



## THE EFFICACY OF RESIN INFILTRATION AND MI PASTE - CPP-ACP'S IN MASKING WHITE SPOT LESIONS

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### Abstract

Patients with poor oral hygiene undergoing intervention are more likely to develop decalcification surrounding orthodontic brackets and bands, commonly known as white spot lesions (WSLs) (Behnan et al., 2010, Rodgers et al., 2010). Many times, even after the removal of fixed appliances and natural remineralization, these WSLs are still discernible. This in vitro research's goal was to use spectrophotometric analysis to examine how well two intervention modalities—resin infiltration (RI) and casein phosphopeptide amorphous calcium phosphate (CPP-ACP)—improved light reflectivity and, consequently, the formation of WSLs. Sixty extracted human third molars underwent partial demineralization to produce artificial WSLs, which were then randomly assigned to two treatment groups and an artificial saliva control group. The quantity of light ( $L^*$ ) reflected from each tooth specimen's surfaces was measured prior to and following treatment using a spectrophotometer (VITA Easyshade compact). The rise in  $L^*$  following treatment revealed statistically significant reflectivity enhancements in all three cohorts. There were no statistically significant variations across the research groups, nevertheless. In the end, WSLs were successfully concealed by the control and both treatment strategies.

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## 1. Introduction

Individuals look for orthodontic therapy to enhance their dentofacial aesthetics, but sometimes as a side effect of poor oral hygiene throughout intervention, white spot lesions (WSLs), which are zones of decalcification surrounding orthodontic brackets and bands, can develop (Behnan et al., Rogers et al., 2010). Unfortunately, following the removal of fixed appliances and natural remineralization, numerous WSLs are still apparent. Additionally, there are several published studies on the subject of WSL remineralization, however few of those investigations have compared the efficacy of multiple therapies for WSL hiding.

The resin infiltration (RI) technique's goal is to stop enamel lesions by micro-invasively injecting polymerizable low viscosity resin into the intercrystalline gaps of enamel. The extremely calcified pseudointact surface layer of enamel must be removed from a WSL using hydrochloric acid prior it can be penetrated (Kielbassa et al., 2009). The RI can pierce the enamel up to 400 microns, as opposed to dental sealants, that only adhere to the enamel's surface (Paris et al., 2007). Regarding the loss of the surface covering, an investigation on adhesive penetration revealed that only 60 microns of infiltration were required to stop additional demineralization (Davila et al., 1975).

Amorphous calcium and phosphate ions are bound by a nanocluster called casein phosphopeptide amorphous calcium phosphate (CPP-ACP). The excretion of the calcium and phosphate ions results in a supersaturated concentration of ions in the saliva, which precipitates a calcium-phosphate complex onto

the damaged tooth surface as the pH of the oral environment decreases (Aimutis, 2004). Although there is inadequate and inconsistent evidence to support its effectiveness, the administration of CPP-ACP in the form of MI Paste (Milk Derived Phosphopeptide Infiltration) may be beneficial as an adjunct in reducing or remineralizing WSLs (Guzman-Armstrong et al., 2010, Tung and Eichmiller, 1999).

The present in vitro research's goal was to use spectrophotometric measurements to examine how well two intervention modalities—resin infiltration (RI) and casein phosphopeptide amorphous calcium phosphate (CPP-ACP)—improved light reflectivity and, consequently, the formation of WSLs. The RI and CPP-ACP groups receiving treatment were contrasted with the control group in order to determine whether there was a statistically significant difference in the way they looked of WSLs (measured by the amount of light reflected from the surfaces of each tooth specimen) before and after therapy.

## 2. Methodology

The study was carried out at Department of Conservative Dentistry and Endodontics, Sri Siddhartha Dental College and Hospital, Agalakote, Tumkur. Previously to the investigation, sixty extracted human permanent molars were obtained, cleaned, and preserved in a 0.1% thymol solution to avoid dehydration (Torres et al., 2010). To stop unintended root degradation brought on by the demineralization procedure, the root surface underneath the cemento-enamel junction (CEJ) of each specimen was coated with two layers of an acid-resistant varnish. (Figure 1).



