



“MOLECULAR PROFILING OF METALLO-BETA LACTAMASE GENE WITH SPECIAL REFERENCE TO BLAIMP-1 GENE IN IMIPENEM RESISTANT *PSEUDOMONAS AERUGINOSA* ISOLATES FROM PATIENTS OF CHRONIC SUPPURATIVE OTITIS MEDIA AT A TERTIARY CARE CENTRE IN ANDHRA PRADESH, INDIA”.

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ABSTRACT

Introduction: A chronic inflammation of the middle ear and mastoid cavity that lasts for longer than two weeks is referred to as chronic suppurative otitis media (CSOM). A frequent bacteria that causes CSOM is *Pseudomonas aeruginosa*. Carbapenems are among the most effective antibiotics used against *Pseudomonas* infections. Resistant to carbapenems is often associated with production of metallo-β lactamases strains. The detection of the strains that produce MBLs can be useful for ensuring that patients receive the best care possible in an effort to curb and spread resistance.

Aim and Objective : To study the molecular profiling of metallo-beta lactamase gene with special reference to blaIMP-1 gene in imipenem resistant *Pseudomonas aeruginosa* isolates from patients of chronic suppurative otitis media at a tertiary care centre.

Material and Methods: The present study was a cross sectional study carried out in the Department of Microbiology and ENT Department for a period of 1 year i.e between June 2022 to June 2023 at Maharajah Institute of Medical Sciences, Vizianagaram. A total of 245 patients clinically suspected cases for CSOM were studied. Swabs taken from discharging ears were sent for Gram's staining, culture and antibiotic sensitivity test was performed according to the CLSI guidelines 2022. The isolates were further tested for MBL by screening test, by Imipenem – EDTA combined disc test, and MBL E test (Imipenem). The DNA was extracted by using Qiagen DNA Extraction kit, which was further proceeded for the blaIMP-1 gene detection for *Pseudomonas aeruginosa* by the conventional PCR.

Results: In the present study the clinically diagnosed suspected cases of CSOM were 245, out of which 80 (32.65%) was found to be positive for CSOM infection . The ratio of Male 48 (60%) was found to be more as compared to the Female 32 (40%), with the maximum cases in the age group of 0-10 and the least in the age group above 41 years. In our study it was observed that the maximum number of cases was observed in the Gram negative bacilli isolates (80%) as compared to the Gram positive isolates (18.5%) with sides of the ear equally affected. *P. aeruginosa* (47.5%) was the most common isolate followed by *Klebsiella* spp with 16 (20%) and among gram positive isolate *Staphylococcus aureus* was found to be 15 %, there was only 1 case found for *candida albicans* (1.25%). The sensitivity observed for *P. aeruginosa* for Colistin was (97.3%), Piperacillin-tazobactam (73.6%), Amikacin (76.3%), and Cefipime (73.6%) were found to be the most effective Antibiotics. The resistance to ciprofloxacin was (55.26%), Levofloxacin (50%), Piperacillin(26.3%), Gentamicin (36.8%), Imipenem (36.8%), Tobramycin(28.9%) and Ceftazidime (28.9%). The molecular characterization of the blaIMP-1 gene was detected in 12 (31.5%) of the isolates of *Pseudomonas aeruginosa* which were screening test-positives for MBL by Imipenem – EDTA combined disc test, and MBL E test (Imipenem).

Conclusion: Knowing the etiological agents of CSOM and their antibiogram is crucial for an effective therapy and avoidance of both illness complications and antimicrobial resistance. Judicial use of broad spectrum antibiotics, like Imipenem, is the need of the hour.

Keywords: CSOM, Metallo-βlactamases, CLSI, Molecular characterization, blaIMP-1 gene , DNA , PCR

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INTRODUCTION

Chronic suppurative otitis media (CSOM) is the most common prevailing infection in developing countries especially in children of low socioeconomic group. It presents as painless discharge and is the most common cause of deafness in India [1,2]. Various studies have shown that both gram positive as well as gram negative organisms are responsible for CSOM [3]. Generally, microbiological culture of the ear discharge simplicates *Pseudomonas aeruginosa*, *Proteus* spp and *Staphylococcus* as the prevalent causative organism [4].

Pseudomonas aeruginosa is the most commonly identified organism in CSOM reported by various studies in India and abroad with incidence ranging from 21% -52.94% [5]. Among the organisms, pseudomonas infection is known to produce deep seated and progressive infection in middle ear and mastoid leading to various intracranial and extracranial complications. *Pseudomonas aeruginosa* is one of the most important hospital-acquired pathogens that causes miscellaneous opportunistic infections [6].

