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**Phenotypic and molecular detection of efflux pumps in *Klebsiella pneumoniae*.**



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**Abstract**

200 samples were collected from patients suffering from different infections and different age groups for both sexes and from different sources distributed on 80 urine samples (Urine), 42 exit samples (Stool), 62 samples (Sputum) and 26 samples from burns and wounds from patients Patients and patients in Al-Diwaniyah Governorate hospitals, whose ages range between 1-60 years, and for a period of five months, from 9/15/2021 to 01/20/2022. Samples were planted on culture media and isolates were diagnosed using traditional methods (culture, microscopic and biochemical).

The vitek2 device was also used to confirm the diagnosis of *K. pneumoniae* isolates, and the confirmation was reinforced by using the molecular diagnosis of the 16 srRNA gene of *K. pneumoniae*, and the diagnostic results showed a return of 36 *K. pneumoniae* isolates with a percentage of 22.1%.

It was detected phenotypically (using the wooden wheel method) and molecularly the Pcr technique to investigate some genes

The results of phenotypic detection of efflux pumps in 18 bacterial isolates showed that they possessed these Acr ABR pumps, and this was confirmed by the molecular detection of efflux pumps, as the results showed that the percentage of the *acrA* gene was 100%, while the *acrB* gene was found at 55.5%, and the *acrR* gene was found in a percentage 100%.

**KEYWORDS:** Phenotypic ; molecular detection ;efflux pumps ; *Klebsiella pneumoniae*

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## INTRODUCTION

*Klebsiella pneumoniae* is a member of the Enterobacteriaceae family, which lives normally in the human intestine, but when present in other parts of the body, it causes many diseases (Siu et al.,2011). It also lives widely in surface water, sewage, soil, plants and mucous membranes of mammals (White et al.,2002). It is one of the most important opportunistic causes of infection in hospitals and society (Togawa et al.,2015).

These bacteria possess many mechanisms of resistance to antibiotics, including their possession of efflux pumps (EPs), which are proteins located in the bacterial plasma membrane. Reaching the goal as well as getting rid of harmful substances for bacteria such as antibiotics and putting them outside (Pagès et al.,2011). These pumps are widely present in Gram-negative bacteria as they pump water-soluble substances such as antibiotics out of the bacterial cell as a method of bacterial resistance to antibiotics (Poole , 2007). Which plays a major role in the pathogenesis of these bacteria, as efflux pumps pump antibiotics out of the cell (Blair et al., 2014). An increase in the number of efflux pumps was observed in *K. pneumoniae* with a type of (MDR) multi drug resistance (Pages et al.,2009).

One of the important pumps in *K. pneumoniae* is the AcrABR pumps, which when lost, the bacteria become more sensitive to a group of antibiotics such as erythromycin, chlorophenicol, fluoroquinolones, gentamycin, and others (Padilla et al., 2010). This mechanism of resistance increased significantly among isolates of *K. pneumoniae* bacteria and to a large group of antibiotics, especially fluoroquinolones (Schneiders et al., 2003).

## MATERIALS AND METHODS

200 samples were collected from different sources, distributed on 80 urine samples, 42 exit samples (stool), 62 sputum samples, and 26 samples of burns and wounds from

patients attending and in Diwanayah Teaching Hospital and Women's and Children's Hospital in the governorate. Al-Diwaniyah, whose ages ranged between 1-60 years and for a period of five months from 9/15/2021 to 01/20/2022. Samples were taken using sterile cotton swabs to collect samples and the dishes were incubated at a temperature of 37°C for 24 hours. On the center of the Macconkey agar and blood agar.

Diagnosis of isolates / The API 20E system was used to diagnose bacterial isolates, as the instructions of the supplied company were followed, and then the diagnosis was made using the Vitek 2 device to diagnose bacterial isolates with a high degree of accuracy after confirming them by preliminary biochemical tests.