Figure 1: Specimen painted with two coats of acid resistant varnish after demineralization

Customized jigs were made from vinyl polysiloxane bite registration material (Dentsply® Regisil Rigid VPS) for each tooth specimen in order to improve the accuracy and repeatability of L\* estimations. The teeth weren't permitted to dehydrate instead were gently wiped dry. The buccal coronal portion of the sample was contacted

by a cylindrical rod that had the exact same diameter as the spectrophotometer's tip. With the rod in position, vinyl polysiloxane formed around the rod and the buccal and occlusal surfaces of the tooth to produce a distinct jig for each specimen. The diameter and length of each cylindrical rod utilised were both the same. Each of the three

categories of control, RI, and CPP-ACP tooth specimens (n=20 each group) were haphazardly created. The samples were assessed using a spectrophotometer (VITA Easyshade® compact)

that was calibrated in accordance with the manufacturer's instructions before being demineralized (T0), following demineralization (T1), and after therapy (T2).

Step #	Resin Infiltration Protocol
1	Clean the affected tooth and rinse well
	↓
2	Icon® Etch for 2 minutes
	↓
3	Rinse with water and air dry for 30 seconds
	↓
4	Apply Icon®-dry for thirty seconds
	↓
5	Apply Icon® Infiltrant for three minutes. Remove excess with cotton or with sharp explorer. Light cure for forty seconds
	↓
6	With a new tip, apply Icon® Infiltrant again for one minute. Remove excess with cotton or with sharp explorer. Light cure for forty seconds
	↓
7	Measure shade after RI treatment

Figure 2. Protocol for the Resin Infiltration Group

Step #	CPP-ACP Protocol
1	Brush manually for 5 seconds using fluoridated toothpaste
	↓
2	Rinse with distilled water
	↓
3	Apply MI Paste® for three minutes
	↓
4	Without rinsing, place specimen into fresh artificial saliva
	↓

5	Repeat daily for four weeks
	↓
6	Measure shade at end of each week

Figure 3. Protocol for the CPP-ACP Group

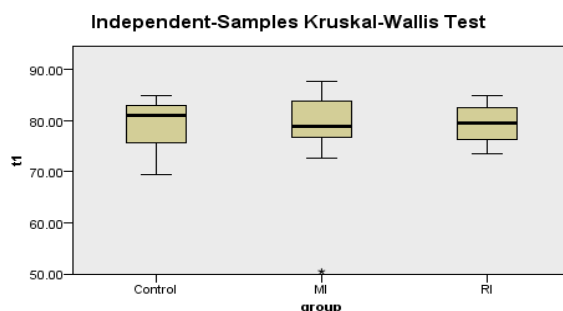
Step #	Control n=20	RI n=20	CPP-ACP n=20
1	Custom jig fabrication	Custom jig fabrication	Custom jig fabrication
	↓	↓	↓
2	Record pre-demineralization shade $T_0$	Record pre-demineralization shade $T_0$	Record pre-demineralization shade $T_0$
	↓	↓	↓
3	Demineralize for 14 days	Demineralize for 14 days	Demineralize for 14 days
	↓	↓	↓
4	Record initial shade after demineralization $T_1$	Record initial shade after demineralization $T_1$	Record initial shade after demineralization $T_1$
	↓	↓	↓
5	Store in artificial saliva for 4 weeks	Resin infiltration application	Daily application of CPP-ACP for 4 weeks
	↓	↓	↓
6	Record final shade $T_2$ at the end of the 4th week	Record final shade $T_2$ immediately after RI	Record final shade $T_2$ at the end of the 4th week

Figure 4. Outline of steps for each of the three groups: Control, RI, and CPP-ACP

Both parametric and nonparametric methods of data analysis were used. All hypothesis tests were two-tailed and carried out using a 0.05 alpha level. With the help of the SAS v9.2 programme (SAS Institute, Cary, NC), statistical analysis was carried out. Both of the MI Paste values ( $T_1$  and  $T_2$ ) and one of the RI values ( $T_2$  alone) may be regarded as outliers (Figures 7 and 8). The Kruskal-Wallis and Wilcoxon Signed Ranks Tests were employed to ascertain whether there were between-group and within-group differences, respectively, in  $T_1$  or  $T_2$  values in order to lessen their influence on the statistical analysis. The ranks of the data were subjected to a two factor (1 between, 1 within) mixed model ANOVA to simultaneously adjust for impacts

### 3. Results

There was no statistically significant between-group variations in  $T_1$  or  $T_2$  values, according to the outcomes of both the parametric and nonparametric statistical analyses. The  $L^*$  values ( $L$ ) within each of the groups did, however, alter in a statistically significant way between  $T_1$  and  $T_2$  (Figures 9, 10, and 11). In other words, there was a substantial difference in  $L$  following treatment in all three groups (Control, RI, and CPP-ACP). The difference between the  $T_2$  and  $T_1$  values was statistically significant ( $p = .001$ ). Overall of the groups'  $T_1$  to  $T_2$  differences in  $L$  were statistically equivalent ( $p = .820$ ). Furthermore, no statistically significant difference in  $T_1$  or  $T_2$  values amongst the groups could be seen ( $p = .891$ ) (See Table 1).