Among the beta lactams, carbapenems are considered as the potent drug of choice for serious treatment of gram-negative bacteria infections. The most effective antibiotics that can be used against *Pseudomonas aeruginosa* are β - lactam antibiotics in which imipenem as a carbapenem is considered as the most appropriate antibiotic to be used against the mentioned organisms [7].

The emergence of multidrug-resistant (MDR: was defined as acquired non-susceptibility to at least one agent in three or more antimicrobial categories) and extremely drug resistant (XDR: was defined as non-susceptibility to at least one agent in all but two or fewer antimicrobial categories) *P. aeruginosa* isolates has been considered as a major concern for the treatment of infections caused by these isolates [8].

Carbapenemases are a wide spectrum group of beta-lactamase which hydrolyzes carbapenems to other β -lactams including monobactams, penicillins, and cephalosporins. Although carbapenems are a commonly last resort treatment used for MDR *P. aeruginosa* infection, the emergence of carbapenem-resistant *P. aeruginosa* is becoming a main public health concern and is associated with high rates of mortality and morbidity among hospitalized patients [9,10].

The MBLs encoding genes such as *blaVIM* and *blaIMP* are one of the most clinically important

classes of beta-lactamases. Carbapenemases acquires resistance belongs to Ambler molecular classes A, B and D. Metallo- β -lactamases (MBL) enzymes are the most significant carbapenemases. Nowadays the emergence of antibiotic resistance strains is one of the challenges in treating patients, such as MBLs producing *Pseudomonas aeruginosa*. The VIM, IMP and SPM types are the most clinically significant carbapenemases which is encoded by *blaVIM*, *blaIMP*, and *blaSPM* genes [10].

Resistance to carbapenems can be related to producing carbapenemase enzymes such as serine carbapenemases and the MBLs encoding genes such as IMP, VIM, and NDM [11].

Therefore, the present study was undertaken to study the molecular profiling of metallo-beta lactamase gene with special reference to *blaIMP-1* gene in imipenem resistant *Pseudomonas aeruginosa* isolates from patients of chronic suppurative otitis media at a tertiary care centre in Andhra Pradesh, India.

MATERIAL AND METHODS

This was a cross sectional study carried out in the Department of Microbiology and ENT Department for a period of 1 year i.e between June 2022 to June 2023 at Maharajah Institute of Medical Sciences, Vizianagaram. The ethical clearance was duly obtained from the Institutional Ethical committee. A total of 245 patients clinically suspected cases for CSOM were studied. Swabs taken from discharging ears were sent for Gram's staining, culture and antibiotic sensitivity test was performed according to the CLSI guidelines 2022 [12]. The isolates were further tested for MBL by screening test, by Imipenem – EDTA combined disc test, and MBL E test (Imipenem). The participants in the study who gave their agreement were included and patients who were taking antibiotics at the time were excluded from the study.

Sample Collection and Processing

The sample was collected using Pus swab from the external auditory canal and introduced into Amies transport medium bottle and sent for laboratory analysis. The sample was processed to primary gram stain for pus cells and inoculated into Blood agar (Oxoid, UK), and MacConkey agar (Oxoid, UK) and incubated aerobically at 37 °C for 24–48 h.

Screening, isolation and identification of organisms :

Identification of the bacteria was based on Microscopy and colony characteristics (colony morphology, hemolysis on blood agar, changes in the physical appearance of the differential media). Gram positive isolates were tested for catalase and Coagulase tests while biochemical tests for gram negative isolated bacteria were tested for oxidase, Triple sugar Iron (TSI), Sulphur indole and motility (SIM), urease production and citrate utilization [13].

Antimicrobial susceptibility testing

Antibiotic susceptibility test of isolated bacteria was performed using modified Kirby Bauer disc diffusion method according to the Clinical and Laboratory Standard Institute (CLSI) guidelines [12]. A colony suspension with concentration equivalent to 0.5 McFarland solution was prepared for each identified isolate and inoculated into Mueller–Hinton-Agar (Oxoid, UK). The Antibiotic discs were placed onto the media and incubated at 37 °C for 24 h. Gram positive isolates were tested against Ampicillin (10 µg), Amoxicillin/clavulanate (20/10µg), Ceftriaxone (30 µg), Gentamycin (10 µg), Ciprofloxacin (5 µg), Trimethoprim/sulfamethoxazole(1.25/23.75µg), Chloramphenicol (30 µg), Amikacin(17 µg) and Cephalexin (18 µg), Cefoxitin (30µg). Gram negative organisms were tested sensitivity to amikacin (AMK, 30 µg), gentamicin (GM, 10 µg), tobramycin (TOB,10 µg), ceftazidime (CAZ, 30 µg), cefepime (CFP, 50 µg),piperacillin (PIP, 100 µg), PIP/tazobactam (PTZ, 100/10µg), imipenem (IMP, 10 µg), ciprofloxacin (CIP, 5 µg), and levofloxacin (LFX, 5 µg) by modified Kirby Bauer disc diffusion method using Mueller Hinton agar (MHA) medium. A suspension of the isolated

