0.2g of the crude extract was weighed, then added 2 mL of dichloromethane and shaken and gradually added concentrated  $H_2SO_4$ . If a reddish-brown color was formed between the junctions of the solution, then terpenoids were found.

### 84 Flavonoids test

About 0.2g of the crude extract was weighed, then the extract was dissolved using 3 mL of 50% ethanol, and 2-3 small pieces of magnesium wire were added. Later, it was boiled and dripped with concentrated hydrochloric acid (HCl). If the solution was yellow, orange, or red, flavonoids were found.

88 Saponins test

A bubble test was performed by weighing about 0.2 g of crude extract, then adding 5 mL of distilled water, bringing to a boil and filtering. Subsequently, the filtrate was added to 2-3 mL of distilled water and shaken vigorously. If bubbles were observed, saponins were found.

#### 92 Tannins test

About 0.2g of the crude extract was weighed, then added 5 mL of distilled water, heated on a water bath and
 filtered. After that, 2-3 drops of ferric chloride(FeCl<sub>3</sub>)were added to the filtered liquid. If the result was green-black
 or blue-black, then tannins were found.

#### Alkaloids test

About 0.2g of the crude extract was weighed, then dissolved in 15mL of2%H<sub>2</sub>SO<sub>4</sub> solution, warmed for 2-3
 minutes and filtered. The filtered liquid was then dripped with Dragendorff's reagent. If there was an orange-red
 precipitate, alkaloids were found.

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### 101 Method for testing theα-glucosidase inhibitors activity of crude extract from kratom leaves<sup>18</sup>

102 The test kit could be divided into 4 sets and pipetted to a 96 well-plate to the required volume as shown in

- 103 **Table 1**.
- 104
- 105 **Table 1.** Volume of  $\alpha$ -glucosidase inhibitors.
- 106

Test kit	0.1 M Sodium phosphate buffer (pH 6.8) (μL)	Test sample (µL)	1 U/mL α- glucosidase (μL)	DMSO (µL)
<b>A</b> (E+S)	100	-	20	20
<b>B</b> (S)	120	-	-	20
<b>C</b> (E+T+S)	100	20	20	-
<b>D</b> (T+S)	120	20	-	-

- 107 Note: E=Enzyme, T=Test Sample, S=Substrate.
- 108

109 All substances were mixed together and incubated at 25 °C for 15 minutes. Next,  $20\mu$ L of 0.02Mp-NPG 110 (Substrate) was added to each well and mixed well then incubated at 250°C for 5 minutes. After that,  $40\mu$ L of 1.0 M 111 Na<sub>2</sub>CO<sub>3</sub> solution was added to inactivate the reaction in each well and absorbance was measured at a wavelength of 112 405nm using an EZ Read 2000 Microplate Reader.

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# 114 Statistical data analysis

Data analysis was performed using three iterations of mean ± standard derivation, analysis of variance (ANOVA), Least Significant Difference (LSD), and Duncan Multiple Range Test (DMRT). Data correlation was searched using SPSS program.

118 The concentration of the extract that inhibits the  $\alpha$ -glucosidase enzyme at 50% (IC<sub>50</sub>) was determined using the 119 IC<sub>50</sub> Calculator program.

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## 121 RESULTS AND DISCUSSION

### 122 Amount of crude extract

According to the study on the extraction of red vein kratom leaves and green vein kratom leaves using solvents: hexane, dichloromethane,ethyl acetate, ethanol and distilled water, the results shown in **Table 2**. The percentage of crude extract of red vein kratom leaves using various solvents showed the mass of the crude as follows: Ethanol had apercentage of mass at a maximum weight of 35% and ethyl acetate; 30%, dichloromethane; 25%, hexane; 24% and distilled water had a percentage of mass at a minimum weight of 3%, respectively. The percentage of crude extract of green vein kratom leaves using various solvents showed the mass of the crudeas follows: Ethanol had a percentage of mass at a maximum weight of 22% and ethyl acetate; 8%, dichloromethane; 6%, hexane; 6% and distilled water

- had a percentage of mass at a minimum weight of 2%, respectively. All of the various solvents showed the maximum
- 131 presence in all ofred vein kratom leaves and green vein kratom leaves can soluble well in ethanol as a polar solvent.
- 132 **Table 2**. Percentage of crude extracts obtained from various solvents.

