



## **Evaluation of Mineralized Plasmatic Matrix Prepared from Xenogeneic Bone in Mandibular Defects Grafting after Cystectomy in Diabetic Patient: A Prospective Randomized Clinical Trial**

**Mohamed Salah Mohamed Mohamed**

*Lecturer of Oral and Maxillofacial Surgery, Department of Oral & Maxillofacial Surgery, Faculty of Dentistry, Aswan University.*

**Corresponding author:** Mohamed Salah Mohamed Mohamed

[Dr.mdsalah86@gmail.com](mailto:Dr.mdsalah86@gmail.com)

---

**Received:16-6-2023**

**Accepted:30-6-2023**

**Published:30-7-2023**

---

**ABSTRACT: Objective:** To evaluate the usage of Mineralized Plasmatic Matrix prepared from Xenogeneic Bone (MPM-XB) in the healing process of mandibular defects after cystectomy in diabetic patients by using density measuring and volumetric analysis with cone beam computed tomography (CBCT). **Patients and Methods:** This study involved 12 diabetic cases with cystic lesions of the mandible. MPM-XB is used to fill the cavity after cystectomy. At the post-operative 3<sup>rd</sup> and 6<sup>th</sup> months, the implanted bone was evaluated by matching the CBCT scan in terms of density and volume compared with the mandibular cystic lesion volume. **Results:** Mean bone density was increased after 6 months as compared to 3 months postoperative with statistically significant difference p value  $\leq 0.05$ . Bone density increased by mean value  $41.75 \pm 20.43$ , ranged from 18 to 102, while volumetric analysis showed a non-significant difference between preoperative and 6 months after, regarding cystic volume and graft volume as well as the ratio between both ( $P > 0.05$ ). **Conclusion:** Mineralized plasmatic matrix prepared from xenogeneic bone appeared to be stable with appropriate bone density and minimal complications in filling bone defects after cystectomy in diabetic patients with mandibular cystic lesions.

**Keywords:** Mineralized Plasmatic Matrix, xenogeneic graft, cystic mandibular lesion.

---

**DOI: 10.48047/ecb/2023.12.Si8.777**

### **Introduction:**

Healing of the bone is a sequence of complex orchestrated biologic processes. Bone healing includes osteo-induction and osteoconduction mediated by intra-cellular and extra-cellular signaling pathways [1]. A bone defect can be congenital or related to infection, trauma or malignancy [2]. Usually, spontaneous healing of small bone defect does occur; however, in critical bone defect, there is a limited healing capacity; thus, surgery is necessary [3].

Large sized jaw defect might cause significant deformities of the face, as well as lingual and masticatory dysfunction. Accordingly, their reconstruction and repair are important issues for plastic and maxillofacial surgeons [4,5,6]. Bone grafting thus has an important role and is necessary for reconstructing the massive defect [7]. Autograft, allograft, xenograft, and synthetic bone graft substitute material (alloplastic) have been used [8].

A xenograft involves grafting from an animal to another of other species or to human [9]. Osteogenesis, osteoconduction, osteo-induction, and osteointegration are biologic processes essential for new bone formation [10]. There are limitations related to the utilization of autograft and allograft [10,11]. Both xenografts and tissue engineering material with enhanced properties could overcome such limitations [12]. Bone-derived xenografts are extensively utilized in maxillofacial surgery [13].

MPM is significantly concentrated in platelets and fibrin in liquid form and combined with bone powder or bone substitute [11]. MPM is a variant of platelet-rich fibrin [14]. Platelet concentrates are appropriate bioactive material due to providing growth factors and cytokines that help in healing process [15,16]. Furthermore, fibrin promotes migration and proliferation of the cells as well as angiogenesis [17]. MPM is more advantageous than platelet concentrates due to incorporation of graft particles into the fibrin meshwork, thus forming sticky bone. The latter is flexible and easy to shape, and it provides high stability to bone particles and prevents soft tissue deposition in the defect [18].

Diabetes mellitus (DM) is common in dental patients. Hyperglycemic microenvironment can delay the healing of post-surgical jaw defects, and is associated with poor outcomes in many oral diseases [19]. Novel improvements have been made in various treatment strategies and medications for managing wound healing in diabetes. Topical plasma-rich growth factor, platelet-rich fibrin, leukocyte- and platelet-rich fibrin and hyaluronic acid is regarded as a promising strategy for diabetics who require jaw surgeries [19].

This study aimed at evaluating the usage of MPM-XB in healing process of mandibular defects after cystectomy in diabetic patients by using volumetric analysis and density measuring with CBCT.

## **Patients and Methods**

This prospective, interventional study involved 12 cases with mandibular cyst in succession who approved to be studied and fulfilled the inclusion criteria. The study was performed in the Dental clinics of Aswan University Hospital at the period from April 2022 to May 2023.

The study included adult diabetic patients more than 18 years of age, both sexes included, they had mandibular cystic bone lesions (large cysts >2.5 cm). Other forms of jaw diseases, pediatric patients, previous dental surgery and major systemic diseases or autoimmune diseases as well as non-diabetic patients were excluded.

The procedures were performed according to ethics research committee, Faculty of oral and dental medicine, Cairo University. Informed consents were taken from all participants and included details about the purpose of this study, and a simple and easy explanation of the technique, possible complications, and benefits of the procedure.

### **Preoperative evaluation:**

Detailed medical and dental history was obtained from each patient including name, age, gender, occupation, address, telephone number.

General examination of systemic condition, blood pressure, temperature and a local examination including jaw inspection and palpation were performed to detect any accompanying infections or lesions.

Laboratory report including complete blood count (CBC), hemoglobin concentration, coagulation study, blood sugar level as well as glycosylated haemoglobin (HbA1c).

Plain X-ray of the jaw (panoramic views) and CBCT (PaX-i3D, Vatech Co., Gyeonggi-do, Republic of Korea) utilizing the following settings: 0.3-mm, 24 s, 106 kV, and 65 mAs was done for all patients for assessment of exact location, size, and extension of jaw cystic lesion.

A sample was taken from the cystic lesion to ensure its histopathological nature before performing its complete surgical removal by using fine-needle aspiration biopsy.