colonies of each test strain equivalent to a 0.5 McFarland’s standard was prepared in sterile normal saline. Briefly, a suspension of each strain was made so that the turbidity was equal to 0.5 McFarland standards and then plated as a lawn culture on to MHA. Antibiotic discs were placed and plates were incubated at 37°C for 18–24 h. Results were interpreted in accordance with CLSI guidelines [12]. *Escherichia coli* ATCC 25922, *Staphylococcus aureus* (American Type Culture Collection; ATCC 25923 and *P. aeruginosa* ATCC 27853 were used as control strains.

The Phenotypic confirmatory test

Imipenem(IMP)- EDTA Combined disc test:

The test organisms were inoculated by lawn culture technique on the plates of Muller-Hinton agar(MHA) as recommended by CLSI [12]. The 10 µg Imipenem Disk and 750 µg Imipenem EDTA Disk(Hi-media SD281) were placed on the plate. The inhibition zones of the imipenem and imipenem-EDTA disks were compared after 16 to 18 hours of incubation at 37°C. In the combined disc test, if the increase in inhibition zone with the imipenem and EDTA disc will be ≥ 7 mm than the imipenem disc alone, it is considered as MBL positive [14].

MBL E test: The E-test MBL Strip contains a double sided seven-dilution range of IP(Imipenem) (4 to 256 µg/ml) and Imipenem (1 to 64 µg/ml) in combination with a fixed concentration of EDTA is considered as the most sensitive method for MBL detection. The E-test was done according to manufacturer’s instructions. MIC ratio of IP/ IPI (Imipenem+EDTA) of >8 or >3 log dilutions indicates MBL production [14].

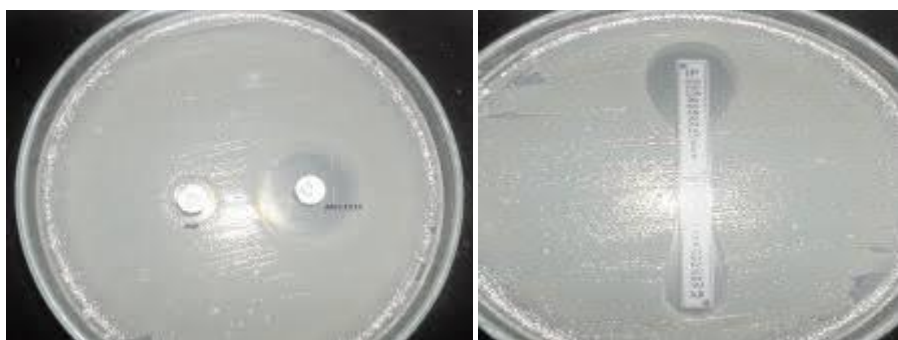


Figure No. 1: Shows MBL positive by Imipenem (IMP)- EDTA Combined disc test MBL positive by E-Test

Genotypic detection of blaIMP-1 gene in *P. aeruginosa*

The DNA was extracted from *P. aeruginosa* isolates using the Qiagen DNA Extraction Kit as per manufactures guidelines. The DNA was eluted

in 60 µl elution buffer and preserve at -20 °C till PCR analysis. For amplification of the target gene, PCR was carried out in a 50 µL reaction mixture with 30 no. of cycles. The primers were purchased

from “Saha gene” and was reconstituted with sterile double distilled water based on the manufacturer’s instruction.