Phenotypic detection of effluent pumps / Decimal dilutions of all bacterial isolates were prepared using sterile physiological salt solution and its turbidity was compared with a standard turbidity constant solution (McFarland). This detection was carried out on bacterial isolates that have antibiotic resistance by adopting the modified EtBr-agar cartwheel method and by using soybean agar agar and ethidium bromide dye with different concentrations according to what was mentioned (Martins et al.,2013). As it comes:

- Different concentrations of ethidium bromide dye (5, 10, 15, 20, 25) mcg/ml were prepared by adding different amounts of the above dye to the medium of trypton agar soybeans after sterilizing and slightly cooling it.
- The media were shaken well and after sterilization, they were poured into sterilized dishes that were previously divided radially and kept at refrigerator temperature until use.
- Sterile cotton swabs were passed over the diluted bacterial suspension, then pressed on the inner wall to get rid of the excess culture, then they were planted in a radial line from the edge of the dish to its center

for each isolate and for each concentration, then the dishes were incubated at a temperature of 37 °C for a period of 16 hours.

- Bacterial isolates were tested using a UV source to observe the fluorescence intensity.

#### Molecular detection of effluent pump

**Chromosomal DNA extraction** The chromosomal DNA of selected colistin-resistant bacterial isolates was extracted using a special kit (Genomic DNA extraction kit) provided by Geneaid Corporation of America.

Preparation of the polymerase chain reaction mixture / The reaction mixture was prepared using the AccuPower® PCR PreMix kit supplied by the Korean company (Bioneer) and using the primers in Table (1) designed with the Efflux Pump operon gene primers and the DNA sequencing initiator in this The study was carried out using the genetic sequence of these genes in the gene bank on the National Biological Information website of the NCBI, using the Primer 3 plus primer design program, and these primers were supplied by the Korean company Bioneer.

**Table 1. The primers used in the study with the sequence of nitrogenous bases and the size of the primer.**

Primer	Nitrogen Bases		Product
<b>AcrA</b>	<b>F</b>	<b>ATG AAC AAA AAC AGA GGG TTA ACG-3'5-</b>	<b>1194</b>
	<b>R</b>	<b>5-TTA AGA CTT GGT TTG TTC TGA TGG-3'</b>	
<b>AcrB</b>	<b>F</b>	<b>ATG CCT AAT TTC TTT ATC GATCGC-35-</b>	<b>3147</b>
	<b>R</b>	<b>TTA ATG ATG CTC AAC CTG ATG GC-35-</b>	
<b>AcrR</b>	<b>F</b>	<b>ATG GCA CGA AAA ACC AAA CAA C-35-</b>	<b>651</b>
	<b>R</b>	<b>TTA AGC TGA CAA GCT CTC CGG-35-</b>	

## RESULTS AND DISCUSSION

The current study dealt with the collection of a number of pathological samples in order to obtain isolates of *K. pneumoniae* bacteria with multiple resistance to antibiotics, as 200 clinical samples were collected from different pathological cases from Al-Diwaniyah Teaching Hospital and Women and Children Hospital in Al-Diwaniyah Governorate for the period from 15/9/2021 until 1/22022 And based on biochemical tests and Vitek2 diagnosis, the results showed that *K.pneumoniae* bacteria constitute 36 isolates with a rate of 22.1%.

#### Phenotypic detection of efflux pumps

The detection of efflux pumps in 18 bacterial isolates was carried out using the EtBr CW wheel method (Gressler et al., 2014). Relying on ethidium bromide stain as a proxy for phenotypic detection. The results showed that 18 isolates (100%) were positive for phenotypic detection (Table 2), depending on the lowest concentration in which the isolates did not appear to shimmer under ultraviolet (UV) rays (Gawad et al., 2018). Thus, our results came close to (Al-Saadi et al.,2019) whose isolates showed a positive percentage of phenotypic detection of effluent pumps at a rate of 92%.