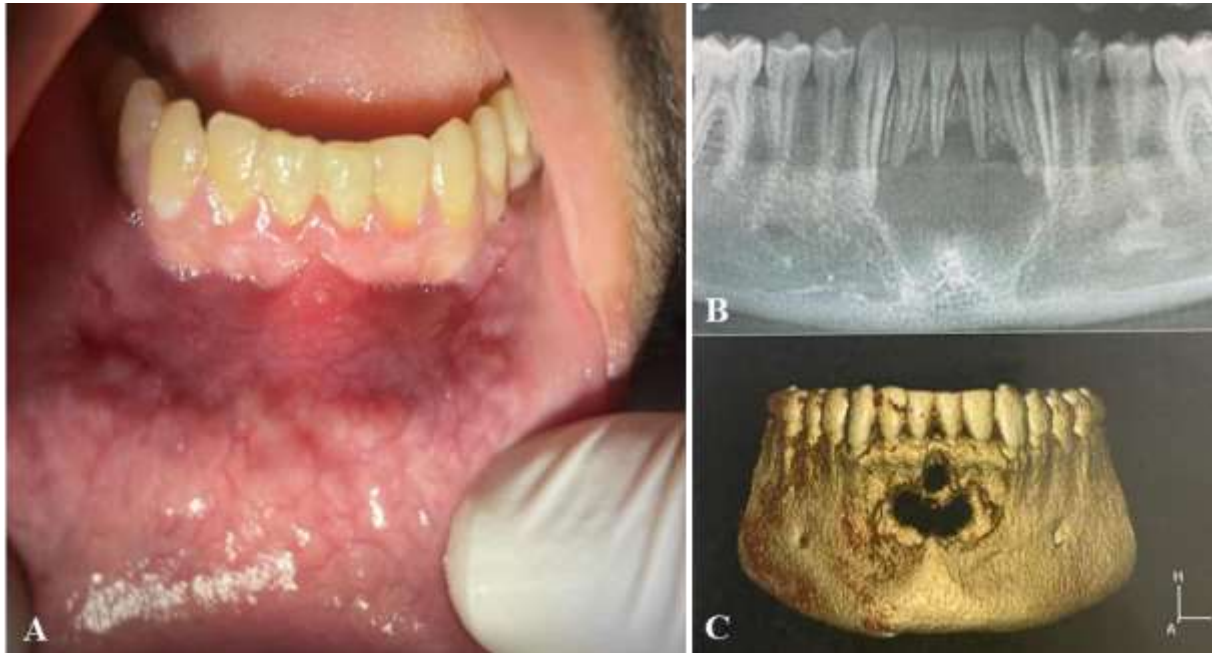


Fig. (1): Preoperative views of mandibular lesion: A. Direct photo. B. Plain x-ray. C. 3D radiographic view.

### **Operative procedures:**

Pre-surgical rubbing and preparation were achieved. Articain (ARTINIBSA 4% 1:100000®)<sup>1</sup> was first injected into the surgical sites. A full 3-lines mucoperiosteal flap was incised and reflected for exposure of the area of cystic lesion. Periosteal releasing incisions were then performed to enable tension-free closure at the end of operation. Small cortical perforations were performed for accessing the lesion if decortication was needed using small round bur. Cysts were excised with adequate curettage and were planned for microscopic examination. Any related restorable teeth were endodontically obturated after apicectomy.

<sup>1</sup> ARTINIBSA® 4% 1:100000 by INIBSA Pharmaceutical Group, Spain.

<sup>2</sup> Xenograft ONEXENO GRAFT® by One Graft, Germany.

