



## ASSESSMENT OF THE BENEFICIAL EFFECT OF CHIA AND QUINOA-FORTIFIED GLUTEN-FREE BISCUIT PRODUCTS ON CELIAC DISEASE IN RATS

Marwa Adel Awad El-Said<sup>1\*</sup>, Maysa Mohamed ElMallah<sup>1</sup>, Mostafa  
Abbas Shalaby<sup>2</sup>, Hany Gaber Elmasry<sup>1</sup>

ArticleHistory:Received:01.05.2023

Revised:03.06.2023

Accepted:09.06.2023

### ABSTRACT

Celiac disease (CD) is a chronic inflammatory disease of the upper small intestine triggered by gluten protein intolerance, which is prevalent in genetically predisposed individuals. The present study aimed to evaluate the beneficial effect of Chia and Quinoa-fortified gluten-free biscuit products on CD in Rats. Thirty-five male Sprague Dawley rats (225±5 g b.wt. and 8 weeks age) were used. Rats were distributed into 5 equal groups; Group1: negative control, group 2: positive control with CD and groups 3, 4 and 5: with CD and were fed Chia, Quinoa and their combination, respectively. Feed intake (FI), body weight gain (BWG %), and feed efficiency ratio (FER) were calculated. Rats were then anesthetized and blood samples were collected for determination of activities of serum liver and kidney function, total cholesterol (TC) and triglyceride (TG) and lipoprotein fractions were measured. Reduced glutathione (GSH), malondialdehyde (MDA) and lactate dehydrogenase (LDH) were measured. Histopathology of small intestine was also carried out. Results revealed that feeding Chia, Quinoa and their combination decreased FI, BWG% and FER, TC and TG and decreased serum parameters of elevated hepatorenal function as compared to the positive control (+ve) group. Histopathological examination of small intestine demonstrated that feeding combination of Chia and Quinoa ameliorated lesions of CD and inflammation. It could be concluded that Chia and Quinoa would be an effective curative for celiac disease- induced histopathological changes in the small intestine due to its improvement of intestinal histopathological lesions.

**Keywords:** Celiac disease- Gluten- Chia - Quinoa- Biochemical analysis, Histopathology.

<sup>1</sup> Nutrition Department, Faculty of Home Economic, Helwan University, Egypt.

<sup>2</sup> Pharmacology Department, Faculty of Veterinary Medicine, Cairo University, Egypt.

\*Corresponding author: Marwa Adel Awad El-Said; Email: [marwaade8284@gmail.com](mailto:marwaade8284@gmail.com); Mobile: 01111361159

DOI: 10.48047/ecb/2023.12.7.31

### INTRODUCTION:

Celiac disease (CD) is a chronic inflammatory disease of the upper small intestine triggered by gluten protein intolerance, which is prevalent in genetically predisposed individuals. The prevalence of the celiac disease in Arab countries varies with sex and age. However, It was found that CD presented similar clinical characteristics independent of the geographic region. (Paivi *et al.*, 2020). The classical manifestation of CD is often present with all related signs and symptoms of malabsorption. Moreover, patients also experience diarrhea, steatorrhea, and loss of weight or growth failure. Meanwhile, in children, the classical characteristics are diarrhea, failure to thrive, muscle wasting, poor appetite, abdominal distension, and sometimes emotional distress and lethargy (Paivi *et al.*, 2020). Currently the only effective treatment for CD is the gluten free diet (GFD), which is extremely difficult to adhere to and significantly reduces the patients' quality of life.

Therefore, investigations are in progress to find new therapies varying from production of genetically modified wheat to blockade of inflammatory cytokines. Meanwhile, the discovery of new endogenous immunoregulatory substances suggests new therapeutic approaches (Rashtak and Murray, 2012).

Gluten is the wheat grain protein richly consumed in Western countries with an average daily intake of 10 to 20 grams/person/day (Cohen *et al.*, 2019). It is comprised of prolamin and glutelin proteins. Both proteins abundantly possess glutamine and proline residues, which defy gastrointestinal digestion and promote the deamination process through the tissue transglutaminase (tTG) enzyme. It may lead to mucosal inflammation and villous atrophy, thus causing malabsorption (Assa and Frenkel., 2017).