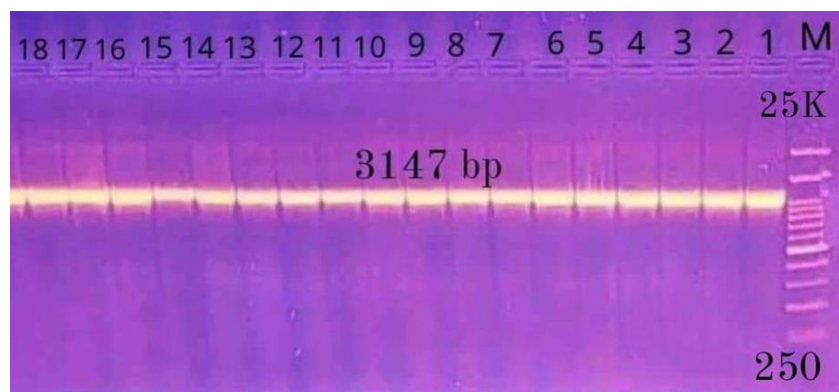
**Table 2. Results of phenotypic detection of efflux pumps in K. Pneumoniae bacteria using different concentrations of ethidium bromide dye.**

	Ethidium bromide dye concentration (µg/ml)				
	5	10	15	20	25
1	-	-	-	+	+
2	-	-	-	-	+
3	-	-	+	+	+
4	-	-	-	+	+
5	-	-	+	+	+
6	-	-	-	+	+
7	-	-	-	-	+
8	-	-	-	-	+
9	-	-	-	+	+
10	-	-	-	+	+
11	-	-	-	-	+
12	-	-	-	+	+
13	-	-	+	+	+
14	-	-	+	+	+
15	-	-	-	-	+
16	-	-	-	-	+
17	-	-	+	+	+
18	-	-	-	+	+

Molecular detection of the AcrAB efflux pump in K. pneumonia

The results of the molecular detection of the *acrA* gene with a size of 3147 base pairs using PCR technology for 18 bacterial isolates under study showed its presence in 100%, by comparing the doubled bundles with the bundles belonging to the DNA ladder. The resulting bundles were also found to have a molecular weight of 3147 base pair. Figure (1)

This study agreed with the findings (Pakzad et al.,2013), as the percentage of bacterial isolates that possessed the *acrA* gene was 100%. Our results (Maleki et al.,2017; Lafta et al.,2022) also confirmed that all isolates of *K. pneumoniae* resistant to ciprofloxacin contained the *acrA* gene, while (Razavi et al.,2020; Margiana et al.,2022) the frequency of the *acrA* gene was found, reaching 82.90%. *acrA* is 52.72%.

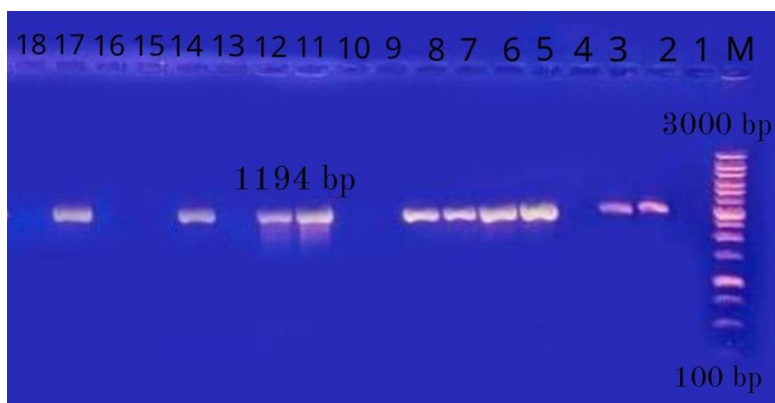


**Figure 1. Electrophoresis of the results of the acrA base pair polymerase chain reaction (3147) of isolates of *K. pneumoniae* on 1.5% agarose gel, voltages 70, 80 amps, 1 hour, column M represents DNA ladder**

#### Genetic detection of the acrB gene

The results of molecular detection of the acrB gene with a size of 1194 base pairs by PCR technique showed for 18 bacterial isolates under study. That 55.5% of the isolates possessed the acrB gene and by comparing the doubled bundles with the bundles belonging to the DNA ladder) (it was found that the resulting bundles had a molecular weight of 1194 base pairs. Figure 2). Thus, our results were almost identical to (Li et al.,2022; Abbas et al.,2022) who

used the PCR technique In order to investigate the acrB gene and they obtained 52.72% of it among their isolates, either) (Pakzad et al.,2013; Hussein et al.,2022) recorded the percentage of appearance of this gene amounted to 100%, and thus our results were inconsistent with them, as (Razavi et al.,2020) violated our results in recording this gene at a rate of 95.90% and in a study conducted by (Schneiders et al.,2003; Al-Jassani et al.,2022) as They recorded the presence of this gene among their isolates by 95%.