Solvents	Percentage of crude extracts obtained from various solvents (%w/w)					
	Red vein kratom leaves	Green vein kratom leaves				
Н	24	6				
D	25	6				
EA	30	8				
E	35	22				
W	3	2				

Note: H = hexane, D = dichloromethane, EA = ethyl acetate, E = ethanol, W = distilled water. The percentage of extraction as a %yield (w/w) could be calculated as (dry weight of the extract/dry weight of the plant before extraction) x 100.

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# 137 Primary phytochemical constituents of extracts from different parts of kratom using different solvents

138 Preliminary phytochemical testing of crude extracts from red vein kratom leaves using various solvents had the 139 following results: hexane contained saponins and alkaloids; dichloromethane contained saponin and alkaloids; ethyl 140 acetate contained alkaloids; ethanol contained terpenoids, tannins and alkaloids; and distilled water contained 141 terpenoids, flavonoids and alkaloids. In addition, phytochemical testing of crude extracts from the green vein kratom 142 leaves using various solvents showed the following results: hexane contained saponins and alkaloids; 143 dichloromethane contained saponins and alkaloids; ethyl acetate contained flavonoids and alkaloids; ethanol 144 contained terpenoids, saponins and alkaloids; and distilled water contained terpenoids, saponins and alkaloids as 145 shown in Table 3.

#### 146 **Table 3.** Phytochemicals found in red vein kratom leaves and green vein kratom leaves.

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	MytragynaspeciosaKorth									
Phytochemical	Red vein kratom leaves					Green vein kratom leaves				
	Н	D	EA	Ε	W	Н	D	EA	Е	W
Anthraquinones	-	-	-	-	-	-	-	-	-	-
Terpenoids	-	-	-	+	+	-	-	-	+	+
Saponins	+	+	-	-	-	+	+	-	+	+
Tannins	-	-	-	+	-	-	-	-	-	-
Flavonoids	-	-	-	-	+	-	-	+	-	+
Alkaloids	+	+	+	+	+	+	+	+	+	+

148 Note: H = hexane, D = dichloromethane, EA = ethyl acetate, E = ethanol and W = distilled water, -means not

149 detected, + means detected.

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# 151 Test on the α-glucosidase inhibitors activity of crude extract in red vein kratom leaves and green vein kratom 152 leaves

153 Mitragynine is an important and abundant substance in kratom, an alkaloid that may play a role in the  $\alpha$ -154 glucosidase inhibitors activity.<sup>19</sup>According to the phytochemical composition test, it was found that kratom leaves 155 contained alkaloids in crude extracts in all solvents. Therefore, the researcher chose the kratom leaves for extraction 156 to test the  $\alpha$ -glucosidase inhibitors activity.

157 The  $\alpha$ -glucosidase inhibitors activity of 1 U/mL in red kratom leaves had a positive control of acarbose.<sup>19</sup>In this 158 regard, there was a percentage ofα-glucosidase inhibitors activity at a minimum concentration of 0.3125 mg/mL and 159 a maximum concentration of 10 mg/mL, representing 4.39±0.79, 26.82±8.93, 33.78±0.84, 40.74±7.38, 56.51±11.38 160 and  $78.94\pm2.97$ , respectively (Figure 2). The crude extract of red vein kratom leaves using various solvents showed 161 the percentage of  $\alpha$ -glucosidase inhibitors activity as follows (**Table 4**): Hexane extract had a percentage of  $\alpha$ -162 glucosidase inhibitors activity at a minimum concentration of 2.5 mg/mL and a maximum concentration of 10 163 mg/mL, representing  $0.78\pm11.52$ ,  $5.71\pm10.82$  and  $14.53\pm8.49$ , respectively.Dichloromethane extract had a percentage 164 of a glucosidase inhibitors activity at a minimum concentration of 2.5 mg/mL and a maximum concentration of 10 165 mg/mL, representing 4.40 $\pm$ 7.73, 22.55 $\pm$ 7.19 and 48.00 $\pm$ 7.95, respectively. Ethyl acetate extract had a percentage of  $\alpha$ -166 glucosidase inhibitors activity at a minimum concentration of 0.3125 mg/mL and a maximum concentration of 10 167 mg/mL, representing 0.51±5.20, 3.27±2.24, 9.27±3.08,21.29±3.21,38.89±2.22 and 61.52±0.47, respectively. Ethanol 168 extract had a percentage of  $\alpha$ -glucosidase inhibitors activity at a minimum concentration of 1.25 mg/mL and a 169 maximum concentration of 10 mg/mL, representing  $0.29\pm2.54$ ,  $2.96\pm5.72$ ,  $12.47\pm0.56$  and  $32.39\pm0.67$ , respectively. 170 Distilled waterextract, on the other hand, showed no the  $\alpha$ -glucosidase inhibitors activity. The percentage of  $\alpha$ -171 glucosidase inhibitors activity in various crude extracts of red vein kratom leaves at maximum concentration of 10 172 mg/mLcompared with acarbose (positive control) as shown inFigure 3.