Chia is commonly known as *Salvia hispanica L.*, an oilseed plant, and is native from southern Mexico and

northern Guatemala. It is a traditional food in Central and South America. Chia seed contains 25 to 40% oil with high essential fatty acids (omega)  $\omega$ -3 alpha-linolenic acid (60%) and (omega)  $\omega$ -6 linoleic acid (20%). It is also high in protein (19-23%), dietary fiber (18-30%), vitamins, minerals and antioxidants (Norlaily *et al.*, 2012). Due to its unique nutritional composition, especially its high unsaturated fatty acid composition, dietary fiber and antioxidant activities its consumption helps to increase satiety index and decreases the risk of various types of diseases such as cardiovascular diseases, cancer, diabetes, inflammatory and autoimmune diseases (Munoz *et al.*, 2013). A number of studies have investigated the inclusion of different seeds into gluten-free breads in order to increase their nutritional value. Because of its nutritional composition, particularly the omega-3 fatty acid, fiber and antioxidant content, Chia is an ideal seed for the enrichment of cereal products, and there has been an increase in the number of commercial products that incorporate chia in their formulation in recent years (Eugenia *et al.*, 2014). Quinoa, a pseudocereal that belongs to the *Chenopodiaceae* family has a higher nutritive value than traditional cereals. It has high protein content (10-18%) with balanced amino acid composition, supplying high contents of lysine and methionine (Nowak *et al.*, 2016). The fat content of raw quinoa seeds was 9.7% (dry-weight basis) and it has similar fatty acid composition with soybean oils with high amounts of essential fatty acids linoleic (52.3%) and linolenic acids (3.9%). These essential fatty acids required for good health, cannot be synthesized in human body and must be obtained from the diet (Costantini *et al.*, 2014). It contains a significant amount of fiber (2.1-4.9%), more calcium, magnesium, potassium, iron, copper, riboflavin (B2),

$\alpha$ -Tocopherol (vitamin E),  $\beta$ -Carotene and ascorbic acid (vitamin C) than wheat, barley and rice. It is also a rich source of other bioactive compounds (polyphenols, phytosterols, etc.) (Aktaş and Levent, 2018). Quinoa used in the bakery industry because the starch present in the seeds has properties similar to those found in wheat. In addition to augmenting the nutritional value, it is free of gluten, so it can be eaten by people who have CD as well as by those who are allergic to wheat (Stikic *et al.* 2012)

## MATERIAL AND METHODS:

### A. Materials

Basal diet was prepared according to the method of Reeves *et al.* (1993). It was consisted of 20 % protein (casein), 10 % carbohydrate, 4.7% fat (corn oil), 2% choline chloride, 1% vitamin mixture, 3.5 % salt mixture and 5% fibers. The remainder was corn starch up to 100 %. Raw material Chia seeds, whole quinoa flour, rice flour, fine granulating sucrose, whole egg, oil, cinnamon, salt, baking powder and ethyl vanillin were obtained from Agricultural Research center in Cairo. Kits for blood analysis were purchased from local distributor of (Sigma Chemical), Cairo, Egypt. Gliadin (G-3375; Sigma chemical substance to induction of celiac disease obtained from El-Gomhoryia Company, Cairo, Egypt. Animals: A total number of thirty-five mice (Sprague Dawley strain) weighing (20 to 25 g) body weight, were purchased from the Agriculture Research Center, Giza, Egypt.

### B. Methods:

#### Preparation of Flour Formulas:

The free gluten flour formula consists of (rice flour), chia flour (CF) and quinoa flour (QF) were mixed together for preparation the biscuits by replacement.