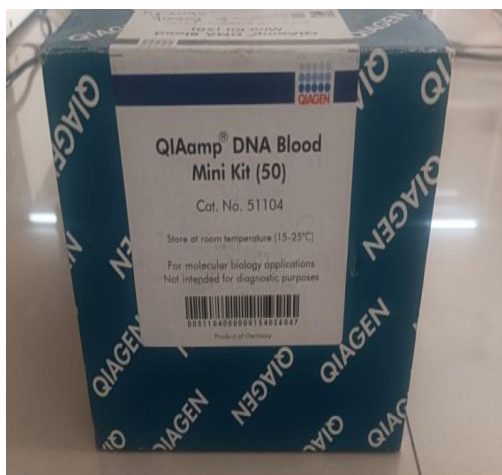


Figure No.2: The DNA Extraction kit



Figure No.3: The Reagents used for the DNA Extraction

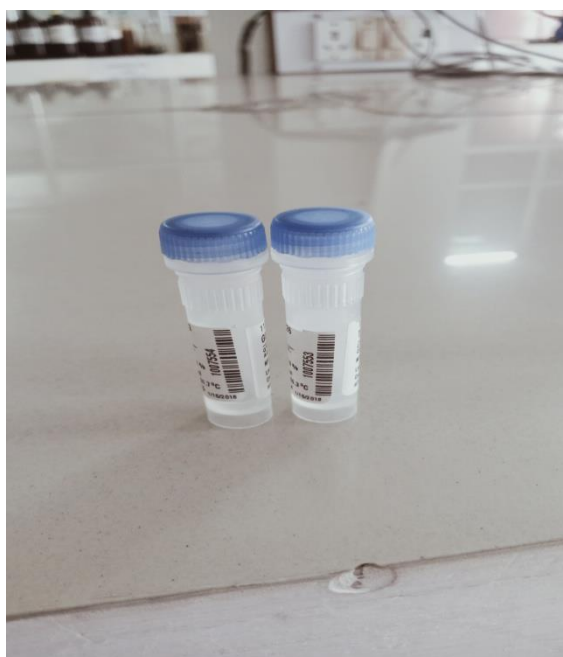


Figure No. 4: The *blaIMP-1* primers from the Saha gene

Molecular Characterization of *blaIMP-1* gene
Polymerase chain reaction (PCR) was carried out for detection of *blaIMP-1*, gene on a thermal cycler (Eppendorf, Germany). The primer pair sequences used in this study and the PCR conditions is described in the below Table 1. The DNA extraction was performed and the electrophoresis unit was run where 2% agarose gel was prepared with ethidium bromide. The

bromophenol blue dye was used for loading our DNA product which was then visualized in the gel documentation system. Positive controls used in this test were SPM-1 producing *P. aeruginosa* 16 strain (provided by Prof. Patrick Nordmann), *blaIMP1* from *Serratia marcescens* (sequenced by Bioneer company), and *blaVIM*. *P. aeruginosa* ATCC 27853 was used as a negative control [15].

Table No. 1: The Nucleotide sequences of primers used for detection of metallo-beta lactamase genes [15]

Primer name	Sequence	PCR Condition				
		Denaturing	Anneal	Extension	Cycles	Size(bp)
bla _{IMP-1}	5' TGAGCAAGTTATCTGTATTC 3' 5' TTAGTTGCTTGGTTTTGATG 3'	94°C, 60 s	57°C, 60 s	72°C, 2 min	35	740

RESULTS

In the present study the clinically diagnosed suspected cases of having CSOM were 245, out of which 80 (32.65%) was found to be positive for CSOM infection [Table No. 2]. The ratio of Male 48 (60%) were found to be more as compared to

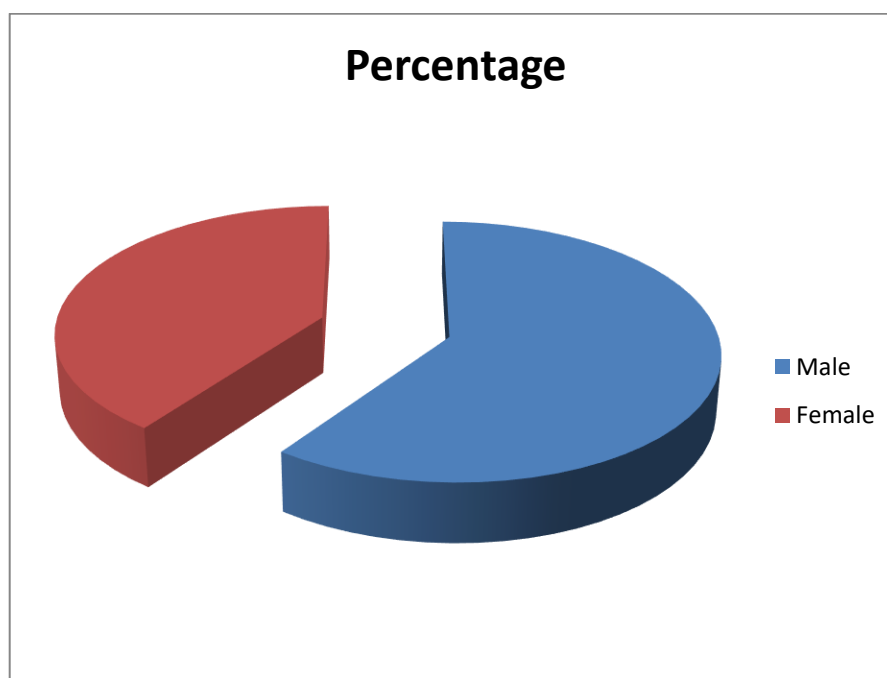
that of Female with 32 (40%) which is illustrated in the Table No.. 3. The maximum number of cases reported were observed in the age group of 0-10 years followed by 11-20 years of age and the least number of cases were seen in the age group above 41 years of age [Table No. 4].