<b>Total N</b>	60
<b>Test Statistic</b>	.067
<b>Degrees of Freedom</b>	2
<b>Asymptotic Sig. (2-sided test)</b>	.967

1. The test statistic is adjusted for ties.
2. Multiple comparisons are not performed because the overall test does not show significant differences across samples.

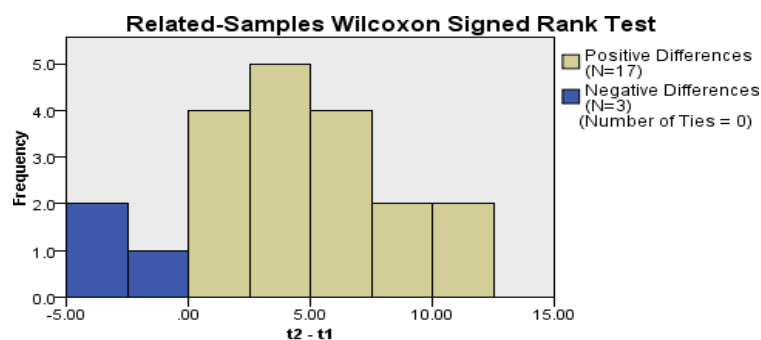
Figure 5. Boxplot of  $T_1$  values of the three groups

### Hypothesis Test Summary

	Null Hypothesis	Test	Sig.	Decision
1	The median of differences between $t_1$ and $t_2$ equals 0.	Related-Samples Wilcoxon Signed Rank Test	.000	Reject the null hypothesis.

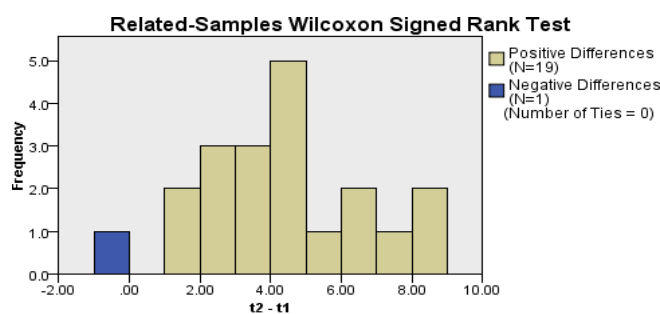
Asymptotic significances are displayed. The significance level is .05.

Figure 6. Boxplot of  $T_2$  values of the three groups



<b>Total N</b>	20
<b>Test Statistic</b>	194.000
<b>Standard Error</b>	26.786
<b>Standardized Test Statistic</b>	3.323
<b>Asymptotic Sig. (2-sided test)</b>	.001

Figure 7:  $T_1 - T_2$  difference in the  $\Delta L^*$  values in Control Group



<b>Total N</b>	20
<b>Test Statistic</b>	209.000
<b>Standard Error</b>	26.786
<b>Standardized Test Statistic</b>	3.883
<b>Asymptotic Sig. (2-sided test)</b>	.000

Figure 8: T<sub>1</sub>- T<sub>2</sub> difference in the ΔL\* values in RI

Table 1: Test of within-subject effects, contrasts and tests of between-subject effects  
 Tests of Within-Subjects Effects

Source		Type III Sum of Squares	df	Mean Square	F	Sig.
time	Sphericity	34307.008	1	34307.008	92.907	.000
	Assumed	34307.008	1.000	34307.008	92.907	.000
	Greenhouse-Geisser	34307.008	1.000	34307.008	92.907	.000
time * group	Lower-bound	34307.008	1.000	34307.008	92.907	.000
	Sphericity	147.467	2	73.733	.200	.820
	Assumed	147.467	2.000	73.733	.200	.820
Error(time)	Greenhouse-Geisser	147.467	2.000	73.733	.200	.820
	Huynh-Feldt	147.467	2.000	73.733	.200	.820
	Lower-bound	147.467	2.000	73.733	.200	.820
	Sphericity	21048.025	57	369.264		
	Assumed	21048.025	57.000	369.264		
	Greenhouse-Geisser	21048.025	57.000	369.264		
	Huynh-Feldt	21048.025	57.000	369.264		
	Lower-bound	21048.025	57.000	369.264		