**Table (1):** Flour formulas (as weight) for preparation of free gluten bakery products (biscuits).

Formula	Rice flour (g)	Chia flour(g)	Quinoa flour(g)
Control	200	-	-
Sample 1	190	10	-
Sample 2	180	20	-
Sample 3	160	40	-
Sample 4	190	-	10
Sample 5	180	-	20
Sample 6	160	-	40
Sample 7	190	5	5
Sample 8	180	10	10
Sample 9	160	20	20

### Preparation of Bakery Products (Biscuits):

Preparation of the different types of free gluten (biscuits). Control gluten-free biscuits sample was prepared using the following recipe: Gluten free flour (rice flour) 200g, fine granulating sucrose (40 g), whole egg (50 g), salt (0.5 g), baking powder (5 g), ethyl vanillin (0.1 g) and sunflower oil (40)g (A.O.A.C.1995). The replacement by chia and quinoa flour with the percentage of chia flour (5, 10, 20 %), Quinoa flour (5, 10, 20 %) and mixture of chia: quinoa flour (1:1) as (5, 10, 20%)

#### **Induction of Celiac Disease (CD):**

To induce celiac disease, Gliadin (G-3375; Sigma) was dilute in 0.02 M acetic acid to form 10% solution. The prepared solution was administered intragastrically by gavage at a dose of 1.5 mg/g body weight for five days as reported by **Nikoukaret al., (2014)**.

#### **Experimental Animal Design:**

Thirty-five rats male Sprague Dawley strain rats was housed in well-aerated cages under hygienic conditions, and fed on basal diet for one week for adaptation.

After the adaptation period, rats will be divided into two main groups, as follows:

**First group:** Negative control group, rats (n=7) was fed on basal diet only during the experimental period.

**Second group:** all remaining rats (n=28), were intragastrically given (1.5 mg/g body weight) from Gliadin diluted in 0.02 M acetic acid to form 10% solution for 5 days to induced celiac disease, and they were divided as follow:

**Subgroup (1):** rats serve as positive control group basal diet only.

**Subgroup (2):** rats were feed Basel diet (with bakery product with chia seed).

**Subgroup (3):** rats were feed Basel diet (with bakery product with quinoa seed).

**Subgroup (4):** rats were feed Basel diet (with bakery product with chia seed and quinoa).

#### **Biological Evaluation:**

Daily feed intake (FI) was recorded day after day throughout the experimental period (3 weeks). Body weight gain (BWG) was calculated. Determination of body weight gain percent and feed efficiency ratio (FER) were assessed according to the method described by **Chapman et al. (1959)** using the following equations:

**BWG %** = final weight (g) – Initial weight (g) / Initial weight (g) ×100

**Feed efficiency ratio (FER)** = Weight gain (g) / feed consumed (g)

#### **Blood Collection and Serum Separation:**

At the end of the experimental period (2 weeks), rats were fasted overnight and authorized by ether, intestinal was dissected out and kept in 10% neutral buffered formalin for histopathological examination

and blood samples were collected from each rat and centrifuged at 3000 rpm for 15 min to obtain the serum for biochemical analysis.

#### **Biochemical Analysis:**

Total cholesterol (TC) was determined according to **Richmond, (1973)**. Triglyceride (TG) was determined according to **Wahlefeld, (1974)**. High density Lipoprotein cholesterol (HDL-C) was determined according to **Albers et al., (1983)**. Low density Lipoprotein cholesterol (LDL-C) and very Low density Lipoprotein cholesterol (VLDL-C) were estimated according to the equation described by **Friedewald et al., (1972)**. Aspartate aminotransferase (AST) and alanine aminotransferase (ALT) concentration were determined according to **Young, (2001)**. Alkaline phosphatase (ALP) concentration was determined according to **Roy, (1970)**. Serum uric acid and creatinine levels were estimated as described by **Lorentz and Brendt (1967)** and Glutathione peroxidase (GPx) was determined according to **Hissin and Hilf, (1976)**. Serum lactate dehydrogenase was determined according to **Vassault (1986)**. Serum malondialdehyde (MDA) was determined by the method of **Ohkawa et al., (1979)**.

#### **Histopathological Examination:**

Immediately after scarifying of rats, intestinal dissected out and kept in 10% neutral buffered formalin for histopathological examination according to **Bancroft and Stevens (1996)**. Histopathological examination was carried out at the Department of Pathology, Faculty of Veterinary medicine, Cairo University, Egypt.