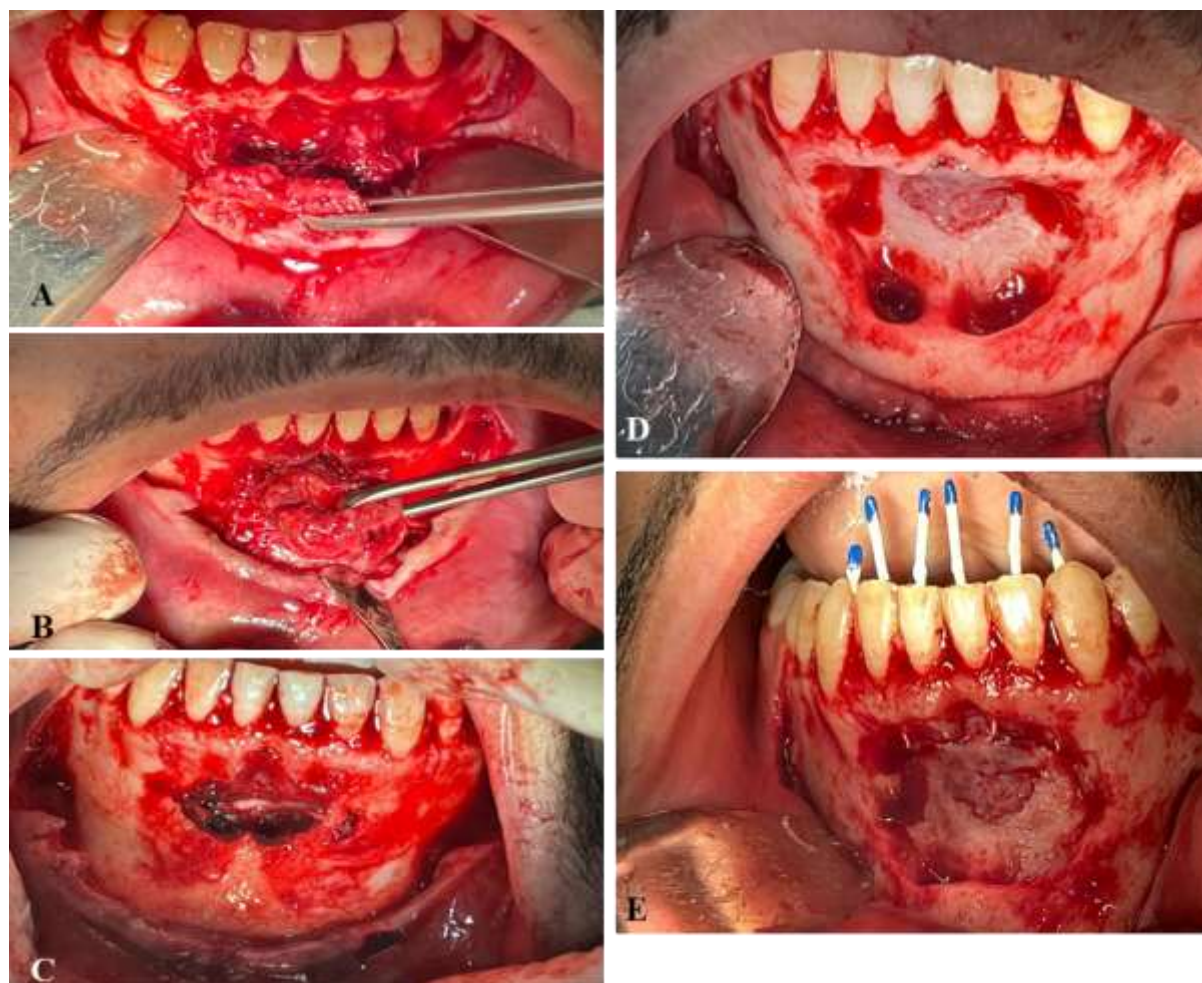


Fig. (2): Steps of cyst removal, curettage, RCT and graft site preparation.

A Xenogenic cancellous bone particulate was utilized for grafting (OneXeno Graft, Cortico-cancellous Bovine Powder, Germany). It was placed in a sterile mixing bowl and underwent hydration with saline. From the patient, 15ml venous blood was collected in uncoated plastic tubes without anticoagulant (B.D. vacutainer) and underwent centrifugation at (2500 rpm) over three minutes to separate the plasma which is rich in platelets and fibrin concentrates at top of the tube while the red corpuscles at its bottom. Then, plasma collection was done using a syringe and placed over the graft and stirred with a mucoperiosteal elevator until the formation of a bone-fibrin mass.

The MPM was adapted to the labial surface of the alveolar bone, then the flap was repositioned and closed using 4-0 Vicryl sutures.



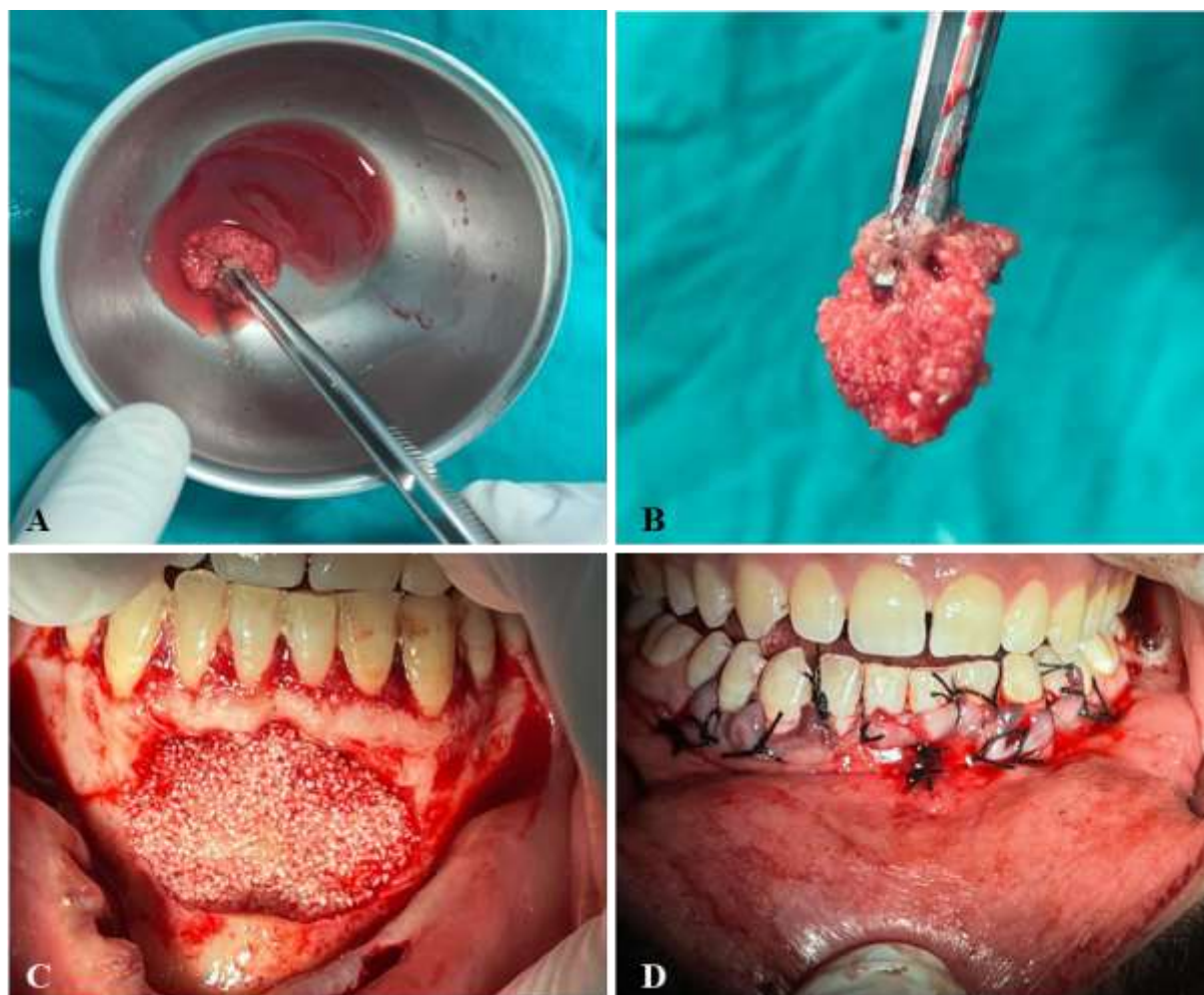


Fig. (3): A & B. Mineralized Plasmatic Matrix Preparation from Xenogeneic Bone. C. Graft insertion in bone defect. D. Wound suturing.