**Tests of Within-Subjects Contrasts**

Source		Type III Sum of Squares	df	Mean Square	F	Sig.
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time	Linear	34307.008	1	34307.008	92.907	.000
time * group	Linear	147.467	2	73.733	.200	.820
Error(time)	Linear	21048.025	57	369.264		

**Tests of Between-Subjects Effects**

**Transformed Variable: Average**

Source	Type III Sum of Squares	df	Mean Square	F	Sig.
Intercept	439230.000		1 439230.000	284.110	.000
group	356.850		2 178.425	.115	.891
Error	88121.150		57 1545.985		

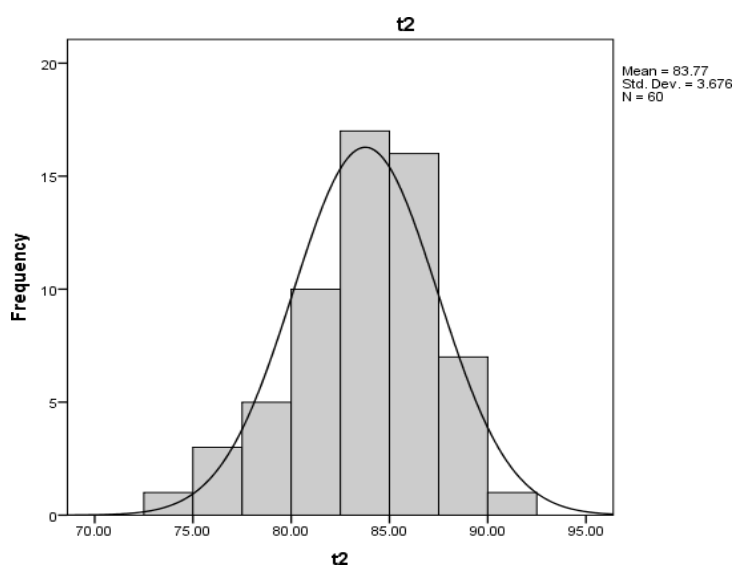


Figure 9: Histogram of mean L\* values of the three groups at T<sub>1</sub> and T<sub>2</sub>

**4. Discussion**

Spectrophotometers emit light to an item using fibre optic technology with the goal to quantify the light that is reflected from the object (Corciolani and Vichi, 2006). In a 3-D colour space, the colour of an object can then be quantified (Torres et al., 2011). Only the L\* value (0-100) was measured because the goal of this study was to assess the masking impact of WSLs using the lightness scale. Whenever lit by the spectrophotometer, things with lower L\* values reflect less light than objects that have greater L\* values. This is generally entities with lower L\* values absorb, scatter, or transmit more light (Fondriest, 2003).

With increasing mineral loss, the scattering coefficient grew exponentially, by more than two orders of magnitude, according to research by Darling et al. on the light scattering characteristics of naturally and artificially demineralized tooth enamel. Ko et al. observed that demineralization of enamel increased the optical scattering coefficient by a factor of

three. They also examined the light scattering of enamel blocks as a consequence of mineral loss. In demineralized teeth, the partial disintegration of individual mineral crystals results in the formation of micropores within the lesion's body. According to Darling et al. (2006), such micropores serve as scattering centres and strongly scatter visible light. As a result, teeth with demineralization have lower L\* values than healthy teeth as they disperse light more and reflects it less. According to the study's findings, the average L\* readings at time T<sub>1</sub> (after demineralization) were actually considerably lower than those at time T<sub>0</sub> (the initial time before demineralization). The removed human third molars were demineralized for fourteen days using the recipe for the demineralization solutions offered by the LLU CDR. This procedure was created based on the findings of our pilot investigation. We found that WSLs that were visible and free of surface cavitation of the enamel's outer surface were generated after fourteen days of demineralization. Over the