#### **STATISTICAL ANALYSIS:**

All data obtained results were analyzed using Statistical Package for the Social Sciences (SPSS) for Windows, version 20 (SPSS Inc., Chicago, IL, USA). Collected data was presented as mean± standard deviation (SD). Analysis of Variance (ANOVA) test was used for determining the significances among different groups according to **Armitage and Berry (1987)**. All differences were considered significant if P-values were (P< 0.05).

#### **RESULTS AND DISCUSSION:**

Data in table (2) show that feeding of rats with Chia and Quinoa-fortified gluten free biscuit products decreased FI, BWG% and FER in the positive group (rats with celiac disease) as compared to the normal control rats. Feeding Chia, Quinoa and their combination to rats significantly increase. FI, BWT% and FER.

These findings were similar to those reported by **Sadeek et al., (2021)** who concluded that the

incorporation of chia and quinoa flour with wheat flour have beneficial effects for improving the

chemical, nutritional, rheological, and sensory properties of food product pasta.

**Table (2):** Effect of Chia and Quinoa-fortified gluten-free biscuit products on feed intake (FI), body weight gain (BWG %) and feed efficiency ratio (FER) in rats

Parameters Groups	FI (g/day/rat)	BWG (%)	FER
Control (-ve)	10.0±0.71a	48.56±2.71a	0.230±0.009a
Control (+ve)	9.0±0.67c	24.07±2.67c	0.120±0.003b
Chia	12.0±0.34bc	30.35±2.34bc	0.121±0.007b
Quinoa	12.5±0.10bc	32.88±2.10bc	0.128±0.006b
Chi and Quinoa	13.0±0.71b	36.28±2.71b	0.132±0.008b

Means in the same column with different letters are significantly different at P≤0.05

**Table 3.** Effect of Chia and Quinoa-fortified gluten-free biscuit products on total(TC) and triglycerides (TG) in rats. (n=7 rats)

Parameters Groups	Total Cholesterol (mg/dL)	Triglycerides mg/dL
Control (-ve)	96.14±1.07b	98.42±2.12b
Control (+ve)	122.42±1.43a	122.71±2.47a
Chia	92.28±1.68bc	83.71±2.14cd
Quinoa	91.42±1.35bc	86.57±1.60c
Chi and Quinoa	88.71±1.83c	74.28±1.83d

Means in the same column with different letters are significantly different at P≤0.05

Results in table (3) revealed that feeding of rats with celiac disease significantly increased TC and TG in the positive group as compared to the normal control group. Rats fed Chia, Quinoa and their combination had a significant decrease in TC and TG.

These results agreed to those reported by **Eugenia et al. (2014)**, **Nowak et al. (2016)** and **Aktaş et al. (2018)**. The previous authors reported that Chia, Quinoa lowered TC and TG in rats with celiac disease.

**Table (4).** Effect of Chia and Quinoa-fortified gluten-free biscuit products on lipoprotein fraction (high density lipoprotein (HDL-c), low density lipoprotein (LDL-c) and very low density lipoprotein (VLDL-c). (n=7 rats)

Parameters Groups	HDL-c (mg/dL)	LDL-c (mg/dL)	VLDL-c (mg/dL)
Control (-ve)	50.57±1.23a	25.88±1.65b	19.68±0.42b
Control (+ve)	42.14±0.50c	55.74±1.77a	24.54±0.69a
Chia	46.57±1.49b	28.79±2.07b	16.47±0.62cd
Quinoa	46.58±0.93b	27.25±1.99b	17.31±0.32c
Chi and Quinoa	49.57±0.57ab	24.28±1.79b	14.85±0.16d

Means in the same column with different letters are significantly different at P≤0.05.

As shown in table (4), Celiac diseases in rats significantly decreased HDL-c, and increased LDL-c and VLDL-c. Rats fed Chia, Quinoa and their combination had a significant increase in HDL-c and a decrease in LDL-c and VLDL-c.

These findings were similar to those of **Norlaily et al. (2013)**; **Nowak et al. (2016)** and **Khatri et al. (2023)**.