Table No. 2: Total Number of Cases for chronic suppurative otitis media patients

S.N.	Type of Isolates	No. of Isolates	Percentage (%)
1.	Clinically diagnosed of having CSOM	245	67.34%
2.	culture positive for CSOM infection	80	32.65%

Table No. 3 : Genderwise Distribution of chronic suppurative otitis media patients

S.N.	Gender	No. of Isolates N=80	Percentage (%)
1.	Male	48	60%
2.	Female	32	40%



Graph No. 1 Genderwise Distribution of chronic suppurative otitis media patients

Table No. 4: Age wise distribution of the chronic suppurative otitis media patients

S.N.	Age group (Years)	Male N= 48	Female N=32	Percentage (%)
1.	0-10	18	10	35%
2.	11-20	11	8	23.75%
3.	21-30	6	7	16.25%
4.	31-40	5	3	10%
5.	41-50	2	2	5%
6.	51-60	3	1	5%
7.	61-70	2	1	3.75%
8.	≤ 80	1	-	1.25%

It was observed that the side of the ear affected was almost in equal distribution, with the left ear being 41(51.25%) and the right ear being 34 (42.5%) It was observed that 5 cases (6.25%) were bilateral Table No. 5. In our study it was observed that the maximum number of cases were found in

Gram negative bacilli isolates (80%) as compared to the Gram positive isolates (18.5%). In the current study it was recorded that 73 isolates (91.25%) samples showed growth of single isolates while 7 (8.75%) were mixed isolates.

Table No. 5: Bilateral distribution of chronic suppurative otitis media culture positive patients

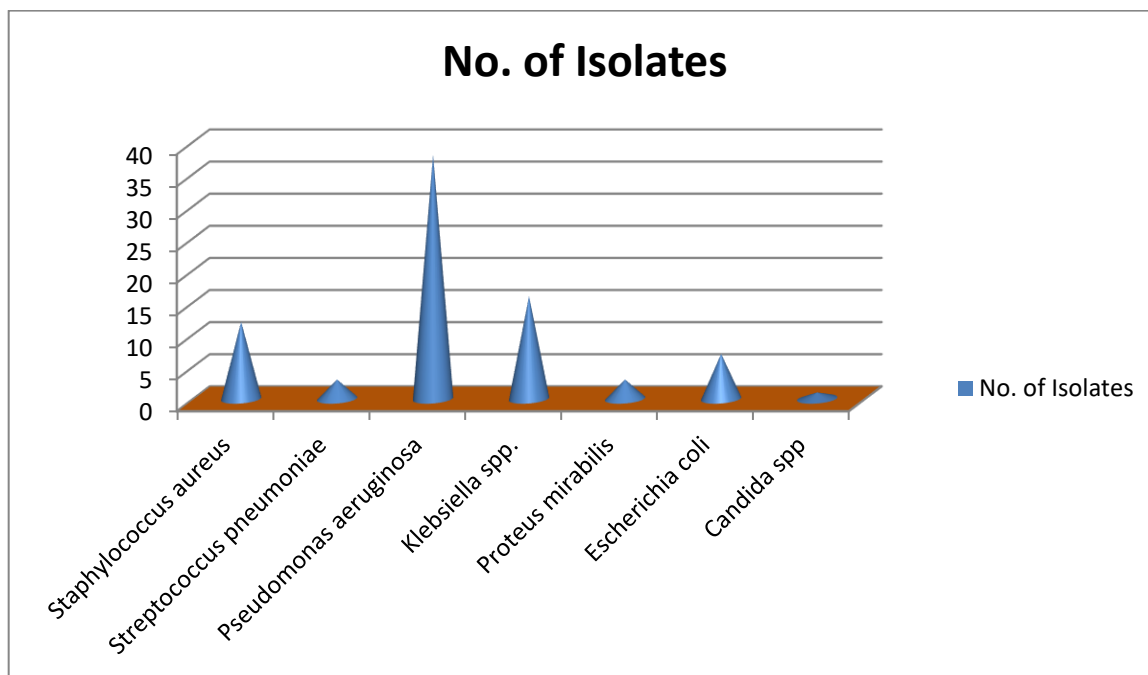
S.N.	Side of the Ear	No. of Isolates N= 80	Percentage (%)
1.	Left	41	51.25%
2.	Right	34	42.5%
3.	Bilateral	5	6.25%
4.	Total	80	100%