**Figure 2. Electrophoresis of the polymerase chain reaction results of acrB 1194 (base pair) gene of isolates of *K. pneumoniae* on 1.5% agarose gel, voltages 70, 80 amps, for 1 hour, column M represents the DNA ladder**

#### Genetic detection of the acR gene

The results of molecular detection of the acrR gene, which has a size of 651 base pairs, by PCR technique for 18 bacterial isolates under study, showed a 100%

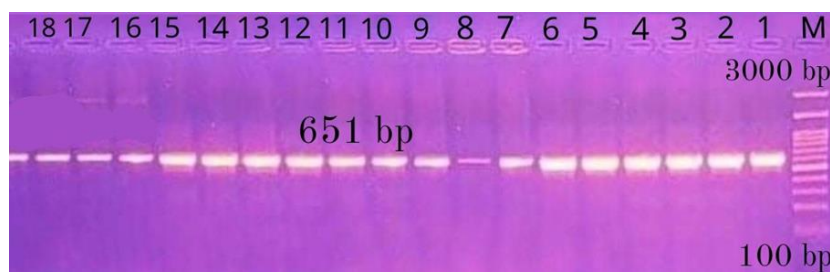
appearance rate (Togawa et al., 2015; Zadeh et al.,2022; Arif et al.,2023)

Studies of some Enterobacter family members indicate that mutations within the AcrR repressor lead to increased expression



of *acrA* and *acrB*. Complementation of mutated *acrR* with a gene of the non-mutated type has been shown to reduce the level of antibiotic resistance, suggesting a role of the functional inhibitor in controlling the highly drug-resistant

phenotype (Padilla et al.,2010;Rohmah et al.,2023). It was found that deletion of the *acrR* gene led to a moderate increase in MICs for most of the antibiotics tested compared to the parent strain.



**Figure 3. Electrophoresis of *acrR*(651) base pair polymerase chain reaction results of isolates of *K. pneumoniae* on 1.5% agarose gel, voltages 70, 80 amps, 1 hour, column M represents DNA ladder.**

### Compliance with Ethical Standards statements

#### Ethical approval:

The manuscript is written in original and all the data, results pertaining to this manuscript are original according to the research performed. The authors followed academic integrity and have not copied any content/results from another source.

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**Conflict of interest:** The authors of the study do not have any conflict of interest

**Informed Consent:** The authors of the manuscript agrees to publish this research in the journal if it's considerable by the editors of the journal. The authors provide full consent for reviewing and publishing this manuscript.

**V.** All the authors of this study contributed equally in terms of performing the research as well as in preparing the manuscript. All the authors of the study followed the guidelines of the corresponding author. Any query/suggestion related to the manuscript can be reached to the corresponding author

### REFERENCES

- Al-Saadi, Z. H. A., & Al-Shwaikh, R. M. . (2019). Phenotypic and molecular detection of *Escherichia coli* efflux pumps from UTI patients. University of Baghdad.
- Blair, J. M., Richmond, G. E., & Piddock, L. J. (2014). Multidrug efflux pumps in Gram-negative bacteria and their role in antibiotic resistance. *Future microbiology*, 9(10), 1165-1177.
- Gawad, W. E., Helmy, O. M., Tawakkol, W. M., & Hashem, A. M. (2018). Antimicrobial resistance, biofilm formation, and phylogenetic grouping of uropathogenic *Escherichia coli* isolates in Egypt: The role of efflux pump-mediated resistance. *Jundishapur J Microbiol*, 11(2), e14444.
- Gressler, L. T.; Vargas, A. C. D.; Costa, M. M. D.; Pötter, L.; Silveira, B. P. D.; Sangioni, L. A. and Botton, S. D. A. (2014). Genotypic and phenotypic detection of efflux pump in *Rhodococcus equi*. *Brazilian J Microbiol*. 45(2): 661-665 .
- Li, Y., Cross, T. S., & Dörr, T. (2022). Analysis of *AcrB* in *Klebsiella pneumoniae* reveals natural variants promoting enhanced multidrug resistance.