Moreover, many studies have described chia and quinoa as protecting against a variety of diseases, particularly cancer, allergies and, inflammatory diseases, and they may reduce the risk of cardiovascular diseases (**Sadeek et al., (2021)**).

**Table (5).** Effect of Chia and Quinoa-fortified gluten-free biscuit products on liver enzymes aspartateaminotransferase (AST); alanine aminotransferase (ALT) and alkaline phosphates (ALP) in rats.(n=7 rats)

Parameters Groups	AST (U/L)	ALT (U/L)	ALP (U/L)
Control (-ve)	14.71±0.42c	28.28±0.86d	115.00±1.38d
Control (+ve)	39.57±0.57a	32.57±0.64a	126.71±1.20a
Chia	19.71±0.42b	29.28±0.80bc	118.42±1.48b
Quinoa	19.57±0.42b	26.57±0.97b	117.58±1.12c
Chi and Quinoa	16.14±0.63c	25.00±0.81cd	116.57±1.84c

Means in the same column with different letters are significantly different at P≤0.05.

Data in table (5) show that Celiac diseases in rats significantly increased liver enzymes AST, ALT and ALP. Rats fed Chia, Quinoa and their combination had a significant decrease in the levels of elevated liver enzymes.

The present results agreed with those of **Rashtak et al. (2012) Muñoz et al. (2013); Khatri, et al., (2023)**. The previous author concluded that feeding Chia and Quinoa to rats lowered the level of liver enzymes.

**Table (6).** Effect of Chia and Quinoa-fortified gluten-free biscuit products on kidney function parameters uric acid (UA) and creatinine (Creat.) in rats. (n=7 rats)

Parameters Groups	UA (mg/dL)	Creat. (mg/Dl)
Control (-ve)	16.71±0.56c	0.78±0.036c
Control (+ve)	24.71±0.68a	1.79±0.058a
Chia	19.14±0.50bc	1.13±0.087b
Quinoa	19.42±0.64b	1.05±0.46b
Chi and Quinoa	17.14±0.63bc	0.82±0.043c

Means in the same column with different letters are significantly different at P≤0.05.

As shown in table (6), Celiac diseases in rats significantly increased kidney function parameters uric acid (UA) and creatinine (Creat.) in control positive groups. Rats fed Chia, Quinoa and their combination had a significant decrease in the levels

of elevated kidney parameters (uric acid and creatinine).

These results were in agreement with those of **Mohdet al. (2012) Muñoz et al.(2013) and Costantiniet al., (2014)**.

**Table (7).** Effect of Chia and Quinoa-fortified gluten-free biscuit products on glutathione (GSH); malondialhyde (MDA) and lactate dehydrogenase (LDH) in rats. (n=7 rats)

Parameters Groups	GSH (mg/dL)	MDA (mg/dL)	LDH (U/L)
Control (-ve)	5.09±0.023a	2.60±0.018c	118.28±3.44c
Control (+ve)	3.43±0.119d	3.67±0.103a	316.14±4.33a
Chia	4.80±0.020b	2.83±0.064b	144.57±2.71b
Quinoa	4.50±0.038c	2.73±0.073bc	139.00±2.41bc
Chi and Quinoa	4.74±0.034bc	2.28±0.040d	129.00±2.60bc

Means in the same column with different letters are significantly different at P≤0.05.

The current results revealed that Celiac diseases in rats significantly increased LDH and MDA but decreased GTH in the positive control group as compared to the negative control group. Rats fed Chia, Quinoa and their combination had a significant decrease in LDH and MDA and a significant increase in GTH.

These findings were nearly similar to those of **Muñoz et al., (2013) and Aktaş and Levent(2018)**. The previous authors reported thatfeeding of rats with Chia, Quinoa and their combination had a significant decrease in LDH and MDA and a significant increase in GTH.