From the Table No. 6 it was clear that *P. aeruginosa* (47.5%) was the most common isolate followed by *Klebsiella* spp with 16 (20%) and

among gram positive isolates *Staphylococcus aureus* was found to be 15 %. There was only 1 case found for *candida albicans* (1.25%).

Table No. 6: Distribution of bacterial species associated with chronic suppurative otitis media patients

Bacterial Isolates	No. of Isolates N= 80	Percentage(%)
Gram positive bacteria		
<i>Staphylococcus aureus</i>	12	15%
<i>Streptococcus pneumoniae</i>	3	3.75%
Gram negative bacteria		
<i>Pseudomonas aeruginosa</i>	38	47.5%
<i>Klebsiella</i> spp.	16	20%
<i>Proteus mirabilis</i>	3	3.75%
<i>Escherichia coli</i>	7	8.75%
Fungal		
Candida spp	1	1.25%
No growth	165	



Graph No. 2: Phenotypic confirmatory test for MBL detection in Imipenem resistant *Pseudomonas aeruginosa* isolates from CSOM patients

Table No. 7: Phenotypic confirmatory test for MBL detection in Imipenem resistant *Pseudomonas aeruginosa* isolates from CSOM patients

Organisms	Imipenem(IMP)- Combined disc test:	EDTA	E-test
<i>Pseudomonas aeruginosa</i>	12/38		12/38

Pseudomonas aeruginosa isolates 12(31.5%) were screening test-positives for MBL by Imipenem –

EDTA combined disc test and MBL E test (Imipenem).

Table No. 8: Shows isolation rate of *Pseudomonas aeruginosa* strains susceptible and resistant to each antibiotic class (n=38)

Antibiotic class	Antibiotics	Percentage(%) Sensitivity	Percentage(%) Resistance
Polymyxins	Colistin	37 (97.36%)	1 (2.6%)
Aminoglycosides	Gentamycin	24 (63.15%)	14 (36.8%)
	Tobramycin	27 (71.05%)	11 (28.9%)
	Amikacin	29 (76.3%)	9 (23.68%)
Cephalosporins	Ceftazidime	27 (71.05%)	11 (28.9%)
	Cefipime	28 (73.6%)	10 (26.31%)
Antipseudomonal Penicillins	Piperacillin/ Tazobactam	28 (73.6%)	10 (26.31%)
Carbapenem	Imipenem	24 (63.15%)	14 (36.8%)
Fluoroquinolones	Ciprofloxacin	17 (44.73%)	21 (55.26%)
	Levofloxacin	19 (50%)	19 (50%)

In the current study the sensitivity observed in *P. aeruginosa* for Colistin was (97.3%), Piperacillin-tazobactam (73.6%), Amikacin (76.3%), and Cefipime (73.6%) were found to be the most effective antibiotics. The resistance to ciprofloxacin was (55.26%),

Levofloxacin (50%), Piperacillin(26.3%), Gentamicin (36.8%), Imipenem (36.8%), Tobramycin(28.9%) and Ceftazidime (28.9%). The blaIMP-1 gene was detected in 12 (31.5%) of the isolates of *Pseudomonas aeruginosa*. *S. aureus* showed a 100% sensitivity to Vancomycin,

Linezolid . *S..Pneumoniae* showed 100% sensitivity to gentamicin, netilmicin, levofloxacin, and ofloxacin. *P. mirabilis* showed a sensitivity of 100% to levofloxacin and 100% to ofloxacin, respectively, followed by ceftazidime , gentamicin 66.66% and ceftriaxone 66.66%.

E. coli showed a sensitivity of 85.71% and 85.71% to gentamicin and levofloxacin, respectively, followed by ofloxacin 85.71%, ceftazidime 71.42%, ceftriaxone 71.42%,

netilmicin 71.42%, and tetracycline 57.14%. *Klebsiella* species showed a sensitivity of 100% to levofloxacin and ofloxacin respectively, followed by netilmicin, ceftazidime, ceftriaxone with 93.71%, tetracycline and gentamicin with 75 %.