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Figure 2. Percentage of  $\alpha$ -glucosidase inhibitors activity in acarbose.

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**Table 4.**Percentage of α-glucosidase inhibitors activity in crude extract of red vein kratom leaves.

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	Perc	Percentage of $\alpha$ -glucosidase inhibitors activity in crude extract of					
Extract	red vein kratom leave (mg/mL)						
	0.3125	0.625	1.25	2.5	5	10	
н	ND	ND	ND	0.78	5.71	14.53	
	ND	ND	ND	$\pm 11.52^{aA}$	$\pm 10.82^{bA}$	$\pm 8.49^{cA}$	
D	ND	ND	ND	4.40	22.55	48.00	
D		ND	$\pm 7.73^{aA}$	$\pm 7.19^{bB}$	±7.95 <sup>cB</sup>		
ГА	0.51	3.27	9.27	21.29	38.89	61.52	
EA	$\pm 5.20^{aA}$	$\pm 2.24^{aA}$	$\pm 3.08^{bA}$	$\pm 3.21^{\text{cB}}$	$\pm 2.22^{dC}$	$\pm 0.47^{eC}$	
Е	ND	ND	0.29	2.96	12.47	32.39	
E	ND	ND	$\pm 2.54^{aB}$	$\pm 5.72^{\mathrm{aA}}$	$\pm 0.56^{bD}$	$\pm 0.67^{cD}$	
W	ND	ND	ND	ND	ND	ND	
	4.39	26.82	33.78	40.74	56.51	78.94	
Α	$\pm 0.79^{aB}$	$\pm 8.93^{bB}$	$\pm 0.84^{bC}$	$\pm 7.38^{bC}$	±11.38 <sup>cE</sup>	$\pm 2.97^{dE}$	

Note: H = hexane, D = dichloromethane, EA = ethyl acetate, E = ethanol, W = distilled water, A = acarbose, and ND means not detected, <sup>a, b, and c</sup>The horizontal mean, labeled with different letters, had a statistically significant difference(p< 0.05)., <sup>A, B, and C</sup> The vertical mean, labeled with different letters, had a statistically significant difference (p < 0.05).

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Figure 3.Percentage of α-glucosidase inhibitors activity in crude extract of red vein kratom leaves at maximum concentration of 10 mg/mL (H = hexane, D = dichloromethane, EA = ethyl acetate, E = ethanol and A = acarbose).

190	The $\alpha$ -glucosidase inhibitors activity of 1 U/mL in green kratom leaves had a positive control of acarbose. <sup>19</sup> In
191	this regard, there was a percentage of $\alpha$ -glucosidase inhibitors activity at a minimum concentration of 0.3125 mg/mL
192	and a maximum concentration of 10 mg/mL, representing 4.39±0.79, 26.82±8.93, 33.78±0.84, 40.74±7.38,
193	56.51±11.38 and 78.94±2.97, respectively(Figure 2). The crude extracts of green vein kratom leaves from various
194	solvents showed percentage of $\alpha$ -glucosidase inhibitors activity as follows( <b>Table 5</b> ): Dichloromethane extract had a
195	percentage of $\alpha$ -glucosidase inhibitors activity at a minimum concentration of 5 mg/mL and a maximum
196	concentration of 10 mg/mL, representing $2.81\pm12.16$ and $19.05\pm11.69$ , respectively. Ethyl acetate extract had a
197	percentage of $\alpha\mbox{-glucosidase}$ inhibitors activity at a minimum concentration of $0.3125 mg/mL$ and a maximum
198	concentration of 10 mg/mL, representing 1.92 $\pm$ 2.21, 6.76 $\pm$ 1.79, 14.35 $\pm$ 0.63, 23.95 $\pm$ 1.29, 33.36 $\pm$ 1.93 and 2.52 \pm
199	$42.63\pm9.08$ , respectively. The $\alpha$ -glucosidase inhibitors activity was not found inhexane, ethanol and distilled water
200	extract. The percentage of $\alpha$ -glucosidase inhibitors activity in various crude extracts of green vein kratom leaves at
201	maximum concentration of 10 mg/mLcompared with acarbose (positive control) as shown inFigure 4.