The excised biopsy was sent to pathology department for pathological differentiation. Antibiotics (Amoxicillin-clavulanate) were prescribed (Augmentin 1gm, SmithKline Beecham, Egypt) two times daily for each patient to prevent infections. An analgesic (Brufen 400mg, Abbot Pharmaceuticals, Egypt)<sup>2</sup> was also prescribed to relieve pain.

---

Augmentin 1gm, SmithKline Beecham, Egypt.

<sup>2</sup> Brufen 400mg, Abbot Pharmaceuticals, Egypt.

### **Postoperative follow-up:**

All patients were assessed in the 1<sup>st</sup> post-operative day looking to exclude any complication and to confirm the proper use of post-operative prescribed drugs. Patients wound was observed one week post-operatively. Then, the follow up was after 1 and 3 months.

CBCT-scans for density measuring and volumetric analysis were carried out and underwent reviewing after 3 and 6 months in relation with the preoperative CBCT-scans.

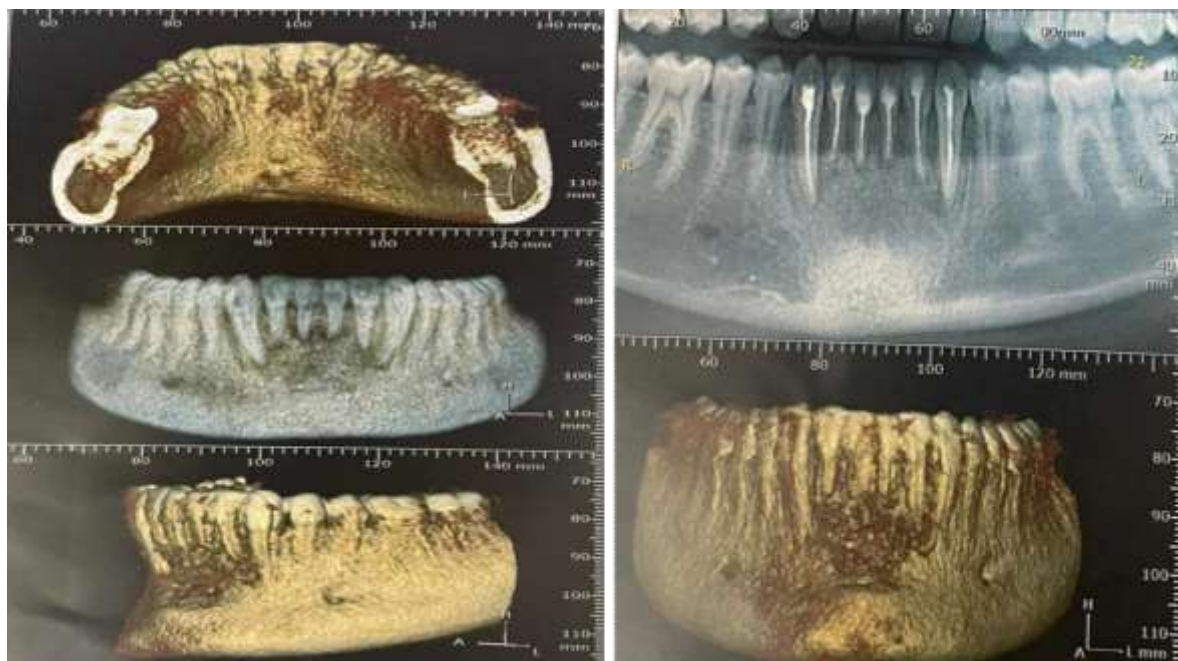


Fig. (4): Cone beam computed tomography (CBCT) during follow-up.

3D volume measurement method consisted of using the INVESALIUS software. A 3D surface of the graft was created from a 2D mask which contains all the pixels of graft and defect. This 2D mask was obtained by customized thresholding tool and then using manual segmentation tool adjusted in all orthogonal plans by interpolation and filling hole of the software, then finally mask was translated into a 3D surface model with volume in mm<sup>3</sup>.

Alterations in buccal and lingual bone density of the cystectomy defect (relative) were evaluated from the coronal sagittal cuts whereas the mesial and distal alterations were evaluated from the panoramic cuts using the density measurement tool to determine the region of interest (ROI) then the software automatically calculated bone density. Relative bone density was measured then the average bone density was calculated [20].