### Histopathology Examination

Evaluation of microscopic lesion score of H&E stained sections of small intestine segments were demonstrated in

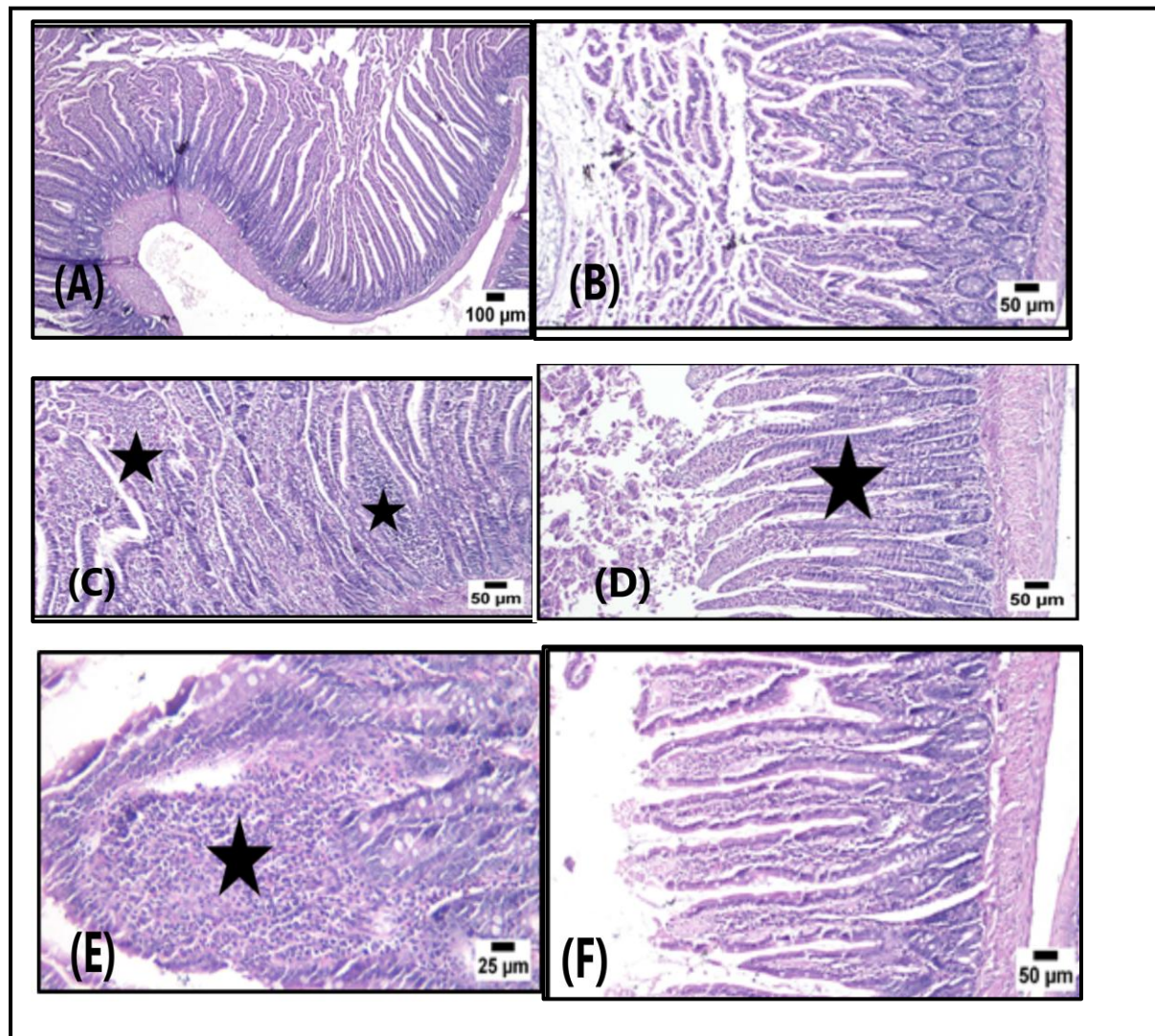


Fig (1).