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203 **Table 5**.Percentage of α-glucosidase inhibitorsactivity in crude extract of green vein kratom leaves.

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	Percentage of $\alpha$ -glucosidase inhibitors activity in crude extract of						
Extract	green vein kratom leave (mg/mL)						
	0.3125	0.625	1.25	2.5	5	10	
Н	ND	ND	ND	ND	ND	ND	
D	ND	ND	ND	ND	2.81 ±12.16 <sup>aA</sup>	19.05 ±11.69 <sup>bA</sup>	
FA	1.92	6.76	14.35	23.95	33.36	42.63	
EA	±2.21 <sup>aA</sup>	$\pm 1.79^{abA}$	$\pm 0.63^{bA}$	±1.29 <sup>cA</sup>	$\pm 1.93^{dB}$	$\pm 9.08^{eB}$	
Ε	ND	ND	ND	ND	ND	ND	
W	ND	ND	ND	ND	ND	ND	
•	4.39	26.82	33.78	40.74	56.51	78.94	
A	$\pm 0.79^{aB}$	$\pm 8.93^{bB}$	$\pm 0.84^{bB}$	$\pm 7.38^{bB}$	±11.38 <sup>cC</sup>	$\pm 2.97^{dC}$	

Note: H = hexane, D = dichloromethane, EA = ethyl acetate, E = ethanol, W = distilled water, A = acarbose, and ND means not detected, <sup>a, b, and c</sup>The horizontal mean, labeled with different letters, had a statistically significant difference (p < 0.05), <sup>A, B, and C</sup> The vertical mean, labeled with different letters, had a statistically significant difference (p < 0.05).

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Figure 4.Percentage of  $\alpha$ -glucosidase inhibitors activity in crude extract of green vein kratom leaves at maximum concentration of 10 mg/mL (D = dichloromethane, EA = ethyl acetate and A = acarbose).

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To compare the concentrations of the extracts from the red vein kratom leaves and the green vein kratom leaves for the determination of  $IC_{50}$ , it was found that the inhibitory concentrations of the  $\alpha$ -glucosidase activity of 50% of the positive controls were as follows: Acarbose was 15.74 mg/mL. Crude extract of red vein kratom leaves with ethyl acetate was 17.28 mg/mL.<sup>19</sup>Crude extract of ethanol was more 50 mg/mL. Green vein kratom leaves with 50 percent of  $\alpha$ -glucosidase inhibitor activity were as follows: Crude extracts of ethyl acetate wasmore50 mg/mL as shown in **Table 6**.

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**Table 6.** Concentrations of  $\alpha$ -glucosidase inhibitors extracts at 50% (IC<sub>50</sub>).

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Extracts	Concentrations of a-glucosidase inhibitors extracts at 50% (IC <sub>50</sub> ) (mg/mL)					
	Red vein kratom leaves	Green vein kratom leaves				
Н	ND	ND				
D	ND	ND				
EA	17.28 <sup>B</sup>	>50 <sup>C</sup>				
E	>50 <sup>C</sup>	ND				
W	ND	ND				
Α	15.74 <sup>A</sup>					

Note: H = hexane, D = dichloromethane, EA = ethyl acetate, E = ethanol, W = distilled water, A = acarbose, and ND means not detected, <sup>A, B, and C</sup>The vertical mean, labeled with different letters, had a statistically significant difference(p < 0.05).

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## 228 CONCLUSIONS

According to preliminary phytochemical testing of extracts from different leaf of the kratom using five solvents: hexane, dichloromethane, ethyl acetate, ethanol and distilled water. Details could be classified as follows. The red vein kratom leaves contained bioactive phytochemicals including terpenoids, saponins, tannins, flavonoids,

and alkaloids, but no anthraquinones. The green vein kratom leaves contained bioactive phytochemicals such asterpenoids, saponins, flavonoids, and alkaloids, but no anthraquinones and tannins.