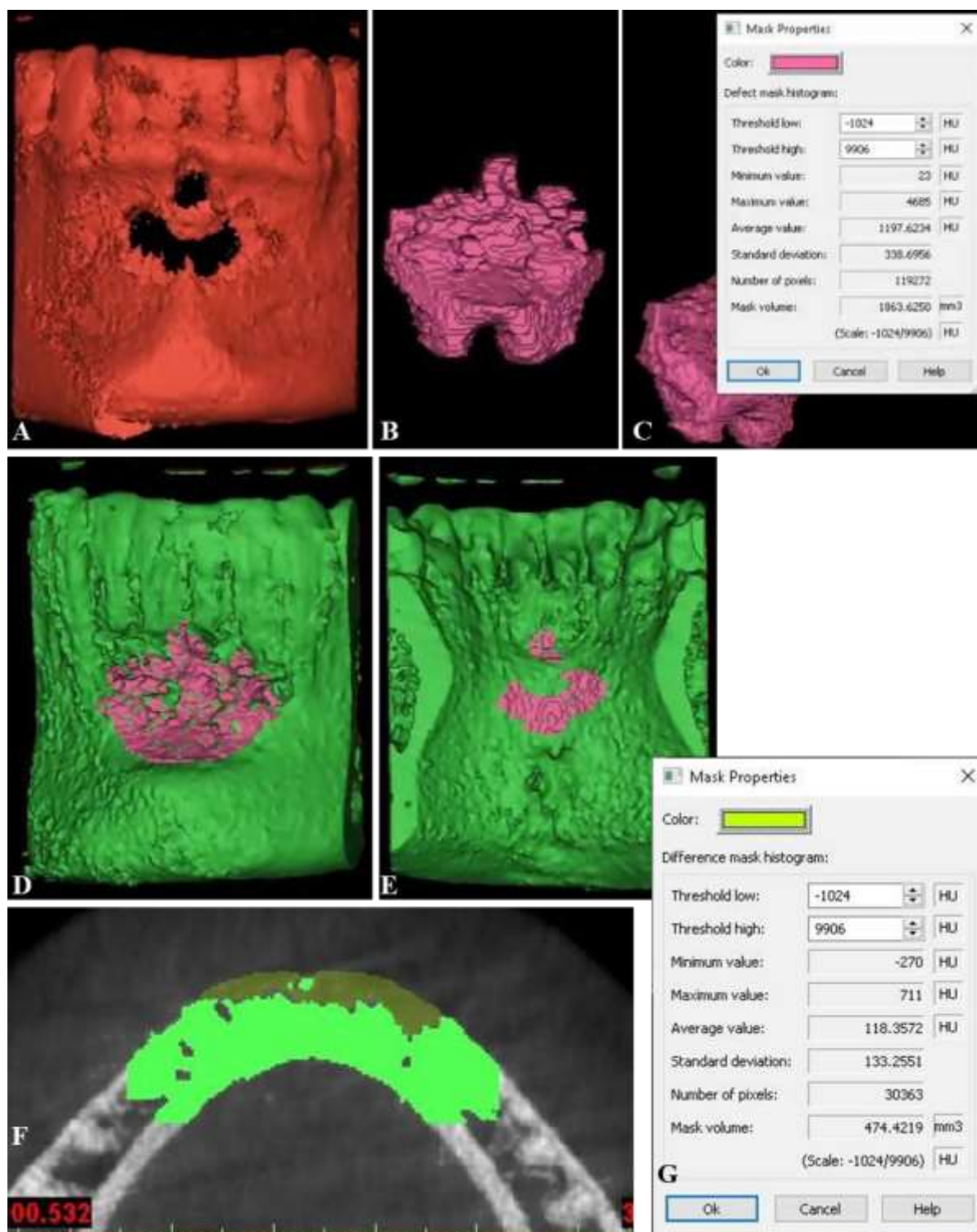


Fig. (5): Examples of X-ray volumetric analysis.

### Statistical analysis:

Data were analysed by IBM SPSS software package v 25 (Armonk, NY: IBM Corp). Frequencies and percents were utilized for qualitative data. The Kolmogorov-Smirnov test was utilized to confirm the normality of distribution; the paired groups were compared with paired



t test. Ranges (minimum and maximum), means and standard deviations (SDs) were utilized for quantitative data. Significance of a result was set at  $p \leq 0.05$ .

## Results

This study included 12 diabetic patients with mandibular cystic lesion: They were 5 males (41.7%) and 7 females (58.3%). The ages ranged between 31 to 65 years with mean  $\pm$  SD of  $52.3 \pm 6.42$  years. Patients' characteristics are presented in (table1).

Mean bone density was increased after 6 months as compared to 3 months postoperative with statistically significant difference p value  $\leq 0.05$ . Bone density increased by mean value  $41.75 \pm 20.43$ , ranged from 18 to 102 (table2), while volumetric analysis shows there a non-significant difference between preoperative and 6 months after, as regard cystic volume and graft volume as well as the ratio between both ( $P > 0.05$ ) (table 3).

Postoperative complications were minimal. Pain was significantly decreased ( $p < 0.001$ ) from  $4.12 \pm 0.79$  by VAS during the first week decreased to  $1.09 \pm 0.04$  in the first month, then disappeared at 3 months of follow-up. Also, cheek edema significantly decreased ( $p = 0.001$ ) from  $14.7 \pm 3.17$  in the first postoperative week to  $7.75 \pm 7.51$  at one month and disappeared at the 3<sup>rd</sup> month. No other complications were found in the follow-up period (table 4).

Table (1): Preoperative patients' criteria of the study population.

	Males		Females		Significance	
	No.	%	No.	%	$\chi^2$	P
Total (n = 12)	5	41.7	7	58.3	0.192	0.542
	Anterior mandibular cysts		Posterior mandibular cysts			
Site of lesion	10	83.3	2	16.7	12.37	0.000*
	Range		Mean $\pm$ SD			
Age (years)	31 – 65		$52.3 \pm 6.42$			
systolic blood pressure (mmHg)	102 – 157		$135.9 \pm 13.4$			
diastolic blood pressure (mmHg)	59 – 92		$78.8 \pm 8.65$			
Fasting blood sugar (mg/dL)	118 – 151		$127.6 \pm 21.4$			
postprandial blood sugar (mg/dL)	135 – 232		$176.5 \pm 42.7$			
HbA1c (U/L)	5.5 – 13.1		$8.63 \pm 3.41$			

$\chi^2$  = Chi square,  $P > 0.05$ : non-significant. \* $p < 0.001$ : highly significant.

Table (2): Bone density measuring outcome during the follow-up period.

Density Measuring	After 3 months (n=12)	After 6 months (n=12)	T6-T3	Paired t-test	P-value
Bone density Mean $\pm$ SD	$852.42 \pm 96.91$ 712-1095	$894.17 \pm 112.7$ 750-1197	$41.75 \pm 20.43$ 18-102	t = 7.08	$\leq 0.001^*$

Mean bone density was increased after 6 months as compared to 3 months postoperative with statistically significant difference p value  $\leq 0.05$ . Bone density increased by mean value  $41.75 \pm 20.43$ , ranged from 18 to 102.

Table (3): Volumetric analysis outcome during follow-up period.

Outcome	Values	Significance
---------	--------	--------------



		F	P
Cystic volume (mm <sup>3</sup> )	27.91 ± 9.97	0.364	0.494
Graft volume (mm <sup>3</sup> )	21.52 ± 6.42	0.398	0.463
Graft ratio (%)	85.6 ± 14.2	0.221	0.185

F: Friedman test, \* p <0.05 = significant.

This table showed that the cystic volume was 27.91 ± 9.97, while the graft volume was 21.52 ± 6.42. The mean graft ratio was 85.6 ± 14.2%.

Table (4): Postoperative complications during the follow-up period.

Complication	1 week	1 month	3 months	Significance	
				F	P
Pain (VAS)	4.12 ± 0.89	1.09 ± 0.04	0.00	21.6	0.000*
Cheek edema	14.7 ± 3.17	7.75 ± 7.51	0.00	15.73	0.001*
Infection	0.00	0.00	0.00	-	-
Graft exposure	0.00	0.00	0.00	-	-
Graft rejection	0.00	0.00	0.00	-	-
Dehiscence	0.00	0.00	0.00	-	-

F: Friedman test, \* p <0.05 = significant. VAS: visual analogue score.