- Research in Microbiology, 173(3), 103901.  
<https://doi.org/10.1016/J.RESMIC.2021.103901>
- Maleki, D.; Jahromy, S. H.; Karizi, S. Z. and Eslami, P. (2017). The Prevalence of *acrA* and *acrB* Genes Among Multiple-Drug Resistant Uropathogenic *Escherichia coli* Isolated from Patients with UTI in Milad Hospital, Tehran. *Avicenna J Clin Microb Infect.* 4(1): 1-7
- Martins, M.; Mccusker, M. P.; Viveiros, M.; Couto, I. ; Fanning, S.; Pages, J. M.; Amaral, L. (2013). A Simple Method for Assessment of MDR Bacteria for Over- Expressed Efflux Pumps. *Open Microbiol J.* 7: 72-82.
- Padilla, E., Llobet, E., Doménech-Sánchez, A., Martínez-Martínez, L., Bengoechea, J. A., & Albertí, S. (2010). *Klebsiella pneumoniae* AcrAB efflux pump contributes to antimicrobial resistance and virulence. *Antimicrobial agents and chemotherapy*, 54(1), 177-183.
- Padilla, E., Llobet, E., Doménech-Sánchez, A., Martínez-Martínez, L., Bengoechea, J. A., & Albertí, S. (2010). *Klebsiella pneumoniae* AcrAB Efflux Pump Contributes to Antimicrobial Resistance and Virulence. *Antimicrobial Agents and Chemotherapy*, 54(1), 177–183. <https://doi.org/10.1128/AAC.00715-09>
- Pagès, J. M., Amaral, L., and Fanning, S. (2011). An original deal for new molecule: reversal of efflux pump activity, a rational strategy to combat Gram-negative resistant bacteria. *Curr. Med. Chem.* 18, 2969–2980. doi: 10.2174/092986711796150469.
- Pages, J. M., Lavigne, J. P., Leflon-Guibout, V., Marcon, E., Bert, F., Noussair, L., & Nicolas-Chanoine, M. H. (2009). Efflux pump, the masked side of  $\beta$ -lactam resistance in *Klebsiella pneumoniae* clinical isolates. *PLoS one*, 4(3), e4817.
- Pakzad I, Zayyen Karin M, Taherikalani M, Boustanshenas M, Lari AR. Contribution of AcrAB efflux pump to ciprofloxacin resistance in *Klebsiella pneumoniae* isolated from burn patients. *GMS Hyg Infect Control.* 2013;8(2):Doc15. doi:10.3205/dgkh000215.
- Poole, K. (2007). Efflux pumps as antimicrobial resistance mechanisms. *Annals of medicine*, 39(3), 162-176.
- Razavi, S., Mirnejad, R., & Babapour, E. (2020). Involvement of AcrAB and OqxAB efflux pumps in antimicrobial resistance of clinical isolates of *Klebsiella pneumoniae*. *Journal of Applied Biotechnology Reports*, 7(4), 251-257.
- Schneiders, T., Amyes, S. G. B., & Levy, S. B. (2003). Role of AcrR and RamA in fluoroquinolone resistance in clinical *Klebsiella pneumoniae* isolates from Singapore. *Antimicrobial agents and chemotherapy*, 47(9), 2831-2837.
- Schneiders, T., Amyes, S. G. B., & Levy, S. B. (2003). Role of AcrR and RamA in Fluoroquinolone Resistance in Clinical *Klebsiella pneumoniae* Isolates from Singapore. *Antimicrobial Agents and Chemotherapy*, 47(9), 2831–2837. <https://doi.org/10.1128/AAC.47.9.2831-2837.2003>
- Siu, L. K., Fung, C. P., Chang, F. Y., Lee, N., Yeh, K. M., Koh, T. H., & Ip, M. (2011). Molecular typing and virulence analysis of serotype K1 *Klebsiella pneumoniae* strains isolated from liver abscess patients and stool samples from noninfectious subjects in Hong Kong, Singapore, and Taiwan. *Journal of clinical microbiology*, 49(11), 3761-3765.
- Togawa, A., Toh, H., Onozawa, K., Yoshimura, M., Tokushige, C., Shimono, N., ... & Tamura, K. (2015). Influence of the bacterial phenotypes on the clinical manifestations in *Klebsiella pneumoniae* bacteremia patients: a retrospective cohort study. *Journal of Infection and Chemotherapy*, 21(7), 531-537.