According to the  $\alpha$ -glucosidase inhibitors activity (1 U/mL) using *in vitro* kratom leaf extract, it was found that the red vein kratom leaves had a higher inhibitory effect on the  $\alpha$ -glucosidase enzyme than the green vein kratom leaves. It was also observed that ethyl acetate extract of red vein kratom with  $\alpha$ -glucosidase inhibitors activity (IC<sub>50</sub>=17.28 mg/mL) was similar to that of acarbose positive control (IC<sub>50</sub>=15.74 mg/mL) (there was a statistically significant difference (p<0.05).

However, a detailed study of the important phytochemicals of crude extracts and their bioactivity is interestingfor further experiments.

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#### 242 ACKNOWLEDGEMENTS

The researcher would like to thank the research subsidies for the publication of research results in academic journals from the Innovation and Creativity Research Support Fund of the Faculty of Science, Silpakorn University,

- 245 grant number SRIF-JRG-2562-08.
- 246

#### 247 **REFERENCE**

- Hassan, Z.,Muzaimi, M.,Navaratnam, V.,Yusoff,N.H.M.,Suhaimi, F.W., Vadivelu, R.,Vicknasingam, B.K.,
   Amato,D.,Hörsten, S., Ismail,N.I.W.,Jayabalan, N.,Hazim, A.I.,Mansor, S.M., Müller, C.P.,*Neurosci. Biobehav. Rev.*, 2013,37 (2),138-151.
- Office of the NACC, Ministry of Justice, Collection of drug laws, ministerial regulations and related
   regulations. Bangkok: Bangkok Block Limited Partnership, 2013.
- 253 3. Raffa, R.B., Pergolizzi, J.V., Taylor, R., Ossipov, M.H., J. Clin. Pharm. Ther., 2018, 43 (3), 437-441.
- Kruegel,A.C.,Gassaway, M.M.,Kapoor, A.,Váradi, A.,Majumdar, S.,Filizola, M.,Javitch, J.A.,Sames.D.,*J. Am. Chem. Soc.*,2016,138 (21),6754-6764.
- 256 5. Macko, E., Weisbach, J.A., Douglas, B., Arch. Int. Pharmacodyn. Ther., 1972, 198 (1), 145-161.
- 257 6. Babu, K.M., McCurdy, C.R., Boyer, E.W., *Clin. Toxicol.*, **2008**, *46* (2), 146-152.
- 258 7. Prozialeck, W.C., J. Am. Osteopath. Assoc., 2016, 116 (12), 802-809.
- 259 8. Boyer, E.W., Babu, K.M., Adkins, J.E., McCurdy, C.R., Halpern, J.H., Addiction, 2008, 103 (6), 1048-1050.
- 260 9. Coe,M.A.,Pillitteri, J.L.,Sembower, M.A.,Gerlach, K.K., Henningfield,J.E., *Drug and Alcohol*261 *Dependence*,2019,202,24-32.
- 262 10. Grundmann, O., Drug and Alcohol Dependence, 2017, 176, 63-70.
- 263 11. Henningfield, J.E., Fant, R.V., Wang, D.W., Psychopharmacology, 2018, 235 (2), 573-589.
- 264 12. Smith, K.E., Lawson, T., Drugand AlcoholDependence, 2017, 180, 340-348.
- Bannalak,S.(Kittikachorn, A.) Scriptures of Traditional Thai Medicine, Volume 1, (The Ancient Thai
   Pharmacy Association of Thailand)

- Asanangkornchai,S.,Siriwong Na Ayudhya, A., Kratom leaves in Thai society, Office of the Narcotics Control
   Board. Ministry of Justice, Bangkok Block Limited Partnership, Bangkok, 2005.
- 269 15. Deutschlander, M.S., Venter, M., Roux, S., Louw, J., Lall, N., Journal of Ethnopharmacology, **2009**, 124(3), 619-624.
- 270 16. Kim,C.U.,Lew, W., Williams, M.A., Wu, H.,Zhang, L., Chen, X., Escarpe, P.A., Mendel, D.B., Laver, W.G.,
  271 Steven,R.C.,*J. Med. Chem.*,1998,41,2451-2460.
- 17. Harborne, J.B., *TextbookofPhytochemicalMethods.* A Guideto Modern TechniquesofPlantAnalysis.
  273 3<sup>rd</sup>ed., Chapman and Hall Ltd, London, **1998**, p.182-190.
- 274 18. Field, E., J. Chem. Soc., Trans., **1921**, 119, 887-891.
- 275 19. Niyomdecha, M., Muandao, K., Kuttiyod, T., Sanongkiet, S., *International Journal of Health*276 Sciences, 2022, 6(S3), 10254-10261.