## Discussion

Cystectomy is the standard technique for treating bone cysts and might be performed in combination with decompression [21]. After cystectomy, healing does occur spontaneously to repair bone defects [22]. However, decompression and enucleation are not always adequate for achieving complete bone regeneration, and thus healing time could be extended. This increases the risk of fractures and infections.

Spontaneous bone regeneration requires an adequate blood supply and mesenchymal cells. But, because of the absence of mechanical support, the massive defect cannot completely heal [23,24]. In agreement with our study, another study by Perić Kačarević Ž et al. stated that under previously mentioned conditions of critical sized defects, external materials are required for the regeneration process [25].

Different bone substitute materials have been generally utilized [26]. However, there is little evidence to support certain treatment or which material to be utilized. Bovine-derived hydroxyapatite and synthetic hydroxyapatite were found to be associated with optimal healing over six months [27]. Other supernatants, like plasma-rich gel, have also been found to have efficacy [28].

Autologous platelet concentrates contain growth factors, which promote healing. These growth factors include platelet derived growth factor, tumor growth factor beta, vascular endothelial growth factor, and other cytokines. For that, autologous platelet concentrates are commonly used in conditions requiring rapid tissue repair and regeneration and they have shown positive outcomes. The autologous platelet concentrates are applied in the form of fibrin network will lead to confinement of growth factors secretion to the ROI [29].

MPM preparation features the simplicity of the PRF protocol, but provides a liquid platelet/fibrin concentrate which can bind to bony particles. Scanning electron microscopy (SEM) shows that MPM forms a fibrin meshwork around the mineral blocks. Bone graft can

be readily conformed and the surgical site refreshed by different contained products. The surgical technique is often not changed and it becomes easier and safer. MPM involves the usage of plastic tubes with no additives, thus initiating the intrinsic coagulation pathway. The high  $\text{Ca}^{+2}$  and thromboplastin levels in plasma induce the extrinsic coagulation pathway. Usefully, a homogeneous filling material, a fibrin membrane, and the valuable biological properties of PRF become simultaneously available [30].

For bone grafting success, many conditions must be ensured including the space maintaining, the scaffolding, graft's stability and the appropriate closure. Platelets' concentration has a little role in the regeneration process as their lifetime is only 4 - 8 days. Thus, few days following the placement of platelet concentrates, their concentration becomes decreased significantly reaching the normal concentration in the body. So, what must be considered in such products is neither the biologic portion, nor is the contained cells, but the significant consideration is the biomechanical part. The MPM is the only natural and autogenous product which offers stability to bony particles. The PRF alone or when mixed with a bone graft, will not achieve adequate stability or the required resistance to chewing forces, and thus is not helpful for bone regeneration [31]

Diabetics have a high risk of hyperlipidaemia, infections, and delayed tissue healing. DM is the 3<sup>rd</sup> most prevalent chronic oral disease [32]. In contrast, hyperglycaemia decrease the concentrations of insulin growth factor-1 (IGF-1), transforming growth factor- $\beta$ 3, epidermal growth factor receptor, and ciliary neurotrophic factor, contributing to poor healing [33].

Delayed tooth extraction socket healing is usually observed in diabetic subjects [33]. Oral wound healing is slower in diabetic cases compared with those without DM, especially on the post-operative 7<sup>th</sup> day [34]. However, not all publications concluded that diabetic patients have increased healing disorders [35].

To avoid longstanding healing procedures and their subsequent complications we used in our study MPM-XB to fill the cavity after cystectomy in 12 diabetic patients with Mandibular jaw cyst of critical size aiming to spare autografting with similar outcome and less complications.

Feichtinger and co-workers had recommended the use of 3D CT to obtain specific results on the volume and width of bone bridge [36].

Bornstein et al. concluded that CBCT could be utilized in dentistry for preoperative anatomical assessment, site design, and planning of treatment, as well as for post-operative evaluation [37]. Also, it was found that CBCT is more advantageous than CT because of less radiation dose and less costs [38].

Osman et al. (2019) study shows that cone beam CT scoring method for assessing the outcomes of alveolar bone graft was an excellent radiographic evaluation. Moreover, CBCT is accurate in terms of quantitative analysis of buccal and lingual alveolar bone thickness at different vertical levels [39].

So CBCT was used in our study instead of conventional CT pre-operatively and postoperatively. Regarding bone density using CBCT, a significant increase in bone density was found after 3 months up to the maximum at 6 months, while volumetric analysis shows that there had a non-significant difference between preoperative and 6 months after, regarding cystic volume and graft volume as well as the ratio between both. This is in accordance with

Abo Serie et al., Sultan et al. [40] and Elbokle et al. [41], who concluded that MPM achieved positive results in terms of bone density. Also, Cinar et al. [42] reported that MPM increased new bone formation.

Other earlier studies assessed the grafted bone alterations following alveolar bone graft through assessing the height, Labio- Lingual Thickness, and volume of the grafted bone pre-grafting in comparison to 3 months and one year after grafting [36]. Other studies assessed the resorption of alveolar bone graft through measuring the graft volume at 30 days and 6 months [43].

In our study, regarding the time of healing we had favorable outcomes as we found MPM-XB can fill the cystectomy cavity with good healing and less complications in diabetic patients. The same results were addressed by Abo Serie et al. [44] who studied 16 cases with anterior maxillary horizontal alveolar defect and concluded that MPM provided a more durable and stable structure which helped new bone formation without utilizing a covering collagen membrane.

Our results show that pain and edema were reduced markedly in the 1<sup>st</sup> month and disappeared by the 3<sup>rd</sup> month. This agrees with Mansour et al. [38] who reported that MPM reduces post-operative infections, oedema, pain, Graft exposure and/or loss and soft tissue dehiscence. Pain in the first month was controlled by analgesics. No infection found in our study as we adequately controlled diabetes with strict sterile condition pre- intra- and postoperatively with systemic broad-spectrum antibiotics. Also, we did not find any graft exposure and soft tissue dehiscence.