- White, D. G., Zhao, S., Simjee, S., Wagner, D. D., & McDermott, P. F. (2002). Antimicrobial resistance of foodborne pathogens. *Microbes and infection*, 4(4), 405-412.
- Zadeh, F. A., Bokov, D. O., Salahdin, O. D., Abdelbasset, W. K., Jawad, M. A., Kadhim, M. M., ... & Khatami, M. (2022). Cytotoxicity evaluation of environmentally friendly synthesis Copper/Zinc bimetallic nanoparticles on MCF-7 cancer cells. *Rendiconti Lincei. Scienze Fisiche e Naturali*, 33(2), 441-447.
- Rohmah, M. K., Salahdin, O. D., Gupta, R., Muzammil, K., Qasim, M. T., Al-Qaim, Z. H., ... & Abarghouei, S. (2022). Modulatory role of dietary curcumin and resveratrol on growth performance, serum immunity responses, mucus enzymes activity, antioxidant capacity and serum and mucus biochemicals in the common carp, *Cyprinus carpio* exposed to abamectin. *Fish & Shellfish Immunology*, 129, 221-230.
- Arif, A., Alameri, A. A., Tariq, U. B., Ansari, S. A., Sakr, H. I., Qasim, M. T., ... & Karampoor, S. (2023). The functions and molecular mechanisms of Tribbles homolog 3 (TRIB3) implicated in the pathophysiology of cancer. *International Immunopharmacology*, 114, 109581.
- Margiana, R., Alsaikhan, F., Al-Awsi, G. R. L., Patra, I., Sivaraman, R., Fadhil, A. A., ... & Hosseini-Fard, S. (2022). Functions and therapeutic interventions of non-coding RNAs associated with TLR signaling pathway in atherosclerosis. *Cellular Signalling*, 100, 110471.
- Lafta, H. A., AbdulHussein, A. H., Al-Shalah, S. A., Alnassar, Y. S., Mohammed, N. M., Akram, S. M., ... & Najafi, M. (2023). Tumor-Associated Macrophages (TAMs) in Cancer Resistance; Modulation by Natural Products. *Current Topics in Medicinal Chemistry*.
- Al-Jassani, M. J., Sayah, M. A., Qasim, M. T., Kadhim, A. J., & Muhammad, E. H. (2022). Isolation and Evaluation of Antibacterial Agents Produced by Soil Bacillus SP. and Study Some of their Immunological Parameters. *Revista Electronica de Veterinaria*, 23(4), 105-111.
- Hussein, H. A., Khudair, S. A., Alwan, M., Aljawahiry, T., T Qasim, M., & V Pavlova, I. (2022). Impact of pollution caused by salmon breeding centers on river water quality. *Caspian Journal of Environmental Sciences*, 20(5), 1039-1045.
- M Abbas, M., W Abooud, K., Qasim Mohammed, A., Hasan Al-Zubaidi, S., Hussain, A., M Hameed, N., ... & Ahmad Batayneh, K. (2022). Effects of Various Irrigation Levels and Biochar-Based Fertilizers on Peanut Production. *Journal of Nuts*, 13(4), 289-300.