The DNA Extraction was performed by the Qiagen DNA kit and the DNA was isolated from the samples.



Figure No. 5: The DNA Extraction

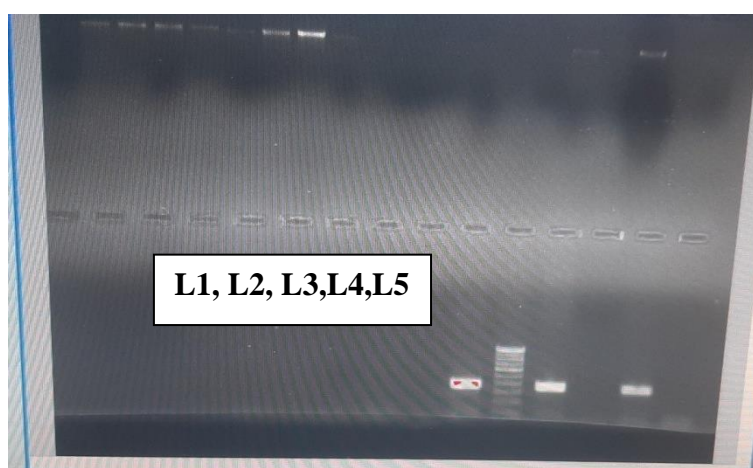


Figure No. 6: Photograph of amplified bla_{IMP-1} gene in *P. aeruginosa* ; the amplified DNA band size was obtained 740bp, , Lane 1 and L 3 is the sample positive for bla_{IMP}; L2 corresponding to 100bp ladder used ; L4 corresponds to the Negative control and L5 corresponds to the Positive control

DISCUSSION

In people with impaired immune systems, *Pseudomonas aeruginosa* is an opportunistic infection that can cause immunocompromised. It has been identified as the most prevalent bacterium in many hospital wards around the world [16]. Nosocomial infections caused by strains of this organism that produce MBL have become more common in recent years.

The carbapenem-resistant *P. aeruginosa* causes serious infections, such as nosocomial pneumonia which based on the reports is increasing in the hospitalized patients [17]. Resistance to carbapenems is often associated with production of metallo- β -lactamases . Nosocomial infections

caused by *Pseudomonas aeruginosa* remains the major cause of mortality, particularly because of emergence of multidrug-resistant strains [16, 17].

In the present study the clinically diagnosed suspected cases of having CSOM were 245, out of which 80 (32.65%) was found to be positive for CSOM infection . This study was similar to the study performed by the other author where the prevalence of csom observed was similar ranging between 21 to50 % [5, 18] but in contrast with the study by Deepthi Maringanti et al.,where, the ear discharge swabs were sent for culture and sensitivity in which only 106 patients out of 180, showed culture positives with the prevalence rate of 58% [19].

The ratio of Male 48 (60%) was found to be more as compared to that of Female with 32 (40%) in our study. This finding was similar to the study by Mohammed Jamiu Kazeem [20] where 198 (52.1%) patients were male while 182 (47.9%) were female. Other studies by Okesola and Fasina and Akingbade et al. [21,22], was also in support with our study but in contrast with the study by Shrestha et al.,[23] and Deepthi Maringanti et al. [19].

The maximum number of cases reported were observed in the age group of 0-10 years followed by 11-20 years of age and the least number of cases was seen in the age group above 41 years of age. This study was parallel to the study performed by the other research worker where the maximum number of cases recorded were in the age group of 11-20 years and least in the age group above 51 years [19]. Another study was also in support with the present study where maximum number of cases was reported in the age group of 10 years age with the fact that CSOM is predominantly a childhood disease, particularly the under 10 and least was observed above the age group of 50 years [20].

In the current study it was noted that the children and adolescents constitute the maximum patient population of CSOM. This may be because of the weak immune system in the young age and also because eustachian tubes are wider, shorter, and straighter compared to that of the adult.

CSOM has been described as disease more common among people of the poorer socio economic status, where there is overcrowding, more siblings under the age of five, poor sanitation and inadequate access to health care facilities; especially in children [24].

It was observed that the side of the ear affected was almost in equal distribution, with the left ear being 41(51.25%) and the right ear being 34 (42.5%) It was observed that 5 cases (6.25%) were bilateral. This study was in support with the study by Mohammed Jamiu Kazeem where the distribution pattern of the right and the left ear was equal while bilateral was 3.4% [20].