## **Conclusion**

For bone grafting success, many conditions must be fulfilled including the space maintaining, scaffolding, graft's stability and the appropriate closure. MPM prepared from xenogeneic bone is significantly effective in restoring critical sized mandibular cystectomy defects in diabetic patients with ease and less complications. Moreover, work with more patients, however, is essential, and the biological qualities of MPM should be appropriately defined.

**Conflict of interests:** None.

## **References:**

1. Dimitriou R, Jones E, McGonagle D, Giannoudis PV. Bone regeneration: Current concepts and future directions. *BMC Med.* 2011; 9: 66.
2. Holban AM, Grumezescu A. *Materials for Biomedical Engineering: Hydrogels and Polymer-Based Scaffolds*; Elsevier: Amsterdam, The Netherlands, 2019.
3. Oryan A, Alidadi S, Moshiri A, Maffulli N. Bone regenerative medicine: Classic options, novel strategies, and future directions. *J. Orthop. Surg. Res.* 2014; 9: 18.
4. Wu J, Liu J, Wang L, Xie A, Liu D. Bone histomorphometry detection of autologous bone powder graft repair of partial mandibular defects in rabbits. *Genet. Mol. Res.* 2015; 14: 13812–13822.



5. Kumar BP, Venkatesh V, Kumar K, Yadav BY, Mohan SR. Mandibular reconstruction: Overview. *J. Maxillofac. Oral Surg.* 2016; 15: 425–441.
6. Zhang Q, Wu W, Qian C, Xiao W, Zhu H, Guo J, Meng Z, Zhu J, Ge Z, Cui W. Advanced biomaterials for repairing and reconstruction of mandibular defects. *Mater. Sci. Eng. C* 2019; 103: 109858.
7. Saikia K, Bhattacharya T, Bhuyan S, Talukdar D, Saikia S, Jitesh P. Calcium phosphate ceramics as bone graft substitutes in filling bone tumor defects. *Indian J. Orthop.* 2008; 42: 169.
8. Bauer TW, Muschler GF. Bone graft materials: An overview of the basic science. *Clin. Orthop. Relat. Res.* 2000; 371: 10–27.
9. Campana V, Milano G, Pagano E, Barba M, Cicione C, Salonna G, Lattanzi W, Logroscino G. Bone substitutes in orthopaedic surgery: From basic science to clinical practice. *J. Mater. Sci. Mater. Med.* 2014; 25: 2445–2461.
10. Zamiri B, Shahidi S, Eslaminejad MB, Khoshzaban A, Gholami M, Bahramnejad E, Moghadasali R, Mardpour S, Aghdami N. Reconstruction of human mandibular continuity defects with allogenic scaffold and autologous marrow mesenchymal stem cells. *J. Craniofacial Surg.* 2013; 24: 1292–1297.
11. Moshiri A, Shahrezaee M, Shekarchi B, Oryan A, Azma K. Three-dimensional porous gelatin–simvastatin scaffolds promoted bone defect healing in rabbits. *Calcif. Tissue Int.* 2015; 96: 552–564.
12. Saravanan S, Leena R, Selvamurugan N. Chitosan based biocomposite scaffolds for bone tissue engineering. *Int. J. Biol. Macromol.* 2016; 93: 1354–1365.
13. Soares MQS, Van Dessel J, Jacobs R, Yaedú RYF, Sant’Ana E, da Silva Corrêa D, Madeira MFC, Duarte MAH, Rubira-Bullen IRF. Morphometric evaluation of bone regeneration in segmental mandibular bone defects filled with bovine bone xenografts in a split-mouth rabbit model. *Int. J. Implant Dent.* 2019; 5: 32.
14. Mohamed EM. The use of growth factors fibrin network to enhance architecture, mechanical and biological aspect of the graft particles. *Int. J. Prev. Clin. Dent. Res.* 2014; 1: 41–44.
15. Mudalal M, Zhou Y. Biological additives and platelet concentrates for tissue engineering on regenerative dentistry basic Science and concise review. *Asian J. Pharm.* 2017; 11: 255–263.
16. Weibrich G, Kleis WK, Hafner G. Growth factor levels in the platelet-rich plasma produced by 2 different methods: Curasantype PRP kit versus PCCS PRP system. *Int. J. Oral Maxillofac. Implant.* 2002; 17: 184–190.
17. Mehrabani D, Khodakaram-Tafti A, Shaterzadeh-Yazdi H, Zamiri B, Omid M. Comparison of the regenerative effect of adipose-derived stem cells, fibrin glue scaffold, and autologous bone graft in experimental mandibular defect in rabbit. *Dent. Traumatol.* 2018; 34: 413–420.