**Fig (1):** Cross sections of small intestine of rats from:

(A) Normal control group showing normal histological structure all layers (H&E), (B) Positive control group showing marked atrophied intestinal villi with hyperplasia of crypts (H&E), (C) Positive control group showing expansion of the lamina propria with numerous infiltrated inflammatory cells (stars) (H&E), (D) Group fed Chia-fortified gluten free biscuit product showing moderate atrophy of the intestinal villi associated with hyperplasia of the crypts (Star) (H&E), (E) Group fed Quinoa -fortified gluten free biscuit product showing variable number of inflammatory cells infiltration in the intestinal mucosa (Star) (H&E) and (F) Group fed Chia and Quinoa -fortified gluten free biscuit product showing apparently normal histological structure.

The histopathological alterations seen in small intestine in rats with CD rats, in this study, were parallel with the reported biochemical parameters. These histopathological alterations in the present study were similar to those demonstrated by Muñoz et al. (2013), Khatri et.al. (2023) who reported that Chia and Quinoa would be an effective curative for

celiac disease- induced histopathological changes in the small intestine.

#### REFERENCES:

A.O.A.C. (1995): official methods of analysis of the association of official analytical chemists.14Ed. Washington, D.C., Published by the association of official Analytical chemists.

- Albers, N.; Benderson, V. and Warnick, G. (1983):** Enzymatic determination of high density lipoprotein cholesterol, Selected Methods. Clin. Chem.; 10: 91-99.
- Aktaş, K. and Levent, H. (2018):** The effects of chia (*Salvia hispanica* L.) and quinoa flours on the quality of rice flour and starch based-cakes. GIDA; 43 (4): 644-654. Doi: 10.15237/gida.GD18042
- Assa, A. and Frenkel-Nir, Y.(2017):** Anthropometric measures and prevalence trends in adolescents with coeliac disease: a population based study, Arch. Dis. Childh.; 102, (2): 139–144.
- Armitage, G.Y. and Berry, W.G. (1987):** Methods 7<sup>th</sup> Ed. Ames., Iowa State University. Press.39-63.
- Bancroft, J.D. and Stevens, A. (1977):** Theory and Practice of Histological Techniques. Edinburgh, Churchill-Livingstone, New York, Pages 740. Doi.org/10.1111/j.1365-2559.1990.tb00755.
- Chapman D. G., Gastilla R. and Campbell J. A. (1959):** Evaluation of protein in foods: 1- A Method for the determination of protein efficiency ratio. Can. J. Biochem. Phys; 37:679- 686.
- Cohen, I.; Day, S. and Shaoul R (2019):** Gluten in celiac disease—more or less? Rambam Maimonides Medical Journal, volume 10, No. 1, article e0007.
- Costantini, L.; Lukšič, L.; Molinari, R.; Kreft, I., Bonafaccia, G.; Manzi, L. and Merendino, N. (2014):** Development of gluten-free bread using tartary buckwheat and chia flour rich in flavonoids and omega-3 fatty acids as ingredients. Food Chem.; 165: 232-240.
- Eugenia, S.; Hera, E., Preze, G.T. and gómez, M. (2014):** Effect of Chia (*Salvia hispanica* L) Addition on the Quality of Gluten-Free Bread. J. Food Quality; 37(5): 309-317. Doi:10.1111/jfq.12098
- Friedewald, W.; Leve, R. and Fredrickson, D. (1972):** Estimation of the concentration of low density lipoprotein cholesterol in plasma without use of the preparative ultracentrifuge. Clin Chem, 18: 499-502.
- Hissin, P. and Hilf, R. (1970):** A fluorometric method for determination of oxidized and reduced glutathione in tissues. Anal Biochem, 74(1): 214-226.