It was also observed that the maximum number of cases was found in Gram negative isolates as compared to the Gram positive isolates and only 1.25% with Fungal isolates. Similar observation was made by the other author where gram

negative isolates was observed to be the maximum. There was another study performed by Deepthi Maringanti et al. [19], which was in contrast to our study where there was no fungal growth recorded.

In our study it was observed that the maximum number of cases was found in Gram negative bacilli isolates (80%) as compared to the Gram positive isolates (18.5%). In the current study it was recorded that 73 isolates (91.25%) samples showed growth of single isolates while 7 (8.75%) were mixed isolates. This finding was in support with the other studies [20], [25].

P. aeruginosa (47.5%) was the most common isolate followed by *Klebsiella* spp with 16 (20%) and among gram positive isolates *Staphylococcus aureus* was found to be 15 %. There was only 1 case found for *candida albicans* (1.25%). This observation was compatible with the findings in other report where negative cultures were documented [26]. This study also correlates with the studies performed by other authors where the incidence of *P. aeruginosa* as the most commonly isolated organism in CSOM ranging from 21%-52.94% [18] Another study by Loy et al. (33.3%) [27] and Mansooret al. (40%) [28] also stated the rate of *P. aeruginosa* was more with no fungal isolate, but in Contrast with the study by Adoga et al. [29] where *Klebsiella* species (40%) as the predominant organism. *Streptococcus pneumonia* , *Proteus mirabilis* were noted to be 3.75% and *Escherichia coli* with 8.75%. This correlate with the study by the Nwankwo and Salisu [30] [31].

In case of Pseudomonas study, the sensitivity observed in *P. aeruginosa* for Colistin was (97.3%), Piperacillin-tazobactam (73.6%), Amikacin (76.3%) and Cefipime (73.6%) were found to be the most effective Antibiotics. This study was in support with the study performed by the other author where generally, ofloxacin (78.6%), gentamycin (76.9%), and ceftazidime (69.2%) were effective against *Pseudomonas* [32]. The resistance to ciprofloxacin was (55.26%), Levofloxacin (50%), Piperacillin(26.3%), Gentamicin (36.8%), Imipenem (36.8%), Tobramycin(28.9%) and Ceftazidime (28.9%). Similar finding was observed by the author co workers [19], [20] and [32].

MBLs are a group of β -lactamase enzymes which need one or two zinc in their active site to cleave the amide bond of the β -lactam ring to inactive β -lactam antibiotics [33]. In the present study blaIMP-1 gene was detected in all the 12 isolates, screened test-positives for MBL by Imipenem – EDTA combined disc test, and MBL E test

(Imipenem). In 2012, Fallah *et al.* checked 100 *P. aeruginosa* isolates from Shahid Motahari hospital in Teheran to detect *bla_{IMP}* and *bla_{VIM}* [34], where forty eight out of 83 (57.9%) imipenem-resistant *P. aeruginosa* showed MBL activity.

Since *Pseudomonas* was the predominant organism isolated in most CSOM cases and is mostly highly sensitive to ciprofloxacin which has none of the ototoxic risks of aminoglycosides and resistant to routinely used penicillin group of drugs and cephalosporins, it may be concluded that ciprofloxacin can be adopted as a first line antimicrobial treatment for CSOM culture positive cases. Piperacillin –tazobactam, imipenem and meropenem though highly sensitive, are considered as reserve drugs in CSOM cases which are not responding to ciprofloxacin and gentamycin [19].

Hence, regarding to horizontal transmission of integron-associated MBL genes, detecting MBL positive strains is necessary. Moreover, by using new methods for rapid identification of MBL positive bacteria in the patients, we could prevent spreading of metallo-beta lactamase strains to other patients.

As due to the advent of broad spectrum antibiotics, but inadvertent use of antibiotics both topical and systemic, will lead to emergence of multidrug resistant strains of bacteria [5] [6].

CONCLUSION

Similar to other chronic diseases, CSOM can affect a person's quality of life and capacity to find work. *Pseudomonas aeruginosa* being the most prevalent in the current study where Colistin, Piperacillin-tazobactam, Amikacin, and cefepime were found to be the most efficient antibiotics against this strain. In the mellimarla, vizianagaram region, the percentage of *P. aeruginosa* isolates resistant to imipenem due to MBL enzymes is rising (47.5%).

It is crucial to prevent the emergence and spread of resistant pathogens. Therefore, knowledge of the etiological agents of CSOM and their antibiogram data should be used when formulating antibiotic policy.

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