18. Gonzalez-Sanchez JG and Jimenez-Barragan K. Closure of recurrent cleft palate fistulas with plasma rich in growth factors. *Acta Otorrinolaringol English Edition*) 2011; 62:448–453.
19. Yang S, Li Y, Liu C, Wu Y, Wan Z, Shen D. Pathogenesis and treatment of wound healing in patients with diabetes after tooth extraction. *Front Endocrinol (Lausanne)*. 2022 Sep 23; 13: 949535. doi: 10.3389/fendo.2022.949535.
20. Silva IMCC, Freitas DQ, Ambrosano GMB, Bóscolo FN, Almeida SM. Bone density: comparative evaluation of Hounsfield units in multislice and cone-beam computed tomography, *Braz Oral Res.*, 2012; 26: 550-6.
21. .Rajendra Santosh AB. Odontogenic Cysts. *Dent Clin North Am*. 2020;64:105–119. [PubMed] [Google Scholar]
22. Perjuci F, Ademi-Abdyli R, Abdyli Y, Morina E, Gashi A, Agani Z, Ahmedi J. Evaluation of spontaneous bone healing after enucleation of large residual cyst in maxilla without graft material utilization: Case report. *Acta Stomatologica Croatica*. 2018; 52:53–60. [PMC free article] [PubMed] [Google Scholar]
23. Nauth A, Schemitsch E, Norris B, Nollin Z, Watson JT. Critical-Size Bone Defects: Is There a Consensus for Diagnosis and Treatment? *J Orthop Trauma*. 2018;32 Suppl 1:S7–S11. [PubMed] [Google Scholar]
24. Hollinger JO, Kleinschmidt JC. The critical size defect as an experimental model to test bone repair materials. *J Craniofac Surg*. 1990;1:60–68. [PubMed] [Google Scholar]
25. Perić Kačarević Ž, Rider P, Alkildani S, Retnasingh S, Pejakić M, Schnettler R, Gosau M, Smeets R, Jung O, Barbeck M. An introduction to bone tissue engineering. *Int J Artif Organs*. 2020;43:69–86. [PubMed] [Google Scholar]
26. Kumar V, Sagheb K, KämmererP. Retrospective clinical study of marginalbone level changes with two different Screw-Implant types: comparison between tissue level and bone level implant. *J Maxillofac Oral Surg* 2014; 13: 259- 66.
27. . Kattimani VS, Chakravarthi SP, Neelima Devi KN, Sridhar MS, Prasad LK. Comparative evaluation of bovine derived hydroxyapatite and synthetic hydroxyapatite graft in bone regeneration of human maxillary cystic defects: a clinico-radiological study. *Indian J Dent Res*. 2014;25:594–601. [PubMed] [Google Scholar]
28. Liu Y, Sun X, Yu J, Wang J, Zhai P, Chen S, Liu M, Zhou Y. Platelet-Rich Fibrin as a Bone Graft Material in Oral and Maxillofacial Bone Regeneration: Classification and Summary for Better Application. *Biomed Res Int*. 2019;2019:3295756. [PMC free article] [PubMed] [Google Scholar]
29. Anitua E, Andia I, Ardanza B, Nurden P, Nurden AT. Autologous platelets as a source of proteins for healing and tissue regeneration. *Thromb Haemost*. 2004; 91: 4–15.
30. Nadon F, Chaput B, Périssé J, de Bérail A, Lauwers F, Lopez R. Interest of mineralized plasmatic matrix in secondary autogenous bone graft for the treatment of alveolar clefts. *J Craniofac Surg*. 2015;26(7): 2148-51.

31. EL Moheb M, Al-Zarea B, Sghaireen M, Toriya J, Mizohata A, Patil S, Siada A, Brad B, Kochaji N, Alam MK, Osuga N. Mineralized Plasmatic Matrix to Enhance the Bone Grafting Technique. *Journal of Hard Tissue Biology* 2017; 26(3): ISSN 1341-7649.
32. Segura-Egea J, Martín-González J, Cabanillas-Balsera D, Fouad A, Velasco-Ortega E, López-López J. Association between Diabetes and the Prevalence of Radiolucent Periapical Lesions in Root-Filled Teeth: Systematic Review and Meta-Analysis. *Clin Oral investigations* (2016) 20(6):1133–41. doi: 10.1007/s00784-016-1805-4 [PubMed] [CrossRef] [Google Scholar]
33. Shen X, Shen X, Li B, Zhu W, Fu Y, Xu R, et al.. Abnormal Macrophage Polarization Impedes the Healing of Diabetes-Associated Tooth Sockets. *Bone* (2021) 143:115618. doi: 10.1016/j.bone.2020.115618 [PubMed] [CrossRef] [Google Scholar]
34. Gadicherla S, Smriti K, Roy S, Pentapati KC, Rajan J, Walia A. Comparison of Extraction Socket Healing in Non-Diabetic, Prediabetic, and Type 2 Diabetic Patients. *Clin Cosmet Investig Dent* (2020) 12:291–6. doi: 10.2147/ccide.S264196 [PMC free article] [PubMed] [CrossRef] [Google Scholar]
35. Aronovich S, Skope LW, Kelly JP, Kyriakides TC. The Relationship of Glycemic Control to the Outcomes of Dental Extractions. *J Oral Maxillofac Surg* (2010) 68(12):2955–61. doi: 10.1016/j.joms.2010.05.006 [PubMed] [CrossRef] [Google Scholar]
36. Feichtinger M., Zemann W., Mossböck R., et al. Three-dimensional evaluation of secondary alveolar bone grafting using a 3D- navigation system based on computed tomography: A two-year follow-up. *Br. J. Oral Maxillofac. Surg.* 2008; 46: 278-82.
37. Bornstein MM, Scarfe WC, Vaughn VM, Jacobs R. Cone beam computed tomography in implant dentistry: a systematic review focusing on guidelines, indications, and radiation dose risks. *Int J Oral Maxillofac Implants.* 2014;29:55-77.
38. Mansour AA, Khalil MA, Shuman MA. Evaluation of mineralized plasmatic matrix as a grafting material around immediate dental implant in mandibular posterior teeth. *Al-Azhar Journal of Dental Science (AJDS)*, 2021; 27(3): 297-305.
39. Osman OAM, Kamel IH, El-Sherief A, Mohamed AM, Magdy A. Effect of Platelet Rich Plasma (PRP) on Bone Graft in Alveolar Cleft Repair. *Egypt, J. Plast. Reconstr. Surg.* 2019; 43(3): 417-424.
40. Sultan OI, Chehata IM, Hossam AM. Implant Stability Parameters & Bone Density Values of Different Graft Materials with Immediately Placed Dental Implants. *Egyptian Dental Journal.* 2018; 64:3135-48.
41. Elbokle NN, Sultan OI, Chehata IM, Hossam AM. Effect of Bone Regeneration with Platelets Rich Fibrin Versus Mineralized Plasmatic Matrix for Immediate Implant Placement. *Egyptian Dental Journal.* 2017; 63:3057-67.
42. Cinar IC, Gultekin BA, Saglanmak A, Yalcin S, Olgac V, Mijiritsky E. Histologic, Histomorphometric, and Clinical Analysis of the Effects of Growth Factors in a Fibrin Network Used in Maxillary Sinus Augmentation. *International Journal of Environmental Research and Public Health*, 2020; 17:1918.



43. Shirota T, Kurabayashi H, Ogura H, et al. Analysis of bone volume using computer simulation system for secondary bone graft in alveolar cleft. *Int. J. Oral Maxillofac. Surg.* 2010; 39: 904-8.
44. Abo Serie YM, El Dibany M, El Halawani GN. Evaluation of mineralized plasmatic matrix with and without collagen membrane in anterior maxillary horizontal alveolar defect (Randomized clinical trial). *Alexandria Dental Journal*, 2022; 47(3) Section A: 63-71.