- Khatri, M.; Singh, A.; Singh, R.; and Kamble, D.B.(2023):**Optimization and evaluation of quinoa and chia based gluten free pasta formulation. Food and Humanity; 1:174-179. Doi.org/10.1016/j.foohum.2023.05.009
- Lorentz, K. and Berndt, W. (1967):**Enzymic determination of uric acid by a colorimetric method. Analyt. Biochem.;18 (1): 58-63. Doi.org/10.1016/0003-2697(67)90056-5.
- Mohd AN, Yeap SK, Ho WY (2012):**The promising future of Chia. *Salvia Hispanica* L J Biomed Biotechnol 171956. Doi:10.1155/2012/171956
- Muñoz, L.A.; Cobos, A.; Diaz, O. and Aguilera, J.M. (2013):**Chia seed (*Salvia hispanica*): An ancient grain and a new functional food. Food Rev. Int.; 29: 394-408.
- Nikoukar, L.R.; Nabavizadeh, F.; Mohamadi, S.M. and Moslehi, A. (2014):**Protective effect of ghrelin in a rat model of celiac disease. Acta Physiologica Hungarica; 101(4):438-447. Doi:10.1556/APhysiol.101.2014.4.5
- Norlaily, M.A.; Yeap, S.K.; Ho, W.Y.; Beh, B.K.; Tan, S.W. and Tan, S.G. (2012):** The promising future of chia, *Salvia hispanica* L. J. Biomed Biotechnol, PMID: 23251075- PMCID: PMC3518271 Doi: 10.1155/2012/171956.
- Nowak, V.; Du, J. and Charrondière, U.R. (2016):** Assessment of the nutritional composition of quinoa (*Chenopodium quinoa* Wild.). Food Chem.; 193: 47-54.
- Ohkawa, H.; Ohishi, N. and Yagi, K. (1979):** Assay for lipid peroxides in animal Tissues by thiobarbituric acid reaction (TBA). Analyt. Biochem. ; 95:351- 358. Doi:10.1016/0003-2697(79)90738-3
- Paivi, T.; AlAhmary, K.; Bahkali, S.; AlKhathaami, A.; Munira K. Al-Saqabi, S.; Al Ammar, A.; Jawed, M. and Alosaimi, S.M. (2020):** The Epidemiology of Celiac Disease in the General Population and High-Risk Groups in Arab Countries: A Systematic Review" Bio Med Research International Volume Article ID 6865917.
- Rashtak, S. and Murray, J.A.(2012):** Celiac disease, new approaches to therapy. Aliment. Pharmacol. Ther. ; 35: 768-781.
- Reeves, P.G.; Nielsen, F.H. and Fahmy, G.G. (1993):** AIN-93. Purified diets for laboratory rodents: Final report of the American Institute of Nutrition ad hoc writing committee on the reformulation of the AIN-76 A Rodent diet. J. Nutri.; 123: 139-151.
- Richmond, N. (1973):** Colorimetric determination of total cholesterol and high density lipoprotein cholesterol (HDL-c). Clin. Chem., 19: 1350-1356.
- Roy, S. (1970):** colorimetric method of serum alkaline phosphatase. Journal of Clinical Chemistry, 16: 431-432.
- Sadeek, R.A.; Areeg S. Aly, A.S. and Abd Elsabor, R.G. (2021):** Incorporation Quinoa and Chia Flour with Wheat Flour to Enhance the Nutritional Value and Improve the Sensory Properties of Pasta. J. College Specif. Educ. Education. Specif. Stud.; 5(1):1-33.
- Štěpánková, R.; Kofronová, O.; Tucková, L.; Kozáková, H., Cebra J.J. and Tlaskalová-Hogenová, H. (2003):** Experimentally induced gluten enteropathy and protective effect of epidermal growth factor in artificially fed neonatal rats. J Pediatr Gastroenterol Nutr. 36(1):96-104. Doi: 10.1097/00005176-200301000-00018.
- Stikić, R.; Glamoclija, D.; Demin, M. and Vuceljac-radović, B.(2012):** Agronomical and nutritional evaluation of quinoa seeds (*Chenopodium quinoa* Wild.) as an ingredient in bread formulations. J. Cer. Sci.; 55:132-138
- Vassault, A.; Azzedine, M.; Bailly, M.; Cam, G.; Dumont, G. and Ekindjian, O.,(1986):** Commission "Validation de techniques" (Commission on "Validation of Techniques"). Annales de Biologie Clinique, 44, 679-685.

**Wahlefeld, A. (1974):** Methods of Enzymatic Analysis.  
Academic Press, Chapter, 5: 1831-1835.

**Young, D. (2001):** Effect of disease on clinical lab  
Tests, 4<sup>th</sup>Ed. AACC press.