objective of this investigation, tooth specimens that had been demineralized for more than 14 days displayed cavitated WSLs. The two pre-defined criteria for successful demineralization in this investigation were the decrease in  $L^*$  value and the sample's appearance as being chalky white. Despite remineralization, a non-cavitated WSL can still be seen clinically and by radiography due to the increased radiolucency and altered physical characteristics. The T2 levels in this investigation came close to, but fell short of, the T0 readings. The potential hiding impact of remineralization is reduced in proportion to the size of the lesion body. According to Gonzalez-Cabezas (2010) and ten Cate et al. (1998), this is caused by a variation in remineralization between the surface of the WSL and the body of the lesion. Despite  $L^*$  values in all three categories statistically improved from T1 to T2, there was no statistically significant difference in  $L^*$  values between the groups. The aesthetic look of WSLs was improved in this work by boosting light reflectivity using RI, CPP-ACP, and artificial saliva. This result was different from that of Torres et al. (Torres et al., 2010), who came to the conclusion that RI therapy was superior to artificial saliva. The disparity in findings could have several causes, one of which could be the size of the WSLs produced by the artificial demineralization process. In 2010, Neuhaus et al. found that RI treatment totally concealed minor WSLs. On the other hand, moderate to large WSLs demonstrated aesthetic enhancements following RI but remained evident following therapy. It's possible that the WSLs generated in the present investigation were smaller than the WSLs used in Torres' research. Irrespective of the chosen intervention method, a WSL with a smaller body cavity will probably experience more remineralization than a WSL with a bigger body cavity. Considering the T2  $L^*$  levels for all groups were nearly equal to the T0 values, it's possible that RI and CPP-ACP don't have enough remineralization ability to compete with artificial saliva.

## 5. Conclusion

This in vitro spectrophotometric study's goal was to determine if RI and CPP-ACP were successful in enhancing the look of WSL as measured by elevated optical reflectivity. The statistically substantial increase in  $L^*$  values from T1 to T2 in this investigation proved the efficacy of RI, CPP-ACP, and artificial saliva in concealing WSLs. Nonetheless weighed against the control group (fake saliva), there

was no statistically significant difference in the efficacy of the experimental groups (RI and CPP-ACP). According to the findings of this research, one group's ability to disguise WSLs is not significantly superior to another group's ability to do so. The benefits of RI and CPP-ACP, nonetheless, might go beyond only improving vision. The preventive remineralization benefits of CPP-ACP might outweigh those of artificial saliva. In bigger sized WSLs, RI, a promising minimally invasive treatment, may be more effective at disguising than synthetic saliva. The research's second important conclusion showed that daily CPP-ACP use for two weeks or four weeks produced similar improvements in  $L^*$  levels. However, prolonged CPP-ACP treatment times may result in further advantages (remineralization, lesions decrease) beyond just the cosmetic improvement in light reflectivity.

## 6. References

- Aimutis W. Bioactive properties of milk proteins with particular focus on anticariogenesis. *J Nutr* 2004;134:989S-95S.
- Arends J, Jongebloed WL. Crystallites dimensions of enamel. *J Biol Buccale* 1978;6:161-71.
- Attin T, Wegehaupt F, Gries D, Wiegand A. The potential of deciduous and permanent bovine enamel as substitute for deciduous and permanent human enamel: Erosion-abrasion experiments. *J Dent* 2007;35:773-7.
- Bailey DL, Adams GG, Tsao CE, Hyslop A, Escobar K, Manton DJ, Reynolds EC, Morgan MV. Regression of post-orthodontic lesions by a remineralizing cream. *J Dent Res* 2009;88(12):1148-53.
- Beerens MW, van der Veen MH, van Beek H, ten Cate JM. Effects of casein phosphopeptide amorphous calcium fluoride phosphate paste on white spot lesions and dental plaque after orthodontic treatment: a 3-month follow-up. *Eur J Oral Sci* 2010;118:610-7.
- Behnan S, Arruda A, Gonzalez-Cabezas C, Sohn W, Peters, M. In-vitro evaluation of various treatments to prevent demineralization next to orthodontic brackets. *Am J Orthod Dentofacial Orthop* 2010;138:712.e1-712.e7.
- Brochner A, Christensen C, Kristensen B, Tranæs S, Karlsson L, Sonnesen L, Twetman S. Treatment of post-orthodontic white spot lesions with casein phosphopeptide-stabilised amorphous calcium phosphate. *Clin Oral Investig* 2011;15(3):369-73.
- Gohring TN, Zehnder M, Sener B, Schmidlin PR. In vitro microleakage of adhesive-sealed dentin with lactic and saliva exposure: a radio-



- isotope analysis. *Journal of Dentistry* 2004;32:235-40.
- Gonzales-Cabezas C. The chemistry of caries: remineralization and demineralization events with direct clinical relevance. *Dental Clinics of North America* 2010;54:469-78.
- Gorelick L, Geiger AM, Gwinnett AJ. Incidence of white spot formation after bonding and banding. *Am J Orthod* 1982;81:93-8.
- Guzman-Armstrong S, Chalmers J, Warren J. White Spot Lesions, Prevention and Treatment. *Am J Orthod Dentofacial Orthop* 2010;138:690-6.
- Hicks MJ, Silverstone LM. Internal morphology of surface zones from acid-etched caries-like lesions: A scanning electron microscopic study. *J Dent Res* 1985;64:1296-1301.
- Holmen L, Thylstrup A, Ogaard B, Kragh F. A scanning electron microscopic study of progressive stages of enamel caries invivo. *Caries Research* 1985;19:355-67.
- Houwink B. The index of refraction of dental enamel apatite. *British Dental Journal* 1974;137:472-5.
- Itthagarun A, Wei SHY, Wefel JS. The effect of different commercial dentifrices on enamel lesion progression: an in vitro pH cycling study. *Int Dent J* 2000;50:21-8.
- Joiner, A. Tooth colour: a review of the literature. *Journal of Dentistry* 2004;32:3-12.
- Kielbassa A, Muller J, and Gernhard C, Closing the gap between oral hygiene and minimally invasive dentistry: A review on the resin infiltration technique of incipient (proximal) enamel lesions *Quin Int* 2009; Sep8(40).
- Ko CC, Tantbiroj D, Want T, Douglas WH. Optical scattering power for characterization of mineral loss. *Journal of Dental Research* 2000;79:1584-9.
- Kumar V, Itthagarun A, Kung NM. The effect of casein phosphopeptide-amorphous calcium phosphate on remineralization of artificial caries-like lesions: an in vitro study. *Australian Dental Journal* 2008;53:34-40.
- Lehmann K, Igiel C, Schmidtman I, Scheller H. Four color-measuring devices compared with a spectrophotometric reference system. *J Dent* 2010;e65-e70.
- Mitchell L. Decalcification during orthodontic treatment with fixed appliances—an overview. *Br J Orthod* 1992;19:199-205.
- Neuhaus K, Graf M, Lussi A, Katsaros C. Late infiltration of post-orthodontic white spot lesions. *J Orofac Orthop* 2010;71:442-7.
- O'Brien WJ, Hemmendinger H, Boenke KM, Linger JB, Groh CL. Color distribution of three regions of extracted human teeth. *Dental Materials* 1997;13:179-85.
- Paris S, Meyer-Lueckel H. Masking of labial enamel white spot lesions by resin infiltration - a clinical report. *Quintessence International* 2009;40:713-8.
- Paul S, Peter A, Pietrobon N, Hammerle CHF. Visual and spectrophotometric shade analysis of human teeth. *J Dent Res* 2002;81(8):578-82.
- Poggio C, Lombardini M, Dagna A, Chiesa M, Bianchi S. Protective effect on enamel demineralization of a CPP-ACP paste: an AFM in vitro study. *J Dent* 2009;37(12):949-54.
- Pulido MT, Wefel JS, Hernandez MM, Denehy GE, Guzman-Armstrong S, Chalmers JM, Qian F. The inhibitory effect of MI paste, fluoride and a combination of both on the progression of artificial caries-like lesions in enamel. *Oper Dent* 2008;33-5:550-5.
- Reynolds EC. Remineralization of enamel sub surface lesions by casein phosphopeptide stabilized calcium phosphate solutions. *J Dent Res* 1997;76:1587-95.
- Robertson MA, Kau CH, English JD, Lee RP, Powers J, Nguyen J. MI Paste Plus to prevent demineralization in orthodontic patients: A prospective randomized controlled trial. *Am J Orthod Dentofacial Orthop* 2011;140:660-8.
- Rogers S, Chadwick B, Treasure E. Fluoride-containing orthodontic adhesives and decalcification in patients with fixed appliances: A systematic review. *Am J Orthod Dentofacial Orthop* 2010;138:390.e1-390.e8.
- Silverstone LM. Remineralization and enamel caries: new concepts. *Dent Update* 1983;10:261-73.
- ten Cate JM, Jongebloed WL, Arends J. Remineralization of artificial enamel lesions invitro IV. Influence of fluorides and diphosphonates on short-and long-term remineralization. *Caries Research* 1981;15:60-9.
- Torrado A, Valiente M, Zhang W, Li Y, Munoz C. Remineralization potential of a new toothpaste formulation: an in-vitro study. *J Contemp Dent Pract* 2004; 5(1):18-30.
- Torres C, Borges A, Torres L, Gomes I, Oliveira R. Effect of caries infiltration technique and fluoride therapy on the colour masking of white spot lesions. *J Dent*, 2010;39